Package 'GmicR'

November 12, 2026

Type Package

Title Combines WGCNA and xCell readouts with bayesian network learrning to generate a Gene-Module Immune-Cell network (GMIC)

Version 1.26.0

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Description This package uses bayesian network learning to detect relationships between Gene Modules detected by WGCNA and immune cell signatures defined by xCell. It is a hypothesis generating tool.

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Encoding UTF-8

LazyData true

RoxygenNote 7.0.2

Imports AnnotationDbi, ape, bnlearn, Category, DT, doParallel, foreach, gRbase, GSEABase, gRain, GOstats, org.Hs.eg.db, org.Mm.eg.db, shiny, WGCNA, data.table, grDevices, graphics, reshape2, stats, utils

Suggests knitr, rmarkdown, testthat

VignetteBuilder knitr

StagedInstall yes

biocViews Software, SystemsBiology, GraphAndNetwork, Network, NetworkInference, GUI, ImmunoOncology, GeneExpression, QualityControl, Bayesian, Clustering

git_url https://git.bioconductor.org/packages/GmicR

git_branch RELEASE_3_23

git_last_commit 7d3ca04

git_last_commit_date 2026-04-28

Repository Bioconductor 3.23

Date/Publication 2026-11-11

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`Auto_WGCNA` *Carries out WGCNA with default settings or custom settings*

Description

Carries out WGCNA with default settings or custom settings

Usage

```
Auto_WGCNA(
  datExpr,
  colname_correct = TRUE,
  minModuleSize = 10,
  deepSplit = 4,
  networkType = "signed hybrid",
  TOMType = "unsigned",
  corFnc = "bicor",
  mergeCutHeight = 0.25,
  sft_RsquaredCut = 0.85,
  removeFirst = FALSE,
  reassignThreshold = 1e-06,
  maxBlockSize = 25000,
  nThreads = NULL
)
```

Arguments

`datExpr`	Expression data. A matrix (preferred) or data frame in which columns are genes and rows ar samples. NAs are allowed, but not too many. See `checkMissingData` below and details.
`colname_correct`	a logical value. If TRUE (default), "." in gene names will be replaced with "-". This corrects a name change that is induced by R when creating a data.frame. If FALSE, no changes will be made.

`minModuleSize` minimum module size for module detection. See `cutreeDynamic` for more details.

`deepSplit` integer value between 0 and 4. Provides a simplified control over how sensitive module detection should be to module splitting, with 0 least and 4 most sensitive. See `cutreeDynamic` for more details.

`networkType` network type. Allowed values are (unique abbreviations of) `"unsigned"`, `"signed"`, `"signed hybrid"`. See `adjacency`.

`TOMType` one of `"none"`, `"unsigned"`, `"signed"`. If `"none"`, adjacency will be used for clustering. If `"unsigned"`, the standard TOM will be used (more generally, TOM function will receive the adjacency as input). If `"signed"`, TOM will keep track of the sign of correlations between neighbors.

`corFnc` the correlation function to be used in adjacency calculation.

`mergeCutHeight` dendrogram cut height for module merging.

`sft_RsquaredCut` desired minimum scale free topology fitting index R^2. Default is 0.80.

`removeFirst` should the first bin be removed from the connectivity histogram?

`reassignThreshold` p-value ratio threshold for reassigning genes between modules. See Details.

`maxBlockSize` integer giving maximum block size for module detection. Ignored if `blocks` above is non-NULL. Otherwise, if the number of genes in `datExpr` exceeds `maxBlockSize`, genes will be pre-clustered into blocks whose size should not exceed `maxBlockSize`.

`nThreads` non-negative integer specifying the number of parallel threads to be used by certain parts of correlation calculations. This option only has an effect on systems on which a POSIX thread library is available (which currently includes Linux and Mac OSX, but excludes Windows). If zero, the number of online processors will be used if it can be determined dynamically, otherwise correlation calculations will use 2 threads.

Value

Returns a lists containing network input parameters used for WGCNA, WGCNA module information, and quality control plots.

Note

This is a wrapper for WGCNA.

See Also

`blockwiseModules`

`adjacency`

Examples

```
sample_dat_dir<-system.file("extdata", "sample_dat.Rdata",
package = "GmicR", mustWork = TRUE)
load(sample_dat_dir)
GMIC_Builder<-Auto_WGCNA(sample_dat, mergeCutHeight = 0.35,
minModuleSize = 10)
```

`Batch_Net` *Generates a subgraph from query nodes*

Description

Generates a subgraph from query nodes

Usage

```
Batch_Net(bn_output, Node_ids, relationship_type = "nbr")
```

Arguments

`bn_output`	R object output from bn_tabu_gen function
`Node_ids`	vector containing the nodes for subgraph generation.
`relationship_type`	the relationship to be returned for the specified query nodes. The options are "mb", "nbr", "parents", "children". Default setting is "nbr".

Value

a subgraph containg the selected nodes and relationships.

`bn_tabu_gen` *Uses tabu search algorithm to learn the structure of discretized data.*

Description

Uses tabu search algorithm to learn the structure of discretized data.

Usage

```
bn_tabu_gen(
  Auto_WGCNA_OUTPUT,
  whitelist = NULL,
  blacklist = NULL,
  score = "bde",
  tabu = 50,
  iss = 10,
  cluster = NULL,
  debug = TRUE,
  bootstraps_replicates = 500
)
```

Arguments

Auto_WGCNA_OUTPUT	an R object generated by Auto_WGCNA and discretized using the Data_Prep function.
whitelist	a data frame with two columns (optionally labeled "from" and "to"), containing a set of arcs to be included in the graph.
blacklist	a data frame with two columns (optionally labeled "from" and "to"), containing a set of arcs not to be included in the graph.
score	character string indicating the score used for structure learning. If "bde" (default), prior is set to "uniform". If bds is used, the prior is set to "marginal".
tabu	a positive integer number, the length of the tabu list used in the tabu function.
iss	the imaginary sample size, used by the Bayesian Dirichlet scores (bde and bds) It is also known as “equivalent sample size”. The default value is equal to 10.
cluster	an optional cluster object from package **parallel**.
debug	a boolean value. If TRUE a lot of debugging output is printed; otherwise the function is completely silent.
bootstraps_replicates	an integer for the number of bootstraps_replicates used for structure learning. Default value is 500

Value

The learned bayesian network

See Also

arc.strength

hc

score

Examples

```
GMIC_Builder_disc_dir<-system.file("extdata", "GMIC_Builder_disc.Rdata",
package = "GmicR", mustWork = TRUE)
load(GMIC_Builder_disc_dir)

no_cores<-1
cl<-parallel::makeCluster(no_cores)

GMIC_net<-bn_tabu_gen(GMIC_Builder_disc,
cluster = cl,
bootstraps_replicates = 50, score = "bds")
parallel::stopCluster(cl)
```

`Data_Prep` *Discretizes biological assay data in preparation for bayensian network learning*

Description

Discretizes biological assay data in preparation for bayensian network learning

Usage

```
Data_Prep(
  Auto_WGCNA_OUTPUT = NULL,
  Remove_ME0 = FALSE,
  Numeric_Pheno_scores = NULL,
  xCell_Signatures = NULL,
  ibreaks = 60
)
```

Arguments

`Auto_WGCNA_OUTPUT`
: R object generated from Auto_WGCNA function.

`Remove_ME0`
: a logical value. If FALSE (default), ME0 is not removed. If TRUE the eigengene for module 0 is removed prior to analysis.

`Numeric_Pheno_scores`
: a data.frame with rows indicating sample ID and columns representing additional phenotype data to be included in BN learning. If NULL (default) no data will be included. If provided, the data.frame will be merged with MEs and discretized into three levels.

`xCell_Signatures`
: the name of the text file generated by xCell that contains the cell signature scores. If NULL (default) the only module eigenegnes will be processed. If not NULL and if Auto_WGCNA_OUTPUT is NULL, cell signature scores will be discretized.

`ibreaks`
: an integer that indicates the number of ibreaks used for discretization. The default value is 60.

Value

a list containing a data.frame with module eigenegnes merged with Xcell signature scores and discretized into three levels: L, M, H. If Auto_WGCNA_OUTPUT is NULL, both scaled and discretized cell signatures will be return.

Note

Please verify that the sample name formatting is consistent between both datasets. Rownames in the module eigengenes data.frame and the column names of xCell signatures scores text file are matched for merging. Only samples that are present in both will be processed!

Examples

```
file_dir<-system.file("extdata", "IRIS_xCell_sig.txt",
package = "GmicR", mustWork = TRUE)
Disc_Xcell_sig<-Data_Prep(xCell_Signatures=file_dir, ibreaks = 10)
Disc_Xcell_sig$disc_data
```

Gmic_viz *Visualized network*

Description

Visualized network

Usage

```
Gmic_viz(Auto_WGCNA_Output, Filter_unconnected_ME = TRUE)
```

Arguments

`Auto_WGCNA_Output`	R object with GMIC bayesian network
`Filter_unconnected_ME`	a logical value. If TRUE, the default, unconnected modules will be removed from the final network. If FALSE, all modules will be shown.

Value

a shiny object for network visualization.

Examples

```
GMIC_Final_dir<-system.file("extdata", "GMIC_Final.Rdata",
package = "GmicR", mustWork = TRUE)
load(GMIC_Final_dir)
if(interactive()){
Gmic_viz(GMIC_Final)}
```

GO_Module_NameR *GO enrichment for module names*

Description

GO enrichment for module names

Usage

```
GO_Module_NameR(Auto_WGCNA_OUTPUT, cutoff_size = NULL)
```

Arguments

Auto_WGCNA_OUTPUT	object returned by GSEAGO_Builder function
cutoff_size	integer indication the GO size cut off. If NULL (default) the term with the smallest Pvalue will be returned.

Value

a data.frame listing the GO name for each module. Additionally, a table similar to the output of Query_Prep function is returned, appended with module GO names.

Examples

```
GMIC_Builder_dir<-system.file("extdata", "GMIC_Builder.Rdata",
package = "GmicR", mustWork = TRUE)
load(GMIC_Builder_dir)
GMIC_Builder<-GO_Module_NameR(GMIC_Builder, cutoff_size = 100)
```

GSEAGO_Builder *GO enrichment for module names*

Description

GO enrichment for module names

Usage

```
GSEAGO_Builder(
  Auto_WGCNA_OUTPUT,
  species = "Homo sapiens",
  no_cores = 4,
  ontology = "BP",
  GO_conditional = FALSE,
  colname_correct = TRUE
)
```

Arguments

Auto_WGCNA_OUTPUT	output from Auto_WGCNA function.
species	either 'Homo sapiens' (default) or 'Mus musculus'.
no_cores	Number of cores to use. Default = 4.
ontology	string either 'BP'(Biological Process; default), 'CC'(Cellular Component), or 'MF' (Molecular Function).
GO_conditional	A logical indicating whether the calculation should condition on the GO structure. will not be carried out. If TRUE,
colname_correct	a logical value. If TRUE (default), "." in gene names will be replaced with "-". This corrects a name change that is induced by R when creating a data.frame. If FALSE, no changes will be made.

Value

Lists with gene ontology enrichment analysis, performed using GOstats, for each module.

Note

gene names must be official gene symbol

Examples

```
GMIC_Builder_dir<-system.file("extdata", "GMIC_Builder.Rdata",
                              package = "GmicR", mustWork = TRUE)
load(GMIC_Builder_dir)
GMIC_Builder$GSEAGO_Builder_Output<-NULL
Test_GMIC_Builder<-GSEAGO_Builder(GMIC_Builder, no_cores = 1)
summary(Test_GMIC_Builder$GSEAGO_Builder_Output)
```

InverseARCs *Identifies arcs between nodes with inverse relationships*

Description

Identifies arcs between nodes with inverse relationships

Usage

```
InverseARCs(Output, threshold = -0.3)
```

Arguments

Output	a data frame containing the output of BN_Conditions function.
threshold	number indicating the maximum slope for defining negative relationships. Default level is -0.3.

Value

arcs with inverse relationships

Examples

```
GMIC_net_dir<-system.file("extdata", "GMIC_net.Rdata",
package = "GmicR", mustWork = TRUE)
load(GMIC_net_dir)
GMIC_Final<-InverseARCs(GMIC_net, threshold = -0.3)
```

Query_Prep *Query Prep*

Description

Query Prep

Usage

```
Query_Prep(
  Auto_WGCNA_OUTPUT,
  numGenes = 500,
  Find_hubs = FALSE,
  nThreads = NULL,
  calculate_intramodularConnectivity = TRUE
)
```

Arguments

Auto_WGCNA_OUTPUT	R object generated by Auto_WGCNA function.
numGenes	integer indicating the number of random genes to test for hub gene detection. Default is 500.
Find_hubs	logical value. If TRUE, module hub genes will be returned. If FALSE (default), intramodularConnectivity will be returned without hub gene identification.
nThreads	non-negative integer specifying the number of parallel threads to be used by certain parts of correlation calculations. This option only has an effect on systems on which a POSIX thread library is available (which currently includes Linux and Mac OSX, but excludes Windows). If zero, the number of online processors will be used if it can be determined dynamically, otherwise correlation calculations will use 2 threads.
calculate_intramodularConnectivity	a logical value. If TRUE (default), the intramodularConnectivity will be calucculated using the intramodularConnectivity function from WGCNA. If FALSE, a table of modules and genes will be returned without intramodularConnectivity values.

Value

a data.frame detailing the gene symbols for each module. Gene intramodularConnectivity may also be returned. If detected, hub genes are annotated.

Examples

```
GMIC_Builder_dir<-system.file("extdata", "GMIC_Builder.Rdata",
package = "GmicR", mustWork = TRUE)
load(GMIC_Builder_dir)
GMIC_Builder<-Query_Prep(GMIC_Builder,  Find_hubs = TRUE)
head(GMIC_Builder$Query)
```

---

xCell_loader *Scales and centers data by sample/row in preparation for discretization*

Description

Scales and centers data by sample/row in preparation for discretization

Usage

```
xCell_loader(File = NULL)
```

Arguments

File	the name of the text file generated by xCell that contains the cell signature scores.

Value

xCell signatures scaled and centered by sample. For GMIC, ImmuneScore, StromaScore, and MicroenvironmentScore are removed.

Examples

```
file_dir<-system.file("extdata", "IRIS_xCell_sig.txt",
package = "GmicR", mustWork = TRUE)
Xcell_sig<-xCell_loader(file_dir)
plot(Xcell_sig$Bcells)
```

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