Package 'enhancerHomologSearch'

November 12, 2026

Type Package

Title Identification of putative mammalian orthologs to given enhancer

Version 1.18.0

Description Get ENCODE data of enhancer region via H3K4me1 peaks and search homolog regions for given sequences. The candidates of enhancer homolog regions can be filtered by distance to target TSS. The top candidates from human and mouse will be aligned to each other and then exported as multiple alignments with given enhancer.

BugReports https://github.com/jianhong/enhancerHomologSearch/issues

URL https://jianhong.github.io/enhancerHomologSearch

Depends R (>= 4.1.0), methods

Imports BiocGenerics, Biostrings, BSgenome, BiocParallel, BiocFileCache, Seqinfo, GenomicRanges, httr, IRanges, jsonlite, motifmatchr, Matrix, pwalign, rtracklayer, Rcpp, S4Vectors, stats, utils

Suggests GenomeInfoDb, knitr, rmarkdown, BSgenome.Drerio.UCSC.danRer10, BSgenome.Hsapiens.UCSC.hg38, BSgenome.Mmusculus.UCSC.mm10, TxDb.Hsapiens.UCSC.hg38.knownGene, org.Hs.eg.db, TxDb.Mmusculus.UCSC.mm10.knownGene, org.Mm.eg.db, MotifDb, testthat, TFBSTools

biocViews Sequencing, GeneRegulation, Alignment

License GPL (>= 2)

Encoding UTF-8

VignetteBuilder knitr

RoxygenNote 7.3.2

LinkingTo Rcpp

git_url https://git.bioconductor.org/packages/enhancerHomologSearch

git_branch RELEASE_3_23

git_last_commit c43232e

git_last_commit_date 2026-04-28

Repository Bioconductor 3.23

Date/Publication 2026-11-11

Author Jianhong Ou [aut, cre] (ORCID: <https://orcid.org/0000-0002-8652-2488>),
Valentina Cigliola [dtc],
Kenneth Poss [fnd]

Maintainer Jianhong Ou <jou@morgridge.org>

Contents

alignment 2
alignmentOne 4
conservedMotifs 4
Enhancers-class 6
getENCODEdata 8
motifs 9
queryEncode 9
saveAlignments 10
searchTFBPS 11
shuffle 12

Index **13**

`alignment` *Output*

Description

Do pairwise alignment for query enhancer to target genome

Usage

```
alignment(
  query,
  subject,
  method = c("ClustalW", "Muscle"),
  cluster = c("nj", "upgma", "upgmamax", "upgmamin", "upgmb"),
  substitutionMatrix = c("iub", "clustalw"),
  gapOpening = ifelse(method[1] == "ClustalW", 15, 400),
  gapExtension = ifelse(method[1] == "ClustalW", 6.66, 0),
  maxiters = ifelse(method[1] == "ClustalW", 3, 16),
  order = c("aligned", "input"),
  ...
)
```

Arguments

query	An object of DNAStringSet to represent enhancer
subject	An list of objects of Enhancers.
method	specifies the multiple sequence alignment to be used; currently, "ClustalW", and "Muscle" are supported. Default is "Muscle"
cluster	The clustering method which should be used. Possible values are "nj" (default) and "upgma". In the original ClustalW implementation, this parameter is called clustering.
substitutionMatrix	substitution matrix for scoring matches and mismatches; The valid choices for this parameter are "iub" and "clustalw". In the original ClustalW implementation, this parameter is called matrix.
gapOpening	gap opening penalty; the default is 400 for DNA sequences and 420 for RNA sequences. The default for amino acid sequences depends on the profile score settings: for the setting le=TRUE, the default is 2.9, for sp=TRUE, the default is 1,439, and for sv=TRUE, the default is 300. Note that these defaults may not be suitable if custom substitution matrices are being used. In such a case, a sensible choice of gap penalties that fits well to the substitution matrix must be made.
gapExtension	gap extension penalty; the default is 0.
maxiters	maximum number of iterations; the default is 16.
order	how the sequences should be ordered in the output object; if "aligned" is chosen, the sequences are ordered in the way the multiple sequence alignment algorithm orders them. If "input" is chosen, the sequences in the output object are ordered in the same way as the input sequences.
...	Parameters can be used by Muscle, or ClustalW.

Value

An object of Enhancers.

Examples

```
library(BSgenome.Hsapiens.UCSC.hg38)
library(BSgenome.Mmusculus.UCSC.mm10)
library(BSgenome.Drerio.UCSC.danRer10)
LEN <- GRanges("chr4", IRanges(19050041, 19051709))
seqEN <- getSeq(BSgenome.Drerio.UCSC.danRer10, LEN)
aln_hs <- readRDS(system.file("extdata", "aln_hs.rds",
               package="enhancerHomologSearch"))
genome(aln_hs) <- Hsapiens
aln_mm <- readRDS(system.file("extdata", "aln_mm.rds",
               package="enhancerHomologSearch"))
genome(aln_mm) <- Mmusculus
al <- alignment(seqEN, list(human=aln_hs, mouse=aln_mm),
                method="ClustalW", order="input")
```

`alignmentOne` *Get alignment scores*

Description

Do pairwise alignment for query enhancer to target genome

Usage

```
alignmentOne(query, subject, block = 1000, bpparam = bpparam(), ...)
```

Arguments

`query`	An object of DNAStringSet to represent enhancer
`subject`	Output of getENCODEdata. An object of Enhancers
`block`	The size of sequences to do alignment. Increase the size will increase the memory cost. Default 1000.
`bpparam`	BiocParallel parameters.
`...`	not used.

Value

An object of Enhancers.

Examples

```
library(BiocParallel)
bpparam <- MulticoreParam(workers = 1, tasks=200, progressbar=TRUE)
library(BSgenome.Hsapiens.UCSC.hg38)
peaks <- GRanges("chr1", IRanges(seq(5000, 50000, by=1000), width=1000))
peaks$id <- paste(seq_along(peaks), 1, sep="_")
subj <- Enhancers(genome=Hsapiens, peaks=peaks)
q <- getSeq(Hsapiens, GRanges("chr1", IRanges(90000, width=1000)))
ao <- alignmentOne(q, subj, bpparam=bpparam)
```

`conservedMotifs` *check the conserved motifs in the orthologs*

Description

Print the conserved motifs in the alignments

Usage

```
conservedMotifs(
  aln,
  aln_list,
  PWMs,
  queryGenome,
  background = "genome",
  ...,
  output_folder,
  format = c("txt", "html")
)
```

Arguments

aln	alignment of multiple DNAs. Output of alignment function.
aln_list	The list of output of searchTFBPS such as for human and mouse.
PWMs	The Position Weight Matrix list represented as a numeric matrix. Object of PWMatrixList or PFMatrixList.
queryGenome	An object of BSgenome for query enhancer.
background	Background nucleotide frequencies. Default is "genome". Refer matchMotifs for details.
...	Other parameters can be passed to to matchMotifs.
output_folder	Output folder name.
format	The format of output files with motif match positions. Available formats are 'txt' and 'html'. Default is 'txt'.

Value

A list of XStringViews

Examples

```
library(BSgenome.Hsapiens.UCSC.hg38)
library(BSgenome.Mmusculus.UCSC.mm10)
library(BSgenome.Drerio.UCSC.danRer10)
LEN <- GRanges("chr4", IRanges(19050041, 19051709))
seqEN <- getSeq(BSgenome.Drerio.UCSC.danRer10, LEN)
aln_hs <- readRDS(system.file("extdata", "aln_hs.rds",
              package="enhancerHomologSearch"))
genome(aln_hs) <- Hsapiens
aln_mm <- readRDS(system.file("extdata", "aln_mm.rds",
              package="enhancerHomologSearch"))
genome(aln_mm) <- Mmusculus
al <- alignment(seqEN, list(human=aln_hs, mouse=aln_mm),
               method="ClustalW", order="input")
data(motifs)
conservedMotifs(al[[1]], list(human=aln_hs, mouse=aln_mm),
               motifs[["dist60"]], Drerio)
```

Enhancers-class *Class* "Enhancers"

Description

An object of class "Enhancers" represents the output of function getENCODEdata, which includes the sequences of enhancers and their genomic coordinates.

Usage

```
Enhancers(genome, peaks, TFBP, TFBP0)

## S4 method for signature 'Enhancers'
x$name

## S4 replacement method for signature 'Enhancers'
x$name <- value

## S4 method for signature 'Enhancers,ANY'
distance(x)

## S4 replacement method for signature 'Enhancers'
distance(x) <- value

## S4 method for signature 'Enhancers'
tfbp(x)

## S4 method for signature 'Enhancers'
query_tfbp(x)

## S4 method for signature 'Enhancers'
getSeq(x, ...)

## S4 method for signature 'Enhancers,ANY'
subsetByOverlaps(
  x,
  ranges,
  maxgap = -1L,
  minoverlap = 0L,
  type = c("any", "start", "end", "within", "equal"),
  invert = FALSE,
  ...
)

## S4 method for signature 'Enhancers'
subset(x, ...)
```

```
## S4 method for signature 'Enhancers'
seqinfo(x)

## S4 method for signature 'Enhancers'
genome(x)

## S4 replacement method for signature 'Enhancers'
genome(x) <- value

## S4 method for signature 'Enhancers'
peaks(x)

## S4 replacement method for signature 'Enhancers'
peaks(x) <- value

## S4 method for signature 'Enhancers'
show(object)
```

Arguments

genome	An object of BSgenome.
peaks	An object of GRanges.
TFBP	An object of lgCMatrix.
TFBP0	An vector of logical. "Enhancers"
x	An object of "Enhancers"
name	Slot name.
value	The values.
...	parameters can be passed to upstream functions.
ranges, maxgap, minoverlap, type, invert	parameters used by subsetByOverlaps
object	An object of "Enhancers"

Value

An object of Enhancers.

Slots

genome An object of BSgenome.

peaks An object of GRanges.

TFBP An object of lgCMatrix.

TFBP0 An vector of logical.

Examples

```
Enhancers()
```

`getENCODEdata` *Download enhancer sequences from ENCODE*

Description

Query enhancer peak and extract sequences from ENCODE

Usage

```
getENCODEdata(
  genome,
  markers = "H3K4me1",
  window_size = 1000L,
  step = 50L,
  output = c("Enhancer", "raw_peaks"),
  ...
)
```

Arguments

`genome`	An object of BSgenome.
`markers`	Enhancer markers. Default 'H3K4me1'. For active enhancer, it can be set as c('H3K4me1', 'H3K27ac'). If markers is a 'GRanges' object, the function will skip the download step.
`window_size, step`	The size of windows and steps to split the peaks into small pieces. These parameter is used because the width of histone marker peaks are different sizes. Break the peaks into small pieces can increase the matching score and align the matching for different peaks into same size. The window_size is also be used for overlapping detection of multiple histone markers.
`output`	Output format. If it is 'raw_peaks', the raw peaks list will be returned. Otherwise, the Enhancer object will be returned.
`...`	Parameters can be passed to queryEncode

Value

An object of Enhancers with genome, and peaks. The peaks is an object of GRanges. The genome is an object of BSgenome.

Examples

```
library(BSgenome.Hsapiens.UCSC.hg38)
hs <- getENCODEdata(genome=Hsapiens,
                    partialMatch=c(biosample_summary="spinal cord"))
```

motifs *Pre-clustered motifs from human and mouse*

Description

The data were extracted from MotifDb package (v 1.34.0) and grouped by motifStack package (v 1.37.2). The data were packaged as PFMatrixList object by TFBSTools (v 1.30.0)

Usage

```
data(motifs)
```

Format

a list of PFMatrixList. The names of the list is the grouop distance.

Source

MotifDb package. Source code for the data generation is in extdata folder

Examples

```
data(motifs)
names(motifs)
motifs[[1]]
```

queryEncode *query data from ENCODE by predefined criteria*

Description

Search ENCODE data by querying the ENCODE Portal using its REST API.

Usage

```
queryEncode(
  exactMatch,
  partialMatch = character(0),
  API_url = "https://www.encodeproject.org/search/",
  ...
)
```

Arguments

exactMatch	character. Exact-match keywords refer to search results that perfectly match all the keywords in the search query, exactly as entered. It is a named character vector. The names will be the keys and characters will be the values for search.
partialMatch	character. Partial-match refer to search results that contain the search query. It is a named character vector. The names will be the keys and characters will be the values for search. The value starting from '!' indicates logical negation(NOT). The value starting from '>', '>=', '<', '==', '<=' indicates string comparison.
API_url	character. The ENCODE REST API url.
...	Not used.

Value

A list of search results

Examples

```
res <- queryEncode(
 exactMatch=c(
  target.label="H3K4me1",
  replicates.library.biosample.donor.organism.scientific_name="Homo sapiens",
  assembly="GRCh38",
  assay_term_name="ChIP-seq"),
  partialMatch=c(biosample_summary="heart"))
```

---

saveAlignments *output alignments*

Description

Save enhancer homologs to file in phylip format.

Usage

```
saveAlignments(
  al,
  output_folder = tempdir(),
  motifConsensus = NULL,
  format = c("txt", "html")
)
```

Arguments

al	output of alignment.
output_folder	output folder.
motifConsensus	Transcription factor binding consensus.
format	The format of output files. Available formats are 'txt' and 'html'. Default is 'txt'.

Value

The I/O status.

Examples

```
al <- readRDS(system.file("extdata", "al.rds",
              package="enhancerHomologSearch"))
tmpfolder <- tempdir()
library(MotifDb)
motifs <- query(MotifDb, "JASPAR_CORE")
consensus <- sapply(motifs, consensusString)
consensus <- DNAStringSet(gsub("\\?", "N", consensus))
saveAlignments(al, output_folder=tmpfolder, motifConsensus=consensus)
```

---

searchTFBPS *Transcription Factor Binding Pattern Similarity (TFBPS) search*

Description

Search the TFBPs for query in subject.

Usage

```
searchTFBPS(
  query,
  subject,
  PWMs,
  queryGenome,
  background = "genome",
  ...,
  maximalShuffleEnhancers = 1000
)
```

Arguments

query	An object of DNAStringSet to represent enhancer
subject	Output of getENCODEdata. An object of Enhancers
PWMs	The Position Weight Matrix list represented as a numeric matrix. Object of PWMatrixList or PFMatrixList.
queryGenome	An object of BSgenome for query data.
background	background nucleotide frequencies. Default is "genome". Refer matchMotifs for details.
...	Parameters will be passed to matchMotifs except 'out' and 'genome'.
maximalShuffleEnhancers	The maximal number of Shuffled enhancers. If the number of the input enhancer candidates is greater than maximalShuffleEnhancers, no shuffled enhancer sequences will be included. The shuffled enhancers will be created by shuffle.

Value

An object of Enhancers.

Examples

```
library(BSgenome.Hsapiens.UCSC.hg38)
peaks <- GRanges("chr1", IRanges(seq(5000, 50000, by=1000), width=1000))
peaks$id <- paste(seq_along(peaks), 1, sep="_")
subj <- Enhancers(genome=Hsapiens, peaks=peaks)
q <- getSeq(Hsapiens, GRanges("chr1", IRanges(90000, width=1000)))
data(motifs)
ao <- searchTFBPS(q, subj, motifs[["dist60"]], queryGenome=Hsapiens,
                  maximalShuffleEnhancers = 50)
```

---

shuffle *shuffle reads*

Description

Uses the uShuffle library to shuffle reads

Usage

```
shuffle(reads, k = 2, n = 2)
```

Arguments

reads	An object of BStringSet.
k	the k-let size.
n	the number of random sequences to generate.

Value

An object of BStringSet.

References

Jiang, M., Anderson, J., Gillespie, J. et al. uShuffle: A useful tool for shuffling biological sequences while preserving the k-let counts. BMC Bioinformatics 9, 192 (2008). https://doi.org/10.1186/1471-2105-9-192

Examples

```
library(Biostrings)
f <- DNAStringSet(c("CTC-NACCAGTAT", "TTGA", "TACCTAGAG"))
shuffle(f)
```

Index

∗ **datasets**
    motifs, 9
$,Enhancers-method *(*Enhancers-class*)*, 6
$<-,Enhancers-method *(*Enhancers-class*)*, 6

alignment, 2, *5*, *10*
alignmentOne, 4

BSgenome, *5*, *7, 8*, *11*
BStringSet, *12*

coerce *(*Enhancers-class*)*, 6
coerce,Enhancers,GRanges-method *(*Enhancers-class*)*, 6
conservedMotifs, 4

distance *(*Enhancers-class*)*, 6
distance,Enhancers,ANY-method *(*Enhancers-class*)*, 6
distance,Enhancers-method *(*Enhancers-class*)*, 6
distance<- *(*Enhancers-class*)*, 6
distance<-,Enhancers,ANY-method *(*Enhancers-class*)*, 6
distance<-,Enhancers-method *(*Enhancers-class*)*, 6

Enhancers, *3, 4*, *8*, *11, 12*
Enhancers *(*Enhancers-class*)*, 6
Enhancers-class, 6

genome *(*Enhancers-class*)*, 6
genome,Enhancers-method *(*Enhancers-class*)*, 6
genome<- *(*Enhancers-class*)*, 6
genome<-,Enhancers,BSgenome-method *(*Enhancers-class*)*, 6
genome<-,Enhancers-method *(*Enhancers-class*)*, 6
getENCODEdata, *6*, 8
getSeq,Enhancers-method *(*Enhancers-class*)*, 6
GRanges, *7*

lgCMatrix, *7*

matchMotifs, *5*, *11*
motifs, 9

peaks *(*Enhancers-class*)*, 6
peaks,Enhancers-method *(*Enhancers-class*)*, 6
peaks<- *(*Enhancers-class*)*, 6
peaks<-,Enhancers,GRanges-method *(*Enhancers-class*)*, 6
peaks<-,Enhancers-method *(*Enhancers-class*)*, 6
PFMatrixList, *5*, *11*
PWMatrixList, *5*, *11*

query_tfbp *(*Enhancers-class*)*, 6
query_tfbp,Enhancers,ANY-method *(*Enhancers-class*)*, 6
query_tfbp,Enhancers-method *(*Enhancers-class*)*, 6
queryEncode, *8*, 9

saveAlignments, 10
searchTFBPS, *5*, 11
seqinfo,Enhancers-method *(*Enhancers-class*)*, 6
show,Enhancers-method *(*Enhancers-class*)*, 6
shuffle, *11*, 12
subset,Enhancers-method *(*Enhancers-class*)*, 6
subsetByOverlaps, *7*
subsetByOverlaps,Enhancers,ANY-method *(*Enhancers-class*)*, 6
subsetByOverlpas,Enhancers-method *(*Enhancers-class*)*, 6

tfbp *(*Enhancers-class*)*, 6
tfbp,Enhancers,ANY-method
*(*Enhancers-class*)*, 6
tfbp,Enhancers-method
*(*Enhancers-class*)*, 6

XStringViews, *5*