Package 'MPAC'

November 12, 2026

Title Multi-omic Pathway Analysis of Cells

Version 1.6.0

Description Multi-omic Pathway Analysis of Cells (MPAC), integrates multi-omic data for understanding cellular mechanisms. It predicts novel patient groups with distinct pathway profiles as well as identifying key pathway proteins with potential clinical associations. From CNA and RNA-seq data, it determines genes' DNA and RNA states (i.e., repressed, normal, or activated), which serve as the input for PARADIGM to calculate Inferred Pathway Levels (IPLs). It also permutes DNA and RNA states to create a background distribution to filter IPLs as a way to remove events observed by chance. It provides multiple methods for downstream analysis and visualization.

License GPL-3

Encoding UTF-8

Roxygen list(markdown = TRUE)

RoxygenNote 7.3.2

Depends R (>= 4.4.0)

URL https://github.com/pliu55/MPAC

BugReports https://github.com/pliu55/MPAC/issues

Imports data.table (>= 1.14.2), SummarizedExperiment (>= 1.30.2), BiocParallel (>= 1.28.3), fitdistrplus (>= 1.1), igraph (>= 1.4.3), BiocSingular (>= 1.10.0), S4Vectors (>= 0.32.3), SingleCellExperiment (>= 1.16.0), bluster (>= 1.4.0), fgsea (>= 1.20.0), scran (>= 1.22.1), ComplexHeatmap (>= 2.16.0), circlize (>= 0.4.16), scales (>= 1.3.0), stringr (>= 1.5.1), viridis (>= 0.6.5), ggplot2 (>= 3.5.1), ggraph (>= 2.2.1), survival (>= 3.7), survminer (>= 0.4.9), grid, stats

Suggests rmarkdown, knitr, svglite, bookdown(>= 0.34), testthat (>= 3.0.0)

Config/testthat/edition 3

VignetteBuilder knitr

biocViews Software, Technology, Sequencing, RNASeq, Survival, Clustering, ImmunoOncology

SystemsRequirements The `runPrd()` function requires an external software named PARADIGM. For details, please see the 'Required external software' section in vignette's 'Run PARADIGM: runPrd()'.

git_url https://git.bioconductor.org/packages/MPAC

git_branch RELEASE_3_23

git_last_commit 2a03ee8

git_last_commit_date 2026-04-28

Repository Bioconductor 3.23

Date/Publication 2026-11-11

Author Peng Liu [aut, cre] (ORCID: <https://orcid.org/0000-0001-5655-2259>),
Paul Ahlquist [aut],
Irene Ong [aut],
Anthony Gitter [aut]

Maintainer Peng Liu <pliu55.wisc+bioconductor@gmail.com>

Contents

clSamp . . . 3
colPermIPL . . . 4
colRealIPL . . . 4
conMtf . . . 5
fltByPerm . . . 6
getSignifOvrOnCl . . . 7
ovrGMT . . . 7
pltConMtf . . . 8
pltMtfPrtIPL . . . 9
pltNeiStt . . . 10
pltOvrHm . . . 11
pltSttKM . . . 12
ppCnInp . . . 13
ppPermInp . . . 13
ppRealInp . . . 14
ppRnaInp . . . 15
ppRunPrd . . . 16
runPermPrd . . . 16
runPrd . . . 17
subNtw . . . 18

Index **20**

clSamp *Cluster samples by pathway over-representation*

Description

Cluster samples by pathway over-representation

Usage

```
clSamp(ovrmat, n_neighbors = 10, n_random_runs = 200, threads = 1)
```

Arguments

ovrmat	A matrix of gene set over-representation adjusted p-values with rows as gene sets and columns as samples. It is the output from `ovrGMT()`.
n_neighbors	Number of neighbors for clustering. A larger number is recommended if the size of samples is large. Default: 10.
n_random_runs	Number of random runs. Due to randomness introduced to the Louvain algorithm in R igraph 1.3.0 (https://github.com/igraph/rigraph/issues/539), a large number of runs are recommended to evaluate randomness in the clustering results. Default: 200, which shall be safe for sample size < 50. Please increase it accordingly for a larger sample size.
threads	Number of threads to run in parallel. Default: 1

Value

A data table with each row representing one clustering result, and the first column denotes the number of occurrences of a clustering result and the rest of columns indicating each sample's cluster index. Rows are ordered by the number of occurrences from high to low.

Examples

```
fovr = system.file('extdata/clSamp/ovrmat.rds', package='MPAC')
ovrmat = readRDS(fovr)

clSamp(ovrmat)
```

colPermIPL *Collect Inferred Pathway Levels (IPLs) from PARADIGM runs on permuted data*

Description

Collect Inferred Pathway Levels (IPLs) from PARADIGM runs on permuted data

Usage

```
colPermIPL(indir, n_perms, sampleids = NULL, threads = 1)
```

Arguments

indir	Input folder that saves PARADIGM results. It should be set as the same as outdir as in runPrd().
n_perms	Number of permutations to collect.
sampleids	Sample IDs for which IPLs to be collected. If not provided, all files with suffix '_ipl.txt' in indir will be collected. Default: NULL.
threads	Number of threads to run in parallel. Default: 1

Value

A data.table object with columns of permutation index, pathway entities and their IPLs.

Examples

```
indir = system.file('/extdata/runPrd/', package='MPAC')
n_perms = 3

colPermIPL(indir, n_perms)
```

colRealIPL *Collect Inferred Pathway Levels (IPLs) from PARADIGM runs on real data*

Description

Collect Inferred Pathway Levels (IPLs) from PARADIGM runs on real data

Usage

```
colRealIPL(indir, sampleids = NULL, file_tag = NULL)
```

Arguments

indir	Input folder that saves PARADIGM results. It should be set as the same as outdir as in runPrd().
sampleids	Sample IDs for which IPLs to be collected. If not provided, all files with suffix '_ipl.txt' in indir will be collected. Default: NULL.
file_tag	A string of output file name tag. Default: NULL

Value

A data.table object with columns of pathway entities and their IPLs.

Examples

```
indir = system.file('/extdata/runPrd/', package='MPAC')

colRealIPL(indir)
```

---

conMtf *Find consensus pathway motifs from a list of pathways*

Description

Find consensus pathway motifs from a list of pathways

Usage

```
conMtf(subntwl, omic_genes = NULL, min_mtf_n_nodes = 5)
```

Arguments

subntwl	A list of igraph objects representing input pathways from different samples. It is the output from subNtw()
omic_genes	A vector of gene symbols to narrow down over-representation calculation to only those with input genomic data. If not provided, all genes in the GMT file will be considered. Default: NULL.
min_mtf_n_nodes	Number of minimum nodes in a motif. Default: 5

Value

A list of igraph objects representing consensus pathway motifs

Examples

```
fsubntwl = system.file('extdata/conMtf/subntwl.rds', package='MPAC')
subntwl = readRDS(fsubntwl)

fomic_gns = system.file('extdata/TcgaInp/inp_focal.rds', package='MPAC')
omic_gns = rownames(readRDS(fomic_gns))

conMtf(subntwl, omic_gns, min_mtf_n_nodes=50)
```

`fltByPerm` *Filter IPLs from real data by distribution from permuted data*

Description

Filter IPLs from real data by distribution from permuted data

Usage

```
fltByPerm(realdt, permdt, threads = 1)
```

Arguments

`realdt`	A data.table object containing entities and their IPLs from real data. It is the output from `colRealIPL()`.
`permdt`	A data.table object containing permutation index, entities and their IPLs from permuted data. It is the output from `colPermIPL()`.
`threads`	Number of threads to run in parallel. Default: 1

Value

A matrix of filtered IPLs with rows as entities and columns as samples. Entities with IPLs observed by chance are set to NA.

Examples

```
freal = system.file('extdata/fltByPerm/real.rds', package='MPAC')
fperm = system.file('extdata/fltByPerm/perm.rds', package='MPAC')
realdt = readRDS(freal)
permdt = readRDS(fperm)

fltByPerm(realdt, permdt)
```

`getSignifOvrOnCl` *Get significantly over-represented gene sets for clustered samples*

Description

Get significantly over-represented gene sets for clustered samples

Usage

```
getSignifOvrOnCl(ovrmat, cldt, min_frc = 0.8)
```

Arguments

`ovrmat`	A matrix containing over-representation adjusted P with rows as gene set names and columns as sample IDs. It is the output of the `ovrGMT()` function.
`cldt`	A data table with each row representing one clustering result, and the first column denotes the number of occurrences of a clustering result and the rest of columns indicating each sample's cluster index. It is the output of the `clSamp()` function. Only the most frequent clustering result will be used to plot.
`min_frc`	A minimum fraction of samples in a cluster that have a gene set significantly over-represented (adjusted P < 0.05). This is used to select gene sets to plot. Default: 0.8

Value

A list of a matrix and a data.table object. The matrix has rows as over-represented gene sets, columns as samples, and each cell stores an adjusted P for over-representation. The data.table has the clustering informations with samples in the same order as the matrix's column.

Examples

```
ovrmat <- system.file('extdata/pltOvrHm/ovr.rds',package='MPAC') |> readRDS()
cldt   <- system.file('extdata/pltOvrHm/cl.rds', package='MPAC') |> readRDS()

getSignifOvrOnCl(ovrmat, cldt)
```

`ovrGMT` *Calculate over-representation of gene sets in each sample by genes from sample's largest sub-pathway*

Description

Calculate over-representation of gene sets in each sample by genes from sample's largest sub-pathway

Usage

```
ovrGMT(subntwlist, fgmt, omic_genes = NULL, threads = 1)
```

Arguments

subntwlist	A list of igraph objects represented the largest sub-pathway for each sample. It is the output of subNtw().
fgmt	A gene set GMT file. This will be the same file used for the gene set over-representation calculation in the next step. It is used here to ensure output sub-pathway contains a minimum number of genes from to-be-used gene sets.
omic_genes	A vector of gene symbols to narrow down over-representation calculation to only those with input genomic data. If not provided, all genes in the GMT file will be considered. Default: NULL.
threads	Number of threads to run in parallel. Default: 1

Value

A matrix containing over-representation adjusted P with rows as gene set names and columns as sample IDs.

Examples

```
fsubntwl  = system.file('extdata/subNtw/subntwl.rds',    package='MPAC')
fgmt      = system.file('extdata/ovrGMT/fake.gmt',       package='MPAC')
fomic_gns = system.file('extdata/TcgaInp/inp_focal.rds', package='MPAC')
subntwl  = readRDS(fsubntwl)
omic_gns = rownames(readRDS(fomic_gns))

ovrGMT(subntwl, fgmt, omic_gns)
```

---

pltConMtf *Plot consensus pathway submodules*

Description

Plot consensus pathway submodules

Usage

```
pltConMtf(grphl, proteins = NULL)
```

Arguments

grphl	A list of igraph objects representing consensus pathway submodules. It is the output from conMtf().
proteins	A vector of protein symbols to highlight in the plot. Default: no protein will be highlighted.

Value

a plot of consensus pathway submodules

Examples

```
grphl <- system.file('extdata/pltMtfPrtIPL/grphl.rds',package='MPAC') |>
         readRDS()

proteins <- system.file('extdata/TcgaInp/inp_focal.rds', package='MPAC') |>
            readRDS() |> rownames() |> c('CD3G')

pltConMtf(grphl, proteins) |> print()
```

`pltMtfPrtIPL` *Plot a heatmap of IPLs on proteins from consensus pathway submodules*

Description

Plot a heatmap of IPLs on proteins from consensus pathway submodules

Usage

```
pltMtfPrtIPL(fltmat, cldt, grphl, proteins = NULL)
```

Arguments

`fltmat`	A matrix contains filterd IPL with rows as entity and column as samples. This is the output from `fltByPerm()`. Entity with NA value will be set to 0 and plotted as in 'normal' state.
`cldt`	A data table with each row representing one clustering result, and the first column denotes the number of occurrences of a clustering result and the rest of columns indicating each sample's cluster index. It is the output of the `clSamp()` function. Only the most frequent clustering result will be used to plot.
`grphl`	A list of igraph objects representing consensus pathway submodules. It is the output from `conMtf()`.
`proteins`	A vector of proteins, of which IPLs to plot. Default: all proteins that in both `grphl` and `fltmat`.

Value

A heatmap of IPLs o proteins from consensus pathway submodules

Examples

```
fltmat <- system.file('extdata/pltSttKM/ipl.rds',package='MPAC') |> readRDS()
cldt <- system.file('extdata/pltMtfPrtIPL/cl.rds',package='MPAC')|> readRDS()
grphl <- system.file('extdata/pltMtfPrtIPL/grphl.rds',package='MPAC') |>
         readRDS()

pltMtfPrtIPL(fltmat, cldt, grphl, proteins=c('CD247', 'FASLG'))
```

`pltNeiStt` *Plot a heatmap of pathway and omic states of a protein and its pathway neighbors*

Description

Plot a heatmap of pathway and omic states of a protein and its pathway neighbors

Usage

```
pltNeiStt(real_se, fltmat, fpth, protein = "")
```

Arguments

`real_se`	A SummarizedExperiment object of PARADIGM CNA and RNA states. It is the output fromm `ppRealInp()` and must contain the omic states for the one defined in the `protein` argument.
`fltmat`	A matrix contains filterd IPL with rows as entity and column as samples. This is the output from `fltByPerm()`. Entity with NA value will be set to 0 and plotted as in 'normal' state.
`fpth`	Name of a pathway file for PARADIGM.
`protein`	Name of the protein to plot. It requires to have CN and RNA state data, as well as pathway data from the input. Default: "

Value

A heatmap of pathway and omic states of a protein and its pathway neighbors

Examples

```
fpth = system.file('extdata/Pth/tiny_pth.txt', package='MPAC')

freal = system.file('extdata/pltNeiStt/inp_real.rds', package='MPAC')
fflt  = system.file('extdata/pltNeiStt/fltmat.rds',   package='MPAC')

real_se = readRDS(freal)
fltmat = readRDS(fflt)
protein = 'CD86'
```

```
pltNeiStt(real_se, fltmat, fpth, protein)
```

`pltOvrHm` *Plot a heatmap of over-represented gene sets for clustered samples*

Description

Plot a heatmap of over-represented gene sets for clustered samples

Usage

```
pltOvrHm(ovrmat, cldt, min_frc = 0.8)
```

Arguments

`ovrmat`	A matrix containing over-representation adjusted P with rows as gene set names and columns as sample IDs. It is the output of the `ovrGMT()` function.
`cldt`	A data table with each row representing one clustering result, and the first column denotes the number of occurrences of a clustering result and the rest of columns indicating each sample's cluster index. It is the output of the `clSamp()` function. Only the most frequent clustering result will be used to plot.
`min_frc`	A minimum fraction of samples in a cluster that have a gene set significantly over-represented (adjusted P < 0.05). This is used to select gene sets to plot. Default: 0.8

Value

A heatmap with rows as over-represented gene sets and columns as samples splited by clusters.

Examples

```
ovrmat <- system.file('extdata/pltOvrHm/ovr.rds',package='MPAC') |> readRDS()
cldt   <- system.file('extdata/pltOvrHm/cl.rds', package='MPAC') |> readRDS()

pltOvrHm(ovrmat, cldt)
```

`pltSttKM` *Plot a Kaplan-Meier curve for samples stratified by given protein(s)' pathway states*

Description

Plot a Kaplan-Meier curve for samples stratified by given protein(s)' pathway states

Usage

```
pltSttKM(
  cdrmat,
  fltmat,
  event = "OS",
  time = "OS_days",
  proteins = NULL,
  strat_func = ">0"
)
```

Arguments

`cdrmat`	A matrix containing survival data with rows as patient samples and columns as survival event and time.
`fltmat`	A matrix contains filterd IPL with rows as entity and column as samples. This is the output from `fltByPerm()`. Entity with NA value will be set to 0 and plotted as in 'normal' state.
`event`	The column name in `cdrmat` to indicate survival event. Default: 'OS'.
`time`	The column name in `cdrmat` to indicate survival time. Default: 'OS_days'.
`proteins`	Rowname(s) in `fltmat`. Its/their pathway states will be used to stratify patient samples. Default: all proteins in `fltmat` will be used.
`strat_func`	A function applied on protein(s) pathway states to stratify patient samples. Available options: '>0', '<0', Default: '>0', i.e., IPL >0 vs. the rest.

Value

A Kaplan-Meier plot

Examples

```
cdrmat <- system.file('extdata/pltSttKM/cdr.rds',package='MPAC') |> readRDS()
fltmat <- system.file('extdata/pltSttKM/ipl.rds',package='MPAC') |> readRDS()

pltSttKM(cdrmat, fltmat, event='OS', time='OS_days',
         proteins=c('CD247', 'FASLG'))
```

ppCnInp *Prepare input copy-number (CN) alteration data to run PARADIGM*

Description

Prepare input copy-number (CN) alteration data to run PARADIGM

Usage

```
ppCnInp(cn_tumor_mat)
```

Arguments

cn_tumor_mat	A matrix of tumor CN focal data with rows as genes and columns as samples. A value of 0 means normal CN, > 0 means amplification, and < 0 means deletion.

Value

A SummarizedExperiment object of CN state for PARADIGM

Examples

```
fcn = system.file('extdata/TcgaInp/focal_tumor.rds', package='MPAC')
cn_tumor_mat = readRDS(fcn)

ppCnInp(cn_tumor_mat)
```

ppPermInp *Permute input genomic state data between genes in the same sample*

Description

Permute input genomic state data between genes in the same sample

Usage

```
ppPermInp(real_se, n_perms=100, threads=1)
```

Arguments

real_se	A SummarizedExperiment object of CN and RNA states from real samples with rows as genes and columns as samples. It is the output from ppRealInp().
n_perms	Number of permutations. Default: 100
threads	Number of threads to run in parallel. Default: 1

Value

A list of SummarizedExperiment objects of permuted CN and RNA states. The metadata `i` in each obbect denotes its permutation index.

Examples

```
freal = system.file('extdata/TcgaInp/inp_real.rds', package='MPAC')
real_se = readRDS(freal)

ppPermInp(real_se, n_perms=3)
```

`ppRealInp` *Prepare input copy-number (CN) alteration and RNA data to run PARADIGM*

Description

Prepare input copy-number (CN) alteration and RNA data to run PARADIGM

Usage

```
ppRealInp(
  cn_tumor_mat,
  rna_tumor_mat,
  rna_normal_mat,
  rna_n_sd = 2,
  threads = 1
)
```

Arguments

`cn_tumor_mat`	A matrix of tumor CN focal data with rows as genes and columns as samples. A value of 0 means normal CN, > 0 means amplification, and < 0 means deletion.
`rna_tumor_mat`	A matrix of RNA data from tumor samples with rows as genes and columns as samples
`rna_normal_mat`	A matrix of RNA data from normal samples with rows as genes and columns as samples
`rna_n_sd`	Standard deviation range from fitted normal samples to define RNA state. Default: 2, i.e. 2*sd
`threads`	Number of threads to run in parallel. Default: 1

Value

A SummarizedExperiment object of CN and RNA state for PARADIGM

Examples

```
fcn = system.file('extdata/TcgaInp/focal_tumor.rds', package='MPAC')
ftumor = system.file('extdata/TcgaInp/log10fpkmP1_tumor.rds', package='MPAC')
fnorm = system.file('extdata/TcgaInp/log10fpkmP1_normal.rds', package='MPAC')

cn_tumor_mat = readRDS(fcn)
rna_tumor_mat = readRDS(ftumor)
rna_norm_mat  = readRDS(fnorm)

ppRealInp(cn_tumor_mat, rna_tumor_mat, rna_norm_mat)
```

---

ppRnaInp *Prepare input RNA data to run PARADIGM*

Description

Prepare input RNA data to run PARADIGM

Usage

```
ppRnaInp(rna_tumor_mat, rna_normal_mat, rna_n_sd = 2, threads = 1)
```

Arguments

rna_tumor_mat	A matrix of RNA data from tumor samples with rows as genes and columns as samples
rna_normal_mat	A matrix of RNA data from normal samples with rows as genes and columns as samples
rna_n_sd	Standard deviation range from fitted normal samples to define RNA state. Default: 2, i.e. 2*sd
threads	Number of threads to run in parallel. Default: 1

Value

A SummarizedExperiment of RNA state for PARADIGM

Examples

```
ftumor = system.file('extdata/TcgaInp/log10fpkmP1_tumor.rds', package='MPAC')
fnorm = system.file('extdata/TcgaInp/log10fpkmP1_normal.rds', package='MPAC')
rna_tumor_mat = readRDS(ftumor)
rna_norm_mat  = readRDS(fnorm)

ppRnaInp(rna_tumor_mat, rna_norm_mat, threads=2)
```

ppRunPrd *Prepare required files to run PARADIGM*

Description

Prepare required files to run PARADIGM

Usage

```
ppRunPrd(pat, cnmat, rnamat, outdir, file_tag=NULL)
```

Arguments

pat	Sample ID
cnmat	CN matrix
rnamat	RNA matrix
outdir	Output folder to save all results.
file_tag	A string of output file name tag. Default: NULL

Value

None

runPermPrd *Run PARADIGM on permuted data*

Description

Run PARADIGM on permuted data

Usage

```
runPermPrd(perml, fpth, outdir,
    PARADIGM_bin=NULL, nohup_bin=NULL, sampleids=NULL, threads=1)
```

Arguments

perml	A list of SummarizedExperiment objects of permuted CNA and RNA states generated by ppPermInp().
fpth	Name of a pathway file for PARADIGM.
outdir	Output folder to save all results.
PARADIGM_bin	PARADIGM binary, which can be downloaded from https://github.com/sng87/paradigm-scripts/tree/master/public/exe. Note that the binary is only available for Linux or MacOS. Default: NULL

nohup_bin	nohup binary, which is used for long running PARADIGM jobs. Default: NULL
sampleids	A vector of sample IDs to run PARADIGM on. If not provided, all the samples that exist in both copy-number alteration and RNA files will be ran. Default: NULL
threads	Number of threads to run in parallel. Default: 1

Value

None

Examples

```
fperm = system.file('extdata/TcgaInp/inp_perm.rds', package='MPAC')
perml = readRDS(fperm)
fpth = system.file('extdata/Pth/tiny_pth.txt', package='MPAC')
outdir = tempdir()
paradigm_bin = '/path/to/PARADIGM'  ## change to binary location
pat = 'TCGA-CV-7100'

# depends on external PARADIGM binary, do not run
runPermPrd(perml, fpth, outdir, paradigm_bin, sampleids=c(pat))
```

---

runPrd *Run PARADIGM on multi-omic data*

Description

Run PARADIGM on multi-omic data

Usage

```
runPrd(real_se, fpth, outdir, PARADIGM_bin=NULL, nohup_bin=NULL,
    sampleids=NULL, file_tag=NULL, threads=1)
```

Arguments

real_se	A SummarizedExperiment object of PARADIGM CNA and RNA states. It is the same matrix as the output from `ppRealInp()`.
fpth	Name of a pathway file for PARADIGM.
outdir	Output folder to save all results.
PARADIGM_bin	PARADIGM binary, which can be downloaded from https://github.com/sng87/paradigm-scripts/tree/master/public/exe. Note that the binary is only available for Linux or MacOS. Default: NULL
nohup_bin	nohup binary, which is used for long running PARADIGM jobs. Default: NULL

sampleids	A vector of sample IDs to run PARADIGM on. If not provided, all the samples that exist in both copy-number alteration and RNA files will be ran. Default: NULL
file_tag	A string of output file name tag. Default: NULL
threads	Number of threads to run in parallel. Default: 1

Value

None

Examples

```
freal = system.file('extdata/TcgaInp/inp_real.rds', package='MPAC')
real_se  = readRDS(freal)

fpth = system.file('extdata/Pth/tiny_pth.txt', package='MPAC')
outdir = tempdir()
paradigm_bin = '/path/to/PARADIGM'  ## change to binary location

# depends on external PARADIGM binary
runPrd(real_se, fpth, outdir, paradigm_bin, sampleids=c('TCGA-CV-7100'))
```

---

subNtw *Subset pathways by IPL results*

Description

Subset pathways by IPL results

Usage

```
subNtw(fltmat, fpth, fgmt, min_n_gmt_gns = 2, threads = 1)
```

Arguments

fltmat	A matrix contains filterd IPL with rows as 'entity' and column as samples. This is the output from `fltByPerm()`. Entity with NA in all columns will be ignored.
fpth	Name of a pathway file for PARADIGM.
fgmt	A gene set GMT file. This will be the same file used for the gene set over-representation calculation in the next step. It is used here to ensure output sub-pathway contains a minimum number of genes from to-be-used gene sets.
min_n_gmt_gns	Minimum number of genes from the GMT file in the output sub-pathway. Default: 2.
threads	Number of threads to run in parallel. Default: 1

Value

A list of igraph objects representing the largest sub-pathway for each sample.

Examples

```
fflt = system.file('extdata/fltByPerm/flt_real.rds', package='MPAC')
fltmat = readRDS(fflt)
fpth = system.file('extdata/Pth/tiny_pth.txt',       package='MPAC')
fgmt = system.file('extdata/ovrGMT/fake.gmt',        package='MPAC')

subNtw(fltmat, fpth, fgmt, min_n_gmt_gns=1)
```

Index

clSamp, 3
colPermIPL, 4
colRealIPL, 4
conMtf, 5

fltByPerm, 6

getSignifOvrOnCl, 7

ovrGMT, 7

pltConMtf, 8
pltMtfPrtIPL, 9
pltNeiStt, 10
pltOvrHm, 11
pltSttKM, 12
ppCnInp, 13
ppPermInp, 13
ppRealInp, 14
ppRnaInp, 15
ppRunPrd, 16

runPermPrd, 16
runPrd, 17

subNtw, 18