Package 'GOexpress'

November 12, 2026

Title Visualise microarray and RNAseq data using gene ontology annotations

Version 1.46.0

Date 2025-09-03

Description The package contains methods to visualise the expression profile of genes from a microarray or RNA-seq experiment, and offers a supervised clustering approach to identify GO terms containing genes with expression levels that best classify two or more predefined groups of samples. Annotations for the genes present in the expression dataset may be obtained from Ensembl through the biomaRt package, if not provided by the user. The default random forest framework is used to evaluate the capacity of each gene to cluster samples according to the factor of interest. Finally, GO terms are scored by averaging the rank (alternatively, score) of their respective gene sets to cluster the samples. P-values may be computed to assess the significance of GO term ranking. Visualisation function include gene expression profile, gene ontology-based heatmaps, and hierarchical clustering of experimental samples using gene expression data.

Depends R (>= 3.4), grid, stats, graphics, Biobase (>= 2.22.0)

Imports biomaRt (>= 2.18.0), stringr (>= 0.6.2), ggplot2 (>= 0.9.0), RColorBrewer (>= 1.0), gplots (>= 2.13.0), randomForest (>= 4.6), RCurl (>= 1.95)

Suggests BiocStyle

License GPL (>= 3)

biocViews Software, GeneExpression, Transcription, DifferentialExpression, GeneSetEnrichment, DataRepresentation, Clustering, TimeCourse, Microarray, Sequencing, RNASeq, Annotation, MultipleComparison, Pathways, GO, Visualization, ImmunoOncology

URL https://github.com/kevinrue/GOexpress

LazyData true

git_url https://git.bioconductor.org/packages/GOexpress

git_branch RELEASE_3_23

git_last_commit f69efc7

git_last_commit_date 2026-04-28

Repository Bioconductor 3.23

Date/Publication 2026-11-11

Author Kevin Rue-Albrecht [aut, cre],
Tharvesh M.L. Ali [ctb],
Paul A. McGettigan [ctb],
Belinda Hernandez [ctb],
David A. Magee [ctb],
Nicolas C. Nalpas [ctb],
Andrew Parnell [ctb],
Stephen V. Gordon [ths],
David E. MacHugh [ths],
Hugo Gruson [ctb]

Maintainer Kevin Rue-Albrecht <kevinrue67@gmail.com>

Contents

GOexpress-package . . . 3
AlvMac . . . 4
AlvMac_allgenes . . . 6
AlvMac_allGO . . . 7
AlvMac_GOgenes . . . 8
AlvMac_results . . . 9
AlvMac_results.pVal . . . 10
cluster_GO . . . 12
expression_plot . . . 13
expression_plot_symbol . . . 16
expression_profiles . . . 18
expression_profiles_symbol . . . 21
GO_analyse . . . 24
heatmap_GO . . . 28
hist_scores . . . 31
list_genes . . . 32
microarray2dataset . . . 33
plot_design . . . 34
prefix2dataset . . . 35
pValue_GO . . . 36
quantiles_scores . . . 37
rerank . . . 38
subEset . . . 39
subset_scores . . . 40
table_genes . . . 42

Index **44**

GOexpress-package *Visualise microarray and RNAseq data with gene ontology annotations.*

Description

Integrates gene expression data with gene ontology annotations to score and visualise genes and gene ontologies best clustering groups of experimental samples. Supports custom annotations, or alternatively provides an interface to the Ensembl annotations using the `biomaRt` package. The default scoring approach is based on the random forest framework, while a one-way ANOVA is available as an alteranative. GO term are scored and ranked according to the average rank (alternatively, average power) of all associated genes. P-values can be generated to assess the significance of GO term ranking. The ranked list of GO terms is returned, with tools allowing to visualise the statistics on a gene- and ontology-basis.

Details

Package:	GOexpress
Type:	Package
Version:	1.7.1
Date:	2016-02-06
License:	GPL (>= 3)

This package requires only two input variables

1. An `ExpressionSet` containing assayData and phenoData. The former should be a gene-by-sample matrix providing gene expression values for each gene in each sample. The latter should be an `AnnotatedDataFrame` from the `Biobase` package providing phenotypic information and grouping factors with two or more levels.
2. The name of the grouping factor to investigate, which must be a valid column name in the `phenoData` slot of the above `ExpressionSet`.

Following analysis, visualisation methods include:

- Histogram and quantiles representations of the scores of GO terms
- Permutation-based P-values to assess the significance of GO term ranking
- Filtering of results on various criteria (e.g. number of genes annotated to GO term)
- Re-ordering of GO terms and gene result tables based on score or rank metric
- Table of statistics for genes annotated to a given GO term
- Hierarchical clustering of samples based on the expression level of genes annotated to a given GO term
- Heatmap of samples and genes based on the expression level of genes annotated to a given GO term

- Expression profile of a gene against one given factor (e.g. Time) while grouping samples on another given factor (e.g. Treatment)
- Univariate analysis of the expression level of a gene in the different groups of each experimental factor.
- Venn diagram of the counts of genes shared between a list of GO terms.

Author(s)

Maintainer: Kevin Rue-Albrecht <kevinrue67@gmail.com>

See Also

Main method for an example usage: GO_analyse.

Packages Biobase, randomForest, RColorBrewer, VennDiagram.

Methods biomaRt:getBM, ggplot2:ggplot, gplots:heatmap.2, gplots:bluered, gplots:greenred, grid:grid.newpage, grid:grid.layout, stringr:str_extract.

Examples

```
# Sample input data available with package:
data(AlvMac)

# Sample output data available with package:
data(AlvMac_results)

# Supported species and microarrays:
data(microarray2dataset)
data(prefix2dataset)
```

AlvMac *Sample data from a RNAseq experiment.*

Description

An example ExpressionSet including expression data and phenotypic information about the samples.

The expression data is saved in the assayData slot of the ExpressionSet. It is a gene-by-sample matrix, containing a subset of data from an *in vitro* stimulation of bovine macrophages with different mycobacterial strains. Column names are sample names, and row names are Ensembl gene identifiers of the *Bos taurus* species. Each cell contains the log2-transformed normalised expression level of each gene in each sample.

The phenotypic information is saved in the phenoData slot of the ExpressionSet. Row names are sample names and columns contain descriptive information about each sample, including experimental factors(e.g. Treatment, Timepoint, Animal).

Usage

```
data(AlvMac)
```

Details

Gene expression was measured in poly-A purified strand-specific RNA libraries using the RNA-Sequencing Illumina(R) HiSeq(R) 2000 platform as paired-end 2 x 90 nucleotide reads. Raw reads from pooled RNA libraries were first deconvoluted according to sample-specific nucleotide barcodes. Read pairs containing adapter sequence in either read mate were discarded, and similarly read pairs of low overall quality in either mate were also discarded. Paired-end reads from each filtered individual library were aligned to the *Bos taurus* reference genome (*B. taurus* UMD3.1.71 genome release) using the STAR aligner software. For each library, raw counts for each gene based on sense strand data were obtained using the featureCounts software from the Subread package. The featureCounts parameters were set to unambiguously assign uniquely aligned paired-end reads in a stranded manner to the exons of genes within the *Bos taurus* reference genome annotation (*B. taurus* UMD3.1.71 genome annotation). The gene count outputs were further processed using the edgeR Bioconductor package.

The gene expression quantitation pipeline within the edgeR package was customised to: (1) filter out all bovine rRNA genes; (2) filter out genes displaying expression levels below the minimally-set threshold of one count per million [CPM] in at least ten individual libraries (number of biological replicates); (3) calculate normalisation factors for each library using the trimmed mean of M-values method; (4) log2-transform CPM values based on the normalised library size.

To generate this test data subset, we extracted 100 genes from the original dataset of 12,121 genes. All 7 genes associated with the GO term "GO:0034142" (i.e. "toll-like receptor 4 signaling pathway") present in the original full-size filtered-normalised dataset were kept, all 3 Ensembl gene identifiers annotated to the gene symbol 'RPL36A', and finally another random 90 random genes, making a total of 100 genes measured in 117 samples. Samples include all 10 biological replicates collected at four different time-points, see `data(targets)`. The TLR4 pathway was found in the full dataset as the top-ranking biological pathway discriminating the different mycobacterial infections (unpublished observations).

Value

`assayData` is a matrix of expression levels for 100 genes (rows) measured in 117 samples (columns).

- `rownames` are Ensembl gene identifiers of the *Bos taurus* species.
- `colnames` are samples identifiers.

`phenoData` is a data frame with 117 samples and 7 descriptive fields (e.g. experimental factors) in the columns listed below:

- `rownames` are unique identifiers. Here, sample names.
- `File` contains local filenames where the RNAseq counts were obtained from.
- `Sample` contains individual sample name.
- `Animal` contains the unique identifier of the animal corresponding to the biological replicate, stored as a factor.
- `Treatment` contains the infection status of the sample, stored as a factor (CN: Control, MB: *M. bovis*, TB: *M. tuberculosis*)

- Time contains the time of measurement in hours post-infection,stored as a factor.
- Group contains a combination of the Treatment and Time factors above, stored as a factor itself.
- Timepoint contains the time of measurement, stored as a numeric value. This field is useful to use on the X-axis of expression plots. See function expression_plot().

Source

Publication in review process.

Examples

```
# Load the data
data(AlvMac)

# Structure of the data
str(AlvMac)

# Dimensions (rows, columns) of the data
dim(AlvMac)

# Subset of first 5 features and 5 samples
AlvMac[1:5, 1:5]

# Phenotypic information
pData(AlvMac)

# Phenotypic information about factor "Group"
AlvMac$Group

# Conversion of a factor to a character vector
as.character(AlvMac$Group)

# Number of samples (rows) and annotations (columns)
dim(pData(AlvMac))
```

AlvMac_allgenes *Example of custom gene feature annotations.*

Description

An example data.frame providing the identifier, name and description corresponding to Ensembl gene identifiers present in the AlvMac example ExpressionSet.

Usage

```
data("AlvMac_allgenes")
```

Details

This data-frame includes only Ensembl gene identifiers present in the AlvMac example ExpressionSet.

See the help page of the GO_analyse function for an example usage.

Value

A data frame detailing information about the 100 gene features present in the AlvMac example ExpressionSet:

- gene_id are Ensembl gene identifiers.
- external_gene_name contains the corresponding gene name.
- description contains the corresponding description.

Source

These annotations were obtained from the Ensembl BioMart server using the biomaRt package to access the Ensembl release 75 http://feb2014.archive.ensembl.org.

Examples

```
# Load the data
data(AlvMac_allgenes)

# Structure of the data
str(AlvMac_allgenes)

# First few rows
head(AlvMac_allgenes)
```

AlvMac_allGO *Example of custom gene ontology annotations.*

Description

An example data.frame providing the identifier, name and namespace corresponding to gene ontology identifiers, compatible with the AlvMac example ExpressionSet.

Usage

```
data("AlvMac_allGO")
```

Details

This data-frame includes all gene ontologies present in the btaurus_gene_ensembl dataset of the Ensembl BioMart server, including those associated with no gene identifier in the AlvMac example ExpressionSet.

See the help page of the GO_analyse function for an example usage.

Value

A data frame detailing information about 13,302 gene ontologies:

- `go_id` are gene ontology identifiers.
- `name_1006` contains the corresponding gene ontology name.
- `namespace_1003` contains the corresponding gene ontology namespace (i.e. "biological_process", "molecular_function", or "cellular_component")

Source

These annotations were obtained from the Ensembl BioMart server using the `biomaRt` package to access the Ensembl release 75 http://feb2014.archive.ensembl.org.

Examples

```
# Load the data
data(AlvMac_allGO)

# Structure of the data
str(AlvMac_allGO)

# First few rows
head(AlvMac_allGO)
```

AlvMac_GOgenes *Example of custom mapping between gene ontology identifiers and gene features.*

Description

An example `data.frame` associating Ensembl gene identifiers to gene ontology identifiers, compatible with the `AlvMac` example `ExpressionSet`.

Usage

```
data("AlvMac_GOgenes")
```

Details

This data-frame includes all annotations between Ensembl gene identifiers and gene ontology identifiers present in the `btaurus_gene_ensembl` dataset of the Ensembl BioMart server, including gene identifiers absent from the `AlvMac` example `ExpressionSet`.

Importantly, gene identifiers present in these annotations but absent from the `ExpressionSet` will be given a score of 0 (minimum valid score; indicates no power to discriminates the predefined groups of sample) and a rank equal to the number of genes present in the entire dataset plus one (worst rank, while preserving discrete continuity of the ranking). This is helpful where features considered uninformative were filtered out of the `ExpressionSet`.

See the help page of the `GO_analyse` function for an example usage.

Value

A data frame with 191,614 associations between the following 2 variables.

- gene_id are Ensembl gene identifiers of the *Bos taurus* species.
- go_id are gene ontology identifiers.

Source

These annotations were obtained from the Ensembl BioMart server using the biomaRt package to access the Ensembl release 75 http://feb2014.archive.ensembl.org.

Examples

```
# Load the data
data(AlvMac_GOgenes)

# Structure of the data
str(AlvMac_GOgenes)

# First few rows
head(AlvMac_GOgenes)
```

AlvMac_results *Sample output from the* GO_analyse() *function on an RNAseq experiment.*

Description

This variable may be used to test the filtering and visualisation methods implemented in the package. It contains the output of the command AlvMac_results = GO_analyse(eSet=AlvMac, f="Treatment") applied to the toy input data AlvMac.

Usage

```
data(AlvMac_results)
```

Value

A list of 9 slots summarising the input and results of the analysis:

- GO contains a table ranking all GO terms related to genes in the expression dataset based on the average ability of their related genes to cluster the samples according to the predefined grouping factor.
- mapping contains the table mapping genes present in the dataset to GO terms.
- genes contains a table ranking all genes present in the expression dataset based on their ability to cluster the samples according to the predefined grouping factor (see 'factor' below).

- `factor` contains the grouping factor analysed.
- `method` contains the statistical framework used.
- `subset` contains the filters used to select a subset of samples from the original `ExpressionSet` for analysis.
- `rank.by` contains the metric used to rank the scoring tables.
- `ntree` contains number of trees built during the randomForest analysis.
- `mtry` contains the number of features randomly sampled as candidates at each split in each tree built during the randomForest analysis.

Warning

Running the above command again, you might obtain slightly different scores and ranks due to the stochastic process of sampling used by the random forest algorithm. However, the ranking metric was found to be robust and stable across run, given adequate number of trees and predictor variables sampled.

To produce reproducible results, use the `set.seed()` function prior to running any randomising or sampling function.

Source

Source data are part of a publication in review.

Examples

```
data(AlvMac_results)
str(AlvMac_results)
head(AlvMac_results$GO, n=20)
head(AlvMac_results$GO$genes, n=20)
```

AlvMac_results.pVal — *Sample output from the* `pValue_GO()` *function on an RNAseq experiment.*

Description

This variable may be used to test the filtering and visualisation methods implemented in the package.
It contains the output of successively applying the commands `AlvMac_results = GO_analyse(eSet=AlvMac, f="Treatment")` and `AlvMac_results.pVal = pValue_GO(AlvMac_results, N=100,ranked.by=result$rank.by, rank.by='P')` to the toy input data `AlvMac`.

Usage

```
data("AlvMac_results.pVal")
```

Value

A list of 9 slots summarising the input and results of the analysis:

- `GO` contains a table ranking all GO terms related to genes in the expression dataset based on the average ability of their related genes to cluster the samples according to the predefined grouping factor.
- `mapping` contains the table mapping genes present in the dataset to GO terms.
- `genes` contains a table ranking all genes present in the expression dataset based on their ability to cluster the samples according to the predefined grouping factor (see 'factor' below).
- `factor` contains the grouping factor analysed.
- `method` contains the statistical framework used.
- `subset` contains the filters used to select a subset of samples from the original `ExpressionSet` for analysis.
- `rank.by` contains the metric used to rank the scoring tables.
- `ntree` contains number of trees built during the randomForest analysis.
- `mtry` contains the number of features randomly sampled as candidates at each split in each tree built during the randomForest analysis.
- `p.iterations` contains the number of permutations performed to compute the P-value in the `GO` slot.

Warning

Running the above command again, you might obtain slightly different scores and ranks due to the stochastic process of sampling used by the random forest algorithm. However, the ranking metric was found to be robust and stable across run, given adequate number of trees and predictor variables sampled.

To produce reproducible results, use the `set.seed()` function prior to running any randomising or sampling function.

Source

Source data are part of a publication in review.

Examples

```
data(AlvMac_results.pVal)
str(AlvMac_results.pVal)
head(AlvMac_results.pVal, n=20)
```

cluster_GO *Generates a hierarchical clustering of the samples*

Description

Clusters the samples using only the expression levels of genes associated with a given go_id.

Usage

```
cluster_GO(
    go_id, result, eSet, f=result$factor, subset=NULL,
    method_dist="euclidean", method_hclust="average", cex=0.8,
    main=paste(go_id, result$GO[result$GO$go_id == go_id, "name_1006"]),
    xlab="Distance", cex.main=1, main.Lsplit=NULL, ...)
```

Arguments

go_id	A Gene Ontology (GO) identifier.
result	The output of GO_analyse() or a subset of it obtained from subset_scores().
eSet	ExpressionSet of the Biobase package including a gene-by-sample expression matrix in the AssayData slot, and a phenotypic information data-frame in the phenodata slot. In the expression matrix, row names are Ensembl gene identifiers or probeset identifiers, and column names are sample identifiers. In the phentypic data-frame, row names are sample idenfifiers, column names are grouping factors and phenotypic traits usable for the one-way ANOVA.
f	The grouping factor in phenodata to label the samples by.
subset	A named list to subset eSet. Names must be column names existing in colnames(pData(eSet)). Values must be vectors of values existing in the corresponding column of pData(eSet).
method_dist	The method used to calculate distance between samples. See the dist() method from package stats.
method_hclust	The method used to cluster samples. See the hclust() method from package stats.
cex	A numeric value defining the character expansion of text in the plot.
main	A character string for the main title of the plot.
xlab	A label for the x axis, defaults to "Distance".
cex.main	Scaling factor of the main title font size. Default is 1. We suggest to use it in combination with the argument main.Lsplit for GO terms with long names.
main.Lsplit	Number of characters after which a new-line character will be inserted in the main title. If this would occur within a word, the new-line character will be inserted before this word. Default is NULL, leaving the title on a single line.
...	Additional parameters passed on to dist(), hclust() and plot().

Value

Returns the output of the `plot()` function.

Author(s)

Kevin Rue-Albrecht

See Also

Method `GO_analyse`.

Examples

```
# load the sample output data
data(AlvMac_results)

# Hierarchical clustering of samples based on the same GO term
cluster_GO(
    go_id="GO:0034142", result=AlvMac_results, eSet=AlvMac, cex=0.7
    )

# Re-label sample by another factor
cluster_GO(
    go_id="GO:0034142", result=AlvMac_results, eSet=AlvMac, cex=0.7, f="Group"
    )
```

---

expression_plot *Plots the expression profile of a gene by levels of a factor*

Description

This function will plot the expression profile of a gene across a valid X-axis variable in the `phenodata` while representing the mean and confidence interval of groups of samples defined by levels of another valid grouping factor in the `phenodata`.

Usage

```
expression_plot(
    gene_id, result, eSet, x_var, f=result$factor, subset=NULL,
    xlab=x_var, ylab="log2(cpm)", ylim=range(exprs(eSet)),
    col.palette="Accent",
    col=brewer.pal(n=length(levels(pData(eSet)[,f])), name=col.palette),
    level=0.95, title=NULL, title.size=2, axis.title.size=20,
    axis.text.size=15, axis.text.angle=0,
    legend.title.size=20, legend.text.size=15, legend.key.size=30)
```

Arguments

gene_id	An gene or probeset identifier present in rownames(expr_data).
result	An output of the GO_analyse() or subset_scores() function.
eSet	ExpressionSet of the Biobase package including a gene-by-sample expression matrix in the AssayData slot, and a phenotypic information data-frame in the phenodate slot. In the expression matrix, row names are Ensembl gene identifiers or probeset identifiers, and column names are sample identifiers. In the phentypic data-frame, row names are sample idenfifiers, column names are grouping factors and phenotypic traits usable for the one-way ANOVA.
x_var	A column name in phenodata to plot on the X-axis. If representing time on the X-axis, users should store the time-points as numeric values in the AnnotatedDataFrame to adequately space the time-points.
f	A column name in phenodata to group the samples when representing mean and confidence interval. The factor specified in the initial GO_analyse() call is used by default. Unexpected grouping factors of samples can reveal interesting trends (e.g. "Animal", "Tissue", "CellType", ...).
subset	A named list to subset eSet. Names must be column names existing in colnames(pData(eSet)). Values must be vectors of values existing in the corresponding column of pData(eSet).
xlab	Title of the X-axis. Default is tha value of x_var.
ylab	Title of the Y-axis. Default is "log2(cpm)".
ylim	Numeric vector of length 2 specifying the lower and upper bounds of the Y axis. Default is scaled to the full range of expression values in the expression dataset, to ease comparison of different genes. If set to NULL, the axis will be scaled to fit the plotted data only.
col.palette	A valid RColorBrewer palette name to fetch the colormap from. Default is palette "Accent".
col	A vector of color names or codes. The number of colors provided must match the number of levels of the grouping factor. If specified, overrides argument col.palette.
level	The confidence interval level to visualise around the mean of each group. Default is 0.95.
title	Changes the plot title. Default is a combination of the gene id and the associated gene.
title.size	Changes the font size of the title. Default is 2.
axis.title.size	Changes the font size of the axes title. Defalt is 20.
axis.text.size	Changes the font size of the axes text labels. Default is 15.
axis.text.angle	Changes the angle of the X axis text labels. Default is 0 (horizontal).
legend.title.size	Changes the font size of the legend title. Default is 20.

`legend.text.size`
: Changes the font size of the legend text labels. Default is 15.

`legend.key.size`
: Changes the size of the legend keys (in points). Default is 30.

Value

The ggplot object.

Warning

Common issues:

- It may not be possible to produce plots where the combination of X-axis variable and grouping factor leaves too few replicates to compute a confidence interval for each X value. This is a limitation imposed by the ggplot2 package to produce proper statistics and confidence intervals. In such cases, it may be preferrable to use the expression_profiles() method.

Author(s)

Kevin Rue-Albrecht

References

- ggplot2 package.

See Also

Package `Biobase`, methods `expression_plot_symbol`, `GO_analyse` and `ggplot`.

Examples

```
# load the sample output data
data(AlvMac_results)

# Expression by gene identifier (TNIP3)
expression_plot(
    gene_id="ENSBTAG00000047107",
    result=AlvMac_results, eSet=AlvMac, x_var="Timepoint"
    )

# Same gene, plotted by animal and grouped by treatment (merging time points)
expression_plot(
    gene_id="ENSBTAG00000047107",
    result=AlvMac_results, eSet=AlvMac, x_var="Animal",
    f="Treatment")

# Same gene, plotted by animal and grouped by time-point (merging treatments)
expression_plot(
    gene_id="ENSBTAG00000047107",
    result=AlvMac_results, eSet=AlvMac, x_var="Animal",
    f="Time")
```

expression_plot_symbol *Plots the expression profile of a gene by levels of a factor*

Description

This function will plot the expression profile of a gene across a valid X-axis variable from the AnnotatedDataFrame while representing the mean and confidence interval of groups of samples defined by levels of a valid grouping factor from the AnnotatedDataFrame.

In the case of a gene name represented by multiple gene or probeset identifiers in the dataset, a lattice of plots will be produced. Each of the plots in this lattice can subsequently be plotted separately using its associated index.

Usage

```
expression_plot_symbol(
    gene_symbol, result, eSet, x_var, f=result$factor, subset=NULL,
    index=0, xlab=x_var, ylab="log2(cpm)", ylim=range(exprs(eSet)),
    col.palette="Accent",
    col=brewer.pal(n=length(levels(pData(eSet)[,f])), name=col.palette),
    level=0.95, titles=c(), title.size=2, axis.title.size=20,
    axis.text.size=15, axis.text.angle=0,
    legend.title.size=20, legend.text.size=20, legend.key.size=30)
```

Arguments

gene_symbol	A gene name present in rownames(AlvMac_results$genes).
result	An output of the GO_analyse() or subset_scores() function.
eSet	ExpressionSet of the Biobase package including a gene-by-sample expression matrix in the AssayData slot, and a phenotypic information data-frame in the phenodate slot. In the expression matrix, row names are Ensembl gene identifiers or probeset identifiers, and column names are sample identifiers. In the phentypic data-frame, row names are sample idenfifiers, column names are grouping factors and phenotypic traits usable for the one-way ANOVA.
x_var	A column name in phenodata to plot on the X-axis. If representing time on the X-axis, users should store the time-points as numeric values in the AnnotatedDataFrame to adequately space the time-points.
f	A column name in phenodata to group the samples when representing mean and confidence interval. The factor specified in the initial GO_analyse() call is used by default. Unexpected grouping factors of samples can reveal interesting trends (e.g. "Animal", "Tissue", "CellType", ...).
subset	A named list to subset eSet. Names must be column names existing in colnames(pData(eSet)). Values must be vectors of values existing in the corresponding column of pData(eSet).

index	In the case where multiple gene or probeset identifiers are associated with the gene name, index=2 will plot the expression profile of the second feature identifier alone, for instance.
xlab	Title of the X-axis. Default is tha value of x_var.
ylab	Title of the Y-axis. Default is "log2(cpm)".
ylim	Numeric vector of length 2 specifying the lower and upper bounds of the Y axis. Default is scaled to the full range of expression values in the expression dataset, to ease comparison of different genes. If set to NULL, the axis will be scaled to fit the plotted data only.
col.palette	A valid RColorBrewer palette name to fetch the colormap from.
col	A vector of color names or codes. The number of colors provided must match the number of levels of the grouping factor. Default to a palette with an adequate set of colors. If specified, overrides argument col.palette.
level	The confidence interval level to visualise around the mean of each group. Default is 0.95.
titles	Character vector providing as many titles as there are plots to replace the default titles. Default is a combination of the gene id and the associated gene.
title.size	Changes the font size of the title. Default is 2.
axis.title.size	Changes the font size of the axes title. Defalt is 20.
axis.text.size	Changes the font size of the axes text labels. Default is 15.
axis.text.angle	Changes the angle of the X axis text labels. Default is 0 (horizontal).
legend.title.size	Changes the font size of the legend title. Default is 20.
legend.text.size	Changes the font size of the legend text labels. Default is 15.
legend.key.size	Changes the size of the legend keys (in points). Default is 30.

Value

The ggplot object, or the vector of feature identifiers if multiple features are annotated to the gene symbol.

Warning

Common issues:

- It may not be possible to produce plots where the combination of X-axis variable and grouping factor leaves too few replicates to compute a confidence interval for each X value. This is a limitation imposed by the ggplot2 package to produce proper statistics and confidence intervals. In such cases, it may be preferrable to use the expression_profiles_symbol() method.

Author(s)

Kevin Rue-Albrecht

References

- ggplot2 package.

See Also

Package Biobase, methods expression_plot, GO_analyse and ggplot.

Examples

```
# load the sample output data
data(AlvMac_results)

# Expression by gene identifier (TNIP3)
expression_plot_symbol(
    gene_symbol="PIK3AP1",
    result=AlvMac_results, eSet=AlvMac, x_var="Timepoint"
    )

# Same gene, plotted by animal and grouped by treatment (merging time points)
expression_plot_symbol(
    gene_symbol="PIK3AP1",
    result=AlvMac_results, eSet=AlvMac, x_var="Animal",
    f="Treatment"
    )

# Same gene, plotted by animal and grouped by time-point (merging treatments)
expression_plot_symbol(
    gene_symbol="PIK3AP1",
    result=AlvMac_results, eSet=AlvMac, x_var="Animal",
    f="Time")
```

---

expression_profiles *Plots the individual expression profile of a gene in samples series*

Description

This function will plot the expression profile of a gene in individual samples series across a valid X-axis variable in the phenodata, while colouring sample groups according to another variable in the phenodata, and using different line types according to yet another (or the same) variable in the phenodata.

Usage

```
expression_profiles(
    gene_id, result, eSet, x_var, seriesF, subset=NULL,
    colourF=result$factor, linetypeF=colourF, line.size=1.5,
    xlab=x_var, ylab="log2(cpm)", ylim=range(exprs(eSet)),
    col.palette="Accent",
```

```
    col=brewer.pal(n=length(levels(pData(eSet)[,colourF])),
                   name=col.palette),
    lty=1:length(levels(pData(eSet)[,linetypeF])),
    title=NULL, title.size=2, axis.title.size=20,
    axis.text.size=15, axis.text.angle=0,
    legend.title.size=20, legend.text.size=15,
    legend.key.size=30)
```

Arguments

gene_id	An gene or probeset identifier present in rownames(expr_data).
result	An output of the GO_analyse() or subset_scores() function.
eSet	ExpressionSet of the Biobase package including a gene-by-sample expression matrix in the AssayData slot, and a phenotypic information data-frame in the phenodate slot. In the expression matrix, row names are Ensembl gene identifiers or probeset identifiers, and column names are sample identifiers. In the phentypic data-frame, row names are sample idenfifiers, column names are grouping factors and phenotypic traits usable for the one-way ANOVA.
x_var	A column name in phenodata to plot on the X-axis. If representing time on the X-axis, users should store the time-points as numeric values in the AnnotatedDataFrame to adequately space the time-points.
seriesF	A column name in phenodata to track samples of an identical series across the X-axis. The combination of seriesF and x_var should uniquely identify each sample in eSet.
subset	A named list to subset eSet. Names must be column names existing in colnames(pData(eSet)). Values must be vectors of values existing in the corresponding column of pData(eSet).
colourF	A column name in phenodata to colour the corresponding groups of samples. The factor specified in the initial GO_analyse() call is used by default.
linetypeF	A column name in phenodata to assign a different line type to the corresponding groups of samples. Default mirrors colourF.
line.size	The width of the plotted lines. Default is 1.5.
xlab	Title of the X-axis. Default is tha value of x_var.
ylab	Title of the Y-axis. Default is "log2(cpm)".
ylim	Numeric vector of length 2 specifying the lower and upper bounds of the Y axis. Default is scaled to the full range of expression values in the expression dataset, to ease comparison of different genes. If set to NULL, the axis will be scaled to fit the plotted data only.
col.palette	A valid RColorBrewer palette name to fetch the colormap from. Default is palette "Accent".
col	A vector of color names or codes. The number of colors provided must match the number of levels of the factor colourF. If specified, overrides argument col.palette. Default colours are obtained from the col.palette.

`lty` A vector of numeric values corresponding to line types. The number of line types provided must match the number of levels of the factor `linetypeF`. Default line types are `1:length(levels(pData(eSet)[, linetypeF]))`.

`title` Changes the plot title. Default is a combination of the gene id and the associated gene.

`title.size` Changes the font size of the title. Default is 2.

`axis.title.size` Changes the font size of the axes title. Defalt is 20.

`axis.text.size` Changes the font size of the axes text labels. Default is 15.

`axis.text.angle` Changes the angle of the X axis text labels. Default is 0 (horizontal).

`legend.title.size` Changes the font size of the legend title. Default is 20.

`legend.text.size` Changes the font size of the legend text labels. Default is 15.

`legend.key.size` Changes the size of the legend keys (in points). Default is 30.

Details

In order to track and visualise individual sample series, each sample in the `ExpressionSet` should be associated with a unique combination of `seriesF` and `x_var`. This may require the generation of a new factor in the `phenodata`, combining all experimental factors except that plotted on the X-axis. See below for an example on the training dataset.

Value

The ggplot object.

Author(s)

Kevin Rue-Albrecht

References

- ggplot2 package.

See Also

Package `Biobase` , methods `expression_plot_symbol`, `expression_plot_symbol` and `ggplot`.

Examples

```
# load the sample output data
data(AlvMac_results)

AlvMac$Series <- paste(AlvMac$Animal, AlvMac$Treatment, sep="_")
```

```
expression_profiles(
    gene_id = "ENSBTAG00000047107", result = AlvMac_results,
    eSet=AlvMac, x_var = "Timepoint", seriesF="Series",
    legend.title.size=10, legend.text.size=10, legend.key.size=15)

expression_profiles(
    gene_id = "ENSBTAG00000047107", result = AlvMac_results,
    eSet=AlvMac, x_var = "Timepoint", seriesF="Series",
    linetypeF="Animal",
    legend.title.size=10, legend.text.size=10, legend.key.size=15)
```

`expression_profiles_symbol` *Plots the individual expression profile of a gene in samples series*

Description

This function will plot the expression profile of a gene in individual samples series across a valid X-axis variable in the `phenodata`, while colouring sample groups according to another variable in the `phenodata`, and using different line types according to yet another (or the same) variable in the `phenodata`.

Usage

```
expression_profiles_symbol(
    gene_symbol, result, eSet, x_var, seriesF, subset=NULL,
    colourF=result$factor, linetypeF=colourF, line.size=1.5,
    index=0, xlab=x_var, ylab="log2(cpm)", ylim=range(exprs(eSet)),
    col.palette="Accent",
    col=brewer.pal(n=length(levels(pData(eSet)[,colourF])),
                   name=col.palette),
    lty=1:length(levels(pData(eSet)[,linetypeF])),
    titles=c(), title.size=2, axis.title.size=20,
    axis.text.size=15, axis.text.angle=0,
    legend.title.size=20, legend.text.size=15,
    legend.key.size=30)
```

Arguments

`gene_symbol`	A gene name present in `rownames(AlvMac_results$genes)`.
`result`	An output of the `GO_analyse()` or `subset_scores()` function.
`eSet`	`ExpressionSet` of the `Biobase` package including a gene-by-sample expression matrix in the `AssayData` slot, and a phenotypic information data-frame in the `phenodate` slot. In the expression matrix, row names are Ensembl gene identifiers or probeset identifiers, and column names are sample identifiers. In the phentypic data-frame, row names are sample idenfifiers, column names are grouping factors and phenotypic traits usable for the one-way ANOVA.

x_var — A column name in phenodata to plot on the X-axis. If representing time on the X-axis, users should store the time-points as numeric values in the AnnotatedDataFrame to adequately space the time-points.

seriesF — A column name in phenodata to track samples of an identical series across the X-axis. The combination of seriesF and x_var should uniquely identify each sample in eSet.

subset — A named list to subset eSet. Names must be column names existing in colnames(pData(eSet)). Values must be vectors of values existing in the corresponding column of pData(eSet).

colourF — A column name in phenodata to colour the corresponding groups of samples. The factor specified in the initial GO_analyse() call is used by default.

linetypeF — A column name in phenodata to assign a different line type to the corresponding groups of samples. Default mirrors colourF.

line.size — The width of the plotted lines. Default is 1.5.

index — In the case where multiple gene or probeset identifiers are associated with the gene name, index=2 will plot the expression profile of the second feature identifier alone, for instance.

xlab — Title of the X-axis. Default is tha value of x_var.

ylab — Title of the Y-axis. Default is "log2(cpm)".

ylim — Numeric vector of length 2 specifying the lower and upper bounds of the Y axis. Default is scaled to the full range of expression values in the expression dataset, to ease comparison of different genes. If set to NULL, the axis will be scaled to fit the plotted data only.

col.palette — A valid RColorBrewer palette name to fetch the colormap from. Default is palette "Accent".

col — A vector of color names or codes. The number of colors provided must match the number of levels of the factor colourF. If specified, overrides argument col.palette. Default colours are obtained from the col.palette.

lty — A vector of numeric values corresponding to line types. The number of line types provided must match the number of levels of the factor linetypeF. Default line types are 1:length(levels(pData(eSet)[, linetypeF])).

titles — Character vector providing as many titles as there are plots to replace the default titles. Default is a combination of the gene id and the associated gene.

title.size — Changes the font size of the title. Default is 2.

axis.title.size — Changes the font size of the axes title. Defalt is 20.

axis.text.size — Changes the font size of the axes text labels. Default is 15.

axis.text.angle — Changes the angle of the X axis text labels. Default is 0 (horizontal).

legend.title.size — Changes the font size of the legend title. Default is 20.

legend.text.size — Changes the font size of the legend text labels. Default is 15.

legend.key.size — Changes the size of the legend keys (in points). Default is 30.

Details

In order to track and visualise individual sample series, each sample in the `ExpressionSet` should be associated with a unique combination of `seriesF` and `x_var`. This may require the generation of a new factor in the `phenodata`, combining all experimental factors except that plotted on the X-axis. See below for an example on the training dataset.

Value

The ggplot object, or the vector of feature identifiers if multiple features are annotated to the gene symbol.

Author(s)

Kevin Rue-Albrecht

References

- ggplot2 package.

See Also

Package `Biobase`, methods `expression_plot_symbol`, `expression_plot_symbol` and `ggplot`.

Examples

```
# load the sample output data
data(AlvMac_results)

AlvMac$Series <- paste(AlvMac$Animal, AlvMac$Treatment, sep="_")

expression_profiles_symbol(
    gene_symbol="TNIP3", result = AlvMac_results,
    eSet=AlvMac, seriesF="Series", x_var = "Timepoint", linetypeF="Animal",
    line.size=1.5, title.size=1.5, legend.title.size=10, legend.text.size=10,
    legend.key.size=30)

expression_profiles_symbol(
    gene_symbol="TNIP3", result = AlvMac_results,
    eSet=AlvMac, seriesF="Series", x_var = "Timepoint", linetypeF="Animal",
    line.size=1.5, lty=rep(1,10), title.size=1.5, legend.title.size=10,
    legend.text.size=10, legend.key.size=30)
```

GO_analyse *Identifies gene ontologies clustering samples according to predefined factor.*

Description

Combines gene expression data with Gene Ontology (GO) annotations to rank and visualise genes and GO terms enriched for genes best clustering predefined groups of samples based on gene expression levels.

This methods semi-automatically retrieves the latest information from Ensembl using the biomaRt package, except if custom GO annotations are provided. Custom GO annotations have two main benefits: firstly they allow the analysis of species not supported in the Ensembl BioMart server, and secondly they save time skipping calls to the Ensembl BioMart server for species that are supported. The latter also presents the possiblity of using an older release of the Ensembl annotations, for example.

Using default settings, a random forest analysis is performed to evaluate the ability of each gene to cluster samples according to a predefined grouping factor (one-way ANOVA available as an alteranative). Each GO term is scored and ranked according to the average rank (alternatively, average power) of all associated genes to cluster the samples according to the factor. The ranked list of GO terms is returned, with tools allowing to visualise the statistics on a gene- and ontology-basis.

Usage

```
GO_analyse(
    eSet, f, subset=NULL, biomart_name = "ENSEMBL_MART_ENSEMBL",
    biomart_dataset="", microarray="",
    method="randomForest", rank.by="rank", do.trace=100, ntree=1000,
    mtry=ceiling(2*sqrt(nrow(eSet))), GO_genes=NULL, all_GO=NULL,
    all_genes=NULL, FUN.GO=mean, ...)
```

Arguments

eSet	ExpressionSet of the Biobase package including a gene-by-sample expression matrix in the AssayData slot, and a phenotypic information data-frame in the phenodate slot. In the expression matrix, row names are identifiers of expressed features, and column names are identifiers of the individual samples. In the phenotypic data-frame, row names are sample idenfifiers, column names are grouping factors and phenotypic traits usable for statistical tests and visualisation methods.
f	A column name in phenodata used as the grouping factor for the analysis.
subset	A named list to subset eSet for the analysis. Names must be column names existing in colnames(pData(eSet)). Values must be vectors of values existing in the corresponding column of pData(eSet). The original ExpressionSet will be left unchanged.

`biomart_name` The Ensembl BioMart database to which connect to.

`biomart_dataset`
The Ensembl BioMart dataset identifier corresponding to the species studied. If not specified and no custom annotations were provided, the method will attempt to automatically identify the adequate dataset from the first feature identifier in the dataset. Use data(prefix2dataset) to access a table listing valid choices.

`microarray` The identifier in the Ensembl BioMart corresponding to the microarray platform used. If not specified and no custom annotations were provided, the method will attempt to automatically identify the platform used from the first feature identifier in the dataset. Use `data(microarray2dataset)` to access a table listing valid choices.

`method` The statistical framework to score genes and gene ontologies. Either "randomForest" or "rf" to use the random forest algorithm, or alternatively either of "anova" or "a" to use the one-way ANOVA model. Default is "randomForest".

`rank.by` Either of "rank" or "score" to chose the metric used to order the gene and GO term result tables. Default to 'rank'.

`do.trace` Only used if method="randomForest". If set to TRUE, gives a more verbose output as randomForest is run. If set to some integer, then running output is printed for every do.trace trees. Default is 100.

`ntree` Only used if method="randomForest". Number of trees to grow. This should be set to a number large enough to ensure that every input row gets predicted at least a few times

`mtry` Only used if method="randomForest". Number of features randomly sampled as candidates at each split. Default value is 2*sqrt(gene_count) which is approximately 220 genes for a dataset of 12,000 genes.

`GO_genes` Custom annotations associating features present in the expression dataset to gene ontology identifiers. This must be provided as a data-frame of two columns, named `gene_id` and `go_id`. If provided, no call to the Ensembl BioMart server will be done, and arguments `all_GO` and `all_genes` should be provided as well, to enable all downstream features of `GOexpress`. An example is provided in `AlvMac_GOgenes`.

`all_GO` Custom annotations used to annotate each GO identifier present in `GO_genes` with the ontology name (e.g. "apoptotic process") and namespace (i.e. "biological_process", "molecular_function", or "cellular_component"). This must be provided as a data-frame containing at least one column named `go_id`, and preferably two more columns named `name_1006` and `namespace_1003` for consistency with the Ensembl BioMart. Supported alternative column headers are `name` and `namespace`. Respectively, `name` should be used to provide the description of the GO term, and `namespace` should contain one of "biological_process", "molecular_function" and "cellular_component". `name` is used to generate the title of ontology-based figured, and `namespace` is important to enable subsequent filtering of results by their corresponding value. An example is provided in `data(AlvMac_allGO)`.

`all_genes` Custom annotations used to annotate each feature identifier in the expression dataset with the gene name or symbol (e.g. "TNF"), and an optional description. This must be provided as a data-frame containing at least a column named

gene_id and preferably two more columns named external_gene_name and description for consistency with the Ensembl BioMart. A supported alternative header is name. While external_gene_name is important to enable subsequent visualisation of results by gene symbol, description is only displayed for readability of result tables. An example is provided in data(AlvMac_allgenes).

FUN.GO — Function to summarise the score and rank of all feature associated with each gene ontology. Default is mean function. If using "lambda-like" (anonymous) functions, these must take a list of numeric values as an input, and return a single numeric value as an output.

... — Additional arguments passed on to the randomForest and useMart methods, if applicable.

Details

The default scoring functions strongly favor GO terms associated with fewer genes at the top of the ranking. This bias may actually be seen as a valuable feature which enables the user to browse through GO terms of increasing "granularity", i.e. associated with increasingly large sets of genes, although consequently being increasingly vague and blurry (e.g. "protein binding" molecular function associated with over 6,000 genes).

It is suggested to use the subset_scores() function to subsequently filter out GO terms with fewer than 5+ genes associated with it. Indeed, those GO terms are more sensitive to outlier genes as they were scored on the average of a handful of genes.

Additionally, the pValue_GO function may be used to generate a permutation-based P-value indicating the chance of seeing each GO term reaching an equal or higher rank – or score – by chance.

Value

A list containing the results of the analysis. Some elements are specific to the output of each analysis method.

Core elements:

GO — A table ranking all GO terms related to genes in the expression dataset based on the average ability of their related genes to cluster the samples according to the predefined grouping factor.

mapping — The table mapping genes present in the dataset to GO terms.

genes — A table ranking all genes present in the expression dataset based on their ability to cluster the samples according to the predefined grouping factor.

factor — The predefined grouping factor.

method — The statistical framework used.

subset — The filters used to run the analysis only on a subet of the samples. NULL if no filter was applied.

rank.by — The metric used to rank order the genes and gene ontologies.

FUN.GO — The function used to summarise the score and rank of all gene features associated with each gene ontology.

Random Forest additional elements:

ntree	Number of trees grown.
mtry	Number of variables randomly sampled as candidates at each split.

One-way ANOVA does not have additional arguments.

Warning

Make sure that the factor f is an actual factor in the R language meaning. This is important for the underlying statistical framework to identify the groups of samples defined by their level of this factor.

If the column defining the factor (e.g. "Treatment") in phenodata is not an R factor, use pData(targets)$Treatment = factor(pData(targets)$Treatment) to convert the character values into an actual R factor with appropriate levels.

Author(s)

Kevin Rue-Albrecht

See Also

Methods subset_scores, pValue_GO, getBM, randomForest, and oneway.test.

Examples

```
# Load example data subset
data(AlvMac)
# Load a local copy of annotations obtained from the Ensembl Biomart server
data(AlvMac_GOgenes)
data(AlvMac_allgenes)
data(AlvMac_allGO)

# Run the analysis on factor "Treatment",
# considering only treatments "MB" and "TB" at time-point "48H"
# using a local copy of annotations obtained from the Ensembl BioMart server
AlvMac_results <- GO_analyse(
    eSet=AlvMac, f="Treatment",
    subset=list(Time=c("48H"), Treatment=c("MB", "TB")),
    GO_genes=AlvMac_GOgenes, all_genes=AlvMac_allgenes, all_GO=AlvMac_allGO
    )

# Valid Ensembl BioMart datasets are listed in the following variable
data(prefix2dataset)

# Valid microarray= values are listed in the following variable
data(microarray2dataset)

## Not run:
# Other valid but time-consuming examples:

# Run the analysis on factor "Treatment" including all samples
GO_analyse(eSet=AlvMac, f="Treatment")
```

```
# Run the analysis on factor "Treatment" using ANOVA method
GO_analyse(eSet=AlvMac, f="Treatment", method="anova")


# Use alternative GO scoring/summarisation functions (Default is: average)

# Named functions
GO_analyse(eSet=AlvMac, f="Treatment", FUN.GO = median)

# Anonymous functions (simple example without scientific value)
GO_analyse(eSet=AlvMac, f="Treatment", FUN.GO = function(x){median(x)/100})


# Syntax examples without actual data:

# To force the use of the Ensembl BioMart for the human species, use:
GO_analyse(eSet, f, biomart_dataset = "hsapiens_gene_ensembl")

# To force use of the bovine affy_bovine microarray annotations use:
GO_analyse(eSet, f, microarray = "affy_bovine")

## End(Not run)
```

---

heatmap_GO *Generates a heatmap and hierarchical clustering of the samples and the genes*

Description

Clusters the samples and the genes associated with a GO term using the expression levels of genes related to a given ontology. Represents expression levels of those genes in a heatmap.

Usage

```
heatmap_GO(
    go_id, result, eSet, f=result$factor, subset=NULL, gene_names=TRUE,
    NA.names=FALSE, margins=c(7 ,5),
    scale="none", cexCol=1.2, cexRow=0.5,
    labRow=NULL,
    cex.main=1, trace="none", expr.col=bluered(75),
    row.col.palette="Accent",
    row.col=c(),
    main=paste(
        go_id, result$GO[result$GO$go_id == go_id,"name_1006"]
        ),
    main.Lsplit=NULL,
    ...)
```

Arguments

go_id	A Gene Ontology (GO) identifier.
result	The output of GO_analyse() or a subset of it obtained from subset_scores().
eSet	ExpressionSet of the Biobase package including a gene-by-sample expression matrix in the assayData slot, and a phenotypic information data-frame in the phenodata slot. In the expression matrix, row names are Ensembl gene identifiers or probeset identifiers, and column names are sample identifiers. In the phentypic data-frame, row names are sample idenfifiers, column names are grouping factors and phenotypic traits usable for the one-way ANOVA.
f	The grouping factor in phenodata to label the samples by.
subset	A named list to subset eSet. Names must be column names existing in colnames(pData(eSet)). Values must be vectors of values existing in the corresponding column of pData(eSet).
gene_names	A boolean value. Default is TRUE, to label genes by their associated gene name. If FALSE, labels the genes by their feature identifier in the expression dataset (i.e. Ensembl gene identifier or microarray probeset).
NA.names	A boolean value. Default is TRUE. If labelling genes by their associated gene name (see argument gene_names), whether to display the gene feature identifier for gene features without associated gene name.
margins	A numeric vector of length 2 specifying the number of lines of margins to apply for the bottom and right margins, respectively. Defaults are 7 (bottom) and 5 (right). For Ensembl gene identifiers, we suggest setting manually a bottom margin of 13. See heatmap.2().
scale	Character indicating if the values should be centered and scaled in either the row direction or the column direction, or none. Default is "none". See heatmap.2().
cexCol, cexRow	Positive numbers, used as cex.axis in for the row or column axis labeling. Defaults are 1.2 and 1, respectively. See heatmap.2().
labRow	A character vector of names to re-label the rows (samples). If vector of length 1, it is assumed to be a column name in the phenoData slot. Default are the values of the factor f. See heatmap.2().
cex.main	Scaling factor of the main title font size. Default is 1. We suggest to use it in combination with the argument main.Lsplit for GO terms with long names.
trace	Character string indicating whether a solid "trace" line should be drawn across each 'row' or down each 'column', both' or 'none'. The distance of the line from the center of each color-cell is proportional to the size of the measurement. Defaults to 'none'.
expr.col	Character vector indicating the colors to represent the different levels of gene expression. Defaults to a colormap of 75 shades ranging from blue (low) to red (high) and centered around white. If using differential expression data, you should probably use greenred(75) instead.
row.col.palette	A valid RColorBrewer palette name to fetch the colormap from, to color-code the groups of samples.

row.col	A vector of color names or codes. The number of colors provided must match the number of levels of the grouping factor. Default to an palette of up to 9 colors marking the different levels of the predefined grouping factor on the left side of the heatmap.
main	Main title of the figure. Default is paste(go_id, go_name).
main.Lsplit	Number of characters after which a new-line character will be inserted in the main title. If this would occur within a word, the new-line character will be inserted before this word. Default is NULL, leaving the title on a single line.
...	Additional arguments passed on to heatmap.2().

Value

Returns the output of the heatmap.2() function.

Author(s)

Kevin Rue-Albrecht

See Also

Method heatmap.2, GO_analyse, and brewer.pal.info.

Examples

```
# load the sample output data
data(AlvMac_results)

# Heatmap the top-ranked GO term (toll-like receptor 4 signaling pathway) as
# example
heatmap_GO(go_id="GO:0034142", result=AlvMac_results, eSet=AlvMac)

# Same with larger sample labels on the right hand side.
heatmap_GO(go_id="GO:0034142", result=AlvMac_results, eSet=AlvMac, cexRow=1)

# Change the color-coding to green-black-red gradient (more appropriate for
# differential expression values)
library(gplots)
heatmap_GO(
    go_id="GO:0034142", result=AlvMac_results, eSet=AlvMac,
    expr.col=greenred(75)
    )
```

hist_scores *Plots the distribution of scores following an GOexpress analysis.*

Description

Plots the an histogram representing the frequencies of scores in the output variable of the GO_analyse() function.

This function can also be used on the output of `subset_scores()` function as it returns a value formatted identically to the output of the `GO_analyse()` function.

Usage

```
hist_scores(result,
            main=paste("Distribution of average scores in",
                       deparse(substitute(result))),
            xlab="Average score", ...)
```

Arguments

`result`	The output of the `GO_analyse()` function.
`main`, `xlab`	These arguments to title have useful defaults here.
`...`	Additional arguments passed on to `hist()`.

Value

Returns the output of the `hist()` function.

Author(s)

Kevin Rue-Albrecht

See Also

Method `hist`, and `GO_analyse`.

Examples

```
# load the sample output data with p.values computed
data(AlvMac_results.pVal)

# Histogram of scores (labelled with counts)
hist_scores(result=AlvMac_results, breaks=20, labels=TRUE)

# filter for Biological Processes associated with 5+ genes and <=0.05 P-value
filtered_results <- subset_scores(
    result=AlvMac_results.pVal, total_count=5, p.val=0.05,
    namespace="BP")
```

```
# Histogram of scores (labelled with counts)
hist_scores(result=filtered_results, breaks=20, labels=TRUE)
```

`list_genes` *Returns the genes associated with a Gene Ontology*

Description

Given a Gene Ontology (GO) identifier represented in the dataset, returns a character vector listing the feature identifiers annotated to it.

Usage

```
list_genes(go_id, result, data.only=TRUE)
```

Arguments

`go_id`	A Gene Ontology (GO) identifier represented in the dataset.
`result`	The output of `GO_analyse()` or a subset of it obtained from `subset_scores()`.
`data.only`	Whether to return only the feature identifiers present in the given dataset or alternatively returns all feature identifiers associated with the GO term in the Ensembl BioMart. Default is TRUE.

Value

A character vector listing the feature identifiers of the genes associated with the GO term.

Author(s)

Kevin Rue-Albrecht

See Also

Method `GO_analyse`.

Examples

```
# load the sample output data
data(AlvMac_results)

# List of genes associated with the GO term "toll-like receptor 4 signaling
# pathway"
list_genes(result=AlvMac_results, go_id="GO:0034142")
```

`microarray2dataset` *Table mapping probeset identifier prefixes to datasets in the Ensembl BioMart.*

Description

The species corresponding to an probeset identifier can often be identified from the prefix of the identifier (e.g. Bt.457.1.S1_at corresponds to *Bos taurus* which is only associated with a single microarray platform in the Ensembl BioMart). This table maps some manually curated unique patterns to the corresponding species and microarray platform.

Usage

```
data(microarray2dataset)
```

Details

All Agilent microarray share the same prefix pattern, making it very difficult to differentiate. Many Affymetrix microarrays also share the same prefix "AFFX" for several probesets.

The `dataset` and `microarray` arguments of the `GO_analyse` method are the best way to specify which BioMart information should be used to annotate the features in your expression dataset.

Value

A data frame with 158 rows and the 6 columns. Rows are sorted alphabetically by species name and each refer to a unique combination of dataset and microarray identifier in the Ensembl BioMart. The columns are described below:

- `dataset` contains species-specific biomaRt dataset names.
- `microarray` contains the microarray identifier in the Ensembl BioMart dataset above.
- `sample` contains a sample probeset from this combination of Ensembl BioMart dataset and microarray.
- `species` contains an example probeset sampled from the corresponding Ensembl BioMart dataset and microarray.
- `pattern` contains the corresponding manually curated pattern representative of probesets for this microarray platform. This pattern may or may not be unique to the microarray. Therefore we encourage users to use the 'microarray' argument of the `GO_analyse` method to specify the Ensembl BioMart dataset to use.
- `unique` contains a boolean value stating whether the pattern was found unique to the microarray or not.

Source

The `microarray2dataset.build` method stored in the `toolkit.R` script was used to query the Ensembl BioMart server and build this table.

Examples

```
data(microarray2dataset)
microarray2dataset
```

`plot_design` *Plot Univariate Effects for genes associated with a Gene Ontology*

Description

Successively plots univariate effects of one or more `factors`, typically for a designed experiment as analyzed by `aov`().

Usage

```
plot_design(
    go_id, result, eSet, subset=NULL,
    factors=colnames(pData(eSet)), main="", main.Lsplit=NULL, ...)
```

Arguments

`go_id`	A Gene Ontology (GO) identifier represented by at least one gene in the dataset.
`result`	The output of `GO_analyse()` or a subset of it obtained from `subset_scores()`.
`eSet`	`ExpressionSet` of the `Biobase` package including a gene-by-sample expression matrix in the `AssayData` slot, and a phenotypic information data-frame in the `phenodata` slot. In the expression matrix, row names are Ensembl gene identifiers or probeset identifiers, and column names are sample identifiers. In the phentypic data-frame, row names are sample idenfifiers, column names are grouping factors and phenotypic traits usable for the one-way ANOVA.
`subset`	A named list to subset `eSet`. Names must be column names existing in colnames(pData(eSet)). Values must be vectors of values existing in the corresponding column of pData(eSet).
`factors`	A set of column names from `phenodata`. Each of these values will be represented on the X-axis to investigate its effect on the average expression of a given genes for each level of that factor.
`main`	Changes the main title of the plots.
`main.Lsplit`	Number of characters after which a new-line character will be inserted in the main title. If this would occur within a word, the new-line character will be inserted before this word. Default is NULL, leaving the title on a single line.
`...`	Additional arguments which will be passed on to the `plot.design`() function.

Value

The output of the `plot.design`() function.

Author(s)

Kevin Rue-Albrecht

See Also

Method plot.design.

Examples

```
# load the sample output data
data(AlvMac_results)

# Univariate plot
plot_design(go_id="GO:0034142", eSet=AlvMac, result=AlvMac_results)
```

---

prefix2dataset *Table mapping Ensembl gene identifier prefixes to BioMart datasets.*

Description

The species corresponding to an Ensembl gene identifier can typically be identified from the prefix of the identifier (e.g. ENSBTAG corresponds to *Bos taurus*). This table maps each known unique prefix to the corresponding species.

Usage

```
data(prefix2dataset)
```

Details

C. elegans, *D. melanogaster*, and *S. cerevisiae* have atypical identifier pattern and prefixes in their Ensembl gene identifiers. However, the automatically extracted prefix for *C. elegans* and *D. melanogaster* – respectively "WBgene" and "FBgn" – can be used as such to identify datasets from those species. On the oher hand, prefixes used for the *S. cerevisiae* include "YHR", "YAL", and many others. Consequently, expression data from *S. cerevisiae* species is identified without referring to the "prefix2dataset"" table; instead, such datasets are identified if the first gene identifier in the dataset starts with "Y".

Value

A data frame with 69 rows and 4 columns. Each row refers to one dataset in the Ensembl BioMart. The columns are described below:

- `dataset` contains the biomaRt dataset name.
- `species` contains the corresponding species name.
- `prefix` contains the corresponding unique prefix.
- `sample` contains a sample Ensembl gene identifier from this dataset.

Source

The `prefix2dataset.build` method stored in the `toolkit.R` script was used to query the Ensembl BioMart server and build this table.

Examples

```
data(prefix2dataset)
str(prefix2dataset)
prefix2dataset
```

`pValue_GO` *Compute p-values for ontologies by randomising gene labels.*

Description

Permutes the rank of genes processed by `GO_analyse()` to compute a P-value for each ontology

Usage

```
pValue_GO(
    result, N=1000, ranked.by=result$rank.by, rank.by='P',
    FUN.GO=result$FUN.GO)
```

Arguments

`result`	The output of `GO_analyse()` or a subset of it obtained from `subset_scores()`.
`N`	The number of permutation desired. Default to 1000.
`ranked.by`	Either of 'rank' or 'score'. The metric used to compare whether GO terms in the randomised data arelative to their original value. Default to the current ordering method of the `result` object.
`rank.by`	Either of 'rank', 'score', or 'p.val'. The metric used to order the GO terms after computing of the P-value. Default to 'p.val'.
`FUN.GO`	Function to summarise the score and rank of all feature associated with each gene ontology. Logically, default is the function used in the call to `GO_analyse`. If using "lambda-like" (anonymous) functions, these must take a list of numeric values as an input, and return a single numeric value as an output.

Details

This function is relatively lengthy. Its procedure can be divided in several steps: (1) assemble a copy of all genes used in the original scoring step, then for each of the N permutations, (2) permute the gene labels, (3) aggregate the rank – or score – of all genes belonging to each GO term, (4) decide whether each GO term is ranked – or scored –better or worst than originally, (5) return a p-value based on the number of permutations where each GO term was better ranked – or scored – than originally.

Value

A list formatted identically to the results of the analysis, with an added 'p.val' column in the GO slot, and re-ordered by the chosen metric.

Author(s)

Kevin Rue-Albrecht

See Also

Method GO_analyse.

Examples

```
## Not run:
# Time-consuming examples:
# Limited here to N=1 for time constraint
# Recommended N=1000 (or at least 100).

# Run the analysis on factor "Treatment" including all samples
AlvMac.Pvals <- pValue_GO(result, N=1, ranked.by=result$rank.by, rank.by='P')

## End(Not run)
```

---

quantiles_scores *Returns the quantiles of scores following an GOexpress analysis.*

Description

Returns a set of quantiles in indicating the scores reached by given proportions of the GO terms.

This function can also be used on the output of `subset_scores()` function as it returns a value formatted identically to the output of the `GO_analyse()` function.

Usage

```
quantiles_scores(result, probs=c(0.9, 0.95, 0.99, 0.999, 0.9999),
    quartiles=FALSE)
```

Arguments

`result`	The output of `GO_analyse()` or a subset of it obtained from `subset_scores()`.
`probs`	Numeric vector of probabilities with values in [0,1]. (Values up to 2e-14 outside that range are accepted and moved to the nearby endpoint.) See `quantile`
`quartiles`	A numeric vector of the percentiles for which the scores are desired.

Value

A named vector of percentiles and corresponding scores.

Author(s)

Kevin Rue-Albrecht

See Also

Method quantile.

Examples

```
# load the sample output data with p.values computed
data(AlvMac_results.pVal)

# filter for Biological Processes associated with 5+ genes and <=0.05 P-value
filtered_results <- subset_scores(
    result=AlvMac_results.pVal, total_count=5, p.val=0.05,
    namespace="BP")

# Quantiles of scores
quantiles_scores(result=filtered_results)
```

---

rerank *Reorder the result variable by alternative metrics.*

Description

Reorder the ranked tables of GO terms and genes either by increasing (average) rank or decreasing (average) score.

Usage

```
rerank(result, rank.by = 'rank')
```

Arguments

result	The output of GO_analyse() or a subset of it obtained from subset_scores().
rank.by	Either of 'rank', 'score' or 'p.val'; the metric to rank the GO terms and genes. Note that 'pval' is only applicable on the output of the pValue_GO() function. See details for breaking ties.

Details

Taking an example, to rank GO terms by P-value and break ties by average rank, rerank first by 'rank', and then rerank the resulting object by 'p.val'.

Value

A list formatted identically to the results of the analysis, but ordered by the chosen metric.

Note

The name reorder() was not used to avoid conflict with package stats.

Author(s)

Kevin Rue-Albrecht

See Also

Method GO_analyse.

Examples

```
# load the sample output data
data(AlvMac_results)

# Re-rank the GO terms and genes based on the actual score instead of the rank
reranked.byScore <- rerank(result=AlvMac_results, rank.by="score")

# load the sample output data with p.values computed
data(AlvMac_results.pVal)

# To rank by P-value, while breaking the ties by ave_rank,
# rank first by ave_rank
reranked.byRank <- rerank(result=AlvMac_results.pVal, rank.by="rank")
# rank the result by P-value
reranked.pVal_rank <- rerank(result=reranked.byRank, rank.by="p.val")
```

---

subEset *Subset an ExpressionSet to values of phenotypic data columns.*

Description

Given a list of column names and corresponding values present in the phenoData slot of an ExpressionSet object, this method returns the subset of the ExpressionSet restricted to samples associated with the given values in the given columns.

Usage

```
subEset(eSet, subset = list())
```

Arguments

eSet	ExpressionSet of the Biobase package including a gene-by-sample expression matrix in the AssayData slot, and a phenotypic information data-frame in the phenodate slot. In the expression matrix, row names are Ensembl gene identifiers or probeset identifiers, and column names are sample identifiers. In the phentypic data-frame, row names are sample idenfifiers, column names are grouping factors and phenotypic traits usable for the one-way ANOVA.

subset — A named list to subset eSet. Names must be column names existing in colnames(pData(eSet)). Values must be vectors of values existing in the corresponding column of pData(eSet).

Value

A subset of the given ExpressionSet restricted to samples associated with the given values in the given columns

Author(s)

Kevin Rue-Albrecht

See Also

Class ExpressionSet.

Examples

```
# Load example data set
data(AlvMac)

# Subset it to only samples of "CN" and "MB" treatments, and also only "2H",
# "6H", and "24H" time-points
sub.AlvMac <- subEset(
    eSet=AlvMac,
    subset=list(
        Treatment=c("CN","MB"),
        Time=c("2H","6H")
        )
    )
```

---

subset_scores *Returns a filtered list from GO_analyse results.*

Description

Given an output variable from a GO_analyse analysis and a set of valid filters and thresholds, returns an identically formatted list keeping only the rows of the score table passing all the filters.

Usage

```
subset_scores(result, ...)

# Suggested use:
# on the output of GO_analyse()
# subset_scores(result, total=5, namespace="BP")
# or after application of pValue_GO()
# subset_scores(result, total=5, namespace="BP", p.val=0.05)
```

Arguments

result	The output of the GO_analyse() function.
...	Additional pairs of filter and threshold values in the format "filter=threshold". Filters must be valid names from colnames(result$GO).

Details

It is highly recommended to filter out GO terms with very few genes (e.g. less than 5 genes), as the scoring function is biased for those GO terms (see UsersGuide).

Suggested filters:

total_count, total:	Filter keeping only GO terms associated with at least the given count of genes in the annotati
p.val, p, P:	Filter keeping only the GO terms with P-value lower or equal to the given cutoff (This filter is
namespace, namespace_1003:	Filter keeping only the GO terms of a given type. Valid values are "biological_process", "mol

Additional filters:

data_count, data:	Filter keeping only GO terms associated with at least the given count of genes, present in the Expression
ave_rank, rank:	Filter keeping only GO terms with an ave_rank value equal or lower than the given cutoff (average of the r
ave_score, score:	Filter keeping only GO terms with an ave_score value equal or higher than the given cutoff (average of the

Value

A list formatted identically to the result object provided, with an added or updated filters.GO slot stating the filters and cutoffs applied, and restricted to:

- the gene ontologies passing the given filters and cutoff values.
- the genes mapped to those remaining ontologies (i.e. all genes not associated with an ontology are discarded in the output object).
- gene-mapping annotations related to the remaining gene ontologies.

Note

It is possible to further filter a filter result object. Some warnings may appear if the new filter cutoff conflict with the ones previously applied (stored in the filters.GO slot of the filtered object). Example of conflicting filter values are:

- Filtering for 'biological_process' a result object that was previously filtered for 'molecular_function' (no more 'BP' terms are present in the first filtered object).
- filtering for a total_count of 5 or more a result object previously filtered for a total_count of 10 or more.

Author(s)

Kevin Rue-Albrecht

See Also

Method GO_analyse.

Examples

```
# load the sample output data
data(AlvMac_results.pVal)

# have an overview of the result variable
str(AlvMac_results.pVal)

# filter for Biological Processes associated with 5+ genes and <=0.05 P-value
filtered_results <- subset_scores(
    result=AlvMac_results.pVal, total_count=5, p.val=0.05,
    namespace="BP")

# have an overview of the filtered result variable
str(filtered_results)
```

---

table_genes *Returns a table listing the genes associated with a given Gene Ontology*

Description

Given a Gene Ontology (GO) identifier represented in the dataset and the output variable of a GO_analyse() function, table_genes() returns a table listing the genes associated with that go_id, their associated gene name, and description.

Usage

```
table_genes(go_id, result, data.only=FALSE, order.by='rank')
```

Arguments

go_id	A Gene Ontology (GO) identifier.
result	The output of the GO_analyse() function.
data.only	Whether to return only the feature identifiers present in the given dataset or alternatively returns all feature identifiers associated with the GO term in the Ensembl BioMart.
order.by	The metric or name to order the output table. Default is increasing 'rank', which corresponds to decreasing score. Valid values are 'rank' (increasing) equivalent to 'score' (decreasing), 'gene_id', 'name', equivalent to 'external_gene_name' and 'external_gene_id'. Text ranking in alphabetical order.

Value

A data frame listing the statistics and annotations for the genes present in the expression dataset and associated with the GO term.

Author(s)

Kevin Rue-Albrecht

See Also

Method GO_analyse.

Examples

```
# load the sample output data
data(AlvMac_results)

# Table of result for genes associated with the GO term
# "toll-like receptor 4 signaling pathway"
table_genes(result=AlvMac_results, go_id="GO:0034142")

# ordered by gene name
table_genes(result=AlvMac_results, go_id="GO:0034142", order.by='name')
```

Index

∗ **ExpressionSet**
subEset, 39
∗ **GOexpress**
AlvMac, 4
AlvMac_allgenes, 6
AlvMac_allGO, 7
AlvMac_GOgenes, 8
AlvMac_results, 9
AlvMac_results.pVal, 10
cluster_GO, 12
expression_plot, 13
expression_plot_symbol, 16
expression_profiles, 18
expression_profiles_symbol, 21
GO_analyse, 24
GOexpress-package, 3
heatmap_GO, 28
hist_scores, 31
list_genes, 32
microarray2dataset, 33
plot_design, 34
prefix2dataset, 35
pValue_GO, 36
quantiles_scores, 37
rerank, 38
subEset, 39
subset_scores, 40
table_genes, 42
∗ **anova**
GO_analyse, 24
GOexpress-package, 3
∗ **clustering**
cluster_GO, 12
GO_analyse, 24
GOexpress-package, 3
heatmap_GO, 28
plot_design, 34
∗ **datasets**
AlvMac, 4
AlvMac_allgenes, 6
AlvMac_allGO, 7
AlvMac_GOgenes, 8
AlvMac_results, 9
AlvMac_results.pVal, 10
microarray2dataset, 33
prefix2dataset, 35
∗ **expression**
cluster_GO, 12
expression_plot, 13
expression_plot_symbol, 16
expression_profiles, 18
expression_profiles_symbol, 21
GO_analyse, 24
GOexpress-package, 3
heatmap_GO, 28
plot_design, 34
∗ **gene**
cluster_GO, 12
expression_plot, 13
expression_plot_symbol, 16
expression_profiles, 18
expression_profiles_symbol, 21
GO_analyse, 24
GOexpress-package, 3
heatmap_GO, 28
list_genes, 32
plot_design, 34
table_genes, 42
∗ **ontology**
GO_analyse, 24
GOexpress-package, 3
heatmap_GO, 28
table_genes, 42
∗ **package**
GOexpress-package, 3
∗ **randomForest**
GO_analyse, 24
GOexpress-package, 3

AlvMac, 4
AlvMac_allgenes, 6
AlvMac_allGO, 7
AlvMac_GOgenes, 8
AlvMac_results, 9
AlvMac_results.pVal, 10
aov, *34*

Biobase, *4*, *15*, *18*, *20*, *23*
biomaRt:getBM, *4*
brewer.pal.info, *30*

cluster_GO, 12

expression_plot, 13, *18*
expression_plot_symbol, *15*, 16
expression_profiles, 18
expression_profiles_symbol, 21
ExpressionSet, *40*

factors, *34*

getBM, *27*
ggplot, *15*, *18*, *20*, *23*
ggplot2:ggplot, *4*
GO_analyse, *4*, *13*, *15*, *18*, 24, *30–32*, *37*, *39*, *42*, *43*
GOexpress *(*GOexpress-package*)*, 3
GOexpress-package, 3
gplots:bluered, *4*
gplots:greenred, *4*
gplots:heatmap.2, *4*
grid:grid.layout, *4*
grid:grid.newpage, *4*

heatmap.2, *30*
heatmap_GO, 28
hist, *31*
hist_scores, 31

list_genes, 32

microarray2dataset, 33

oneway.test, *27*

plot.design, *35*
plot_design, 34
prefix2dataset, 35
pValue_GO, *27*, 36

quantile, *38*
quantiles_scores, 37

randomForest, *4*, *27*
RColorBrewer, *4*
rerank, 38

stringr:str_extract, *4*
subEset, 39
subset_scores, *27*, 40

table_genes, 42

VennDiagram, *4*