Package 'imcRtools'

November 12, 2026

Version 1.18.1

Title Methods for imaging mass cytometry data analysis

Description This R package supports the handling and analysis of imaging mass cytometry and other highly multiplexed imaging data. The main functionality includes reading in single-cell data after image segmentation and measurement, data formatting to perform channel spillover correction and a number of spatial analysis approaches. First, cell-cell interactions are detected via spatial graph construction; these graphs can be visualized with cells representing nodes and interactions representing edges. Furthermore, per cell, its direct neighbours are summarized to allow spatial clustering. Per image/grouping level, interactions between types of cells are counted, averaged and compared against random permutations. In that way, types of cells that interact more (attraction) or less (avoidance) frequently than expected by chance are detected.

License GPL-3

Depends R (>= 4.1), SpatialExperiment

Imports S4Vectors, stats, utils, SummarizedExperiment, methods, pheatmap, scrapper, stringr, readr, EBImage, cytomapper, abind, BiocParallel, viridis, dplyr, magrittr, DT, igraph, SingleCellExperiment, vroom, BiocNeighbors, RTriangle, ggraph, tidygraph, ggplot2, data.table, sf, concaveman, tidyselect, distances, MatrixGenerics, rlang, grDevices

Suggests CATALYST, grid, tidyr, BiocStyle, knitr, rmarkdown, markdown, testthat

biocViews ImmunoOncology, SingleCell, Spatial, DataImport, Clustering

VignetteBuilder knitr

URL https://github.com/BodenmillerGroup/imcRtools

BugReports https://github.com/BodenmillerGroup/imcRtools/issues

RoxygenNote 7.3.3

Encoding UTF-8

git_url https://git.bioconductor.org/packages/imcRtools

git_branch RELEASE_3_23

git_last_commit 075e056

git_last_commit_date 2026-05-19

Repository Bioconductor 3.23

Date/Publication 2026-11-11

Author Nils Eling [aut],
Tobias Hoch [ctb],
Vito Zanotelli [ctb],
Jana Fischer [ctb],
Daniel Schulz [ctb, cre] (ORCID:
<https://orcid.org/0000-0002-0913-1678>),
Lasse Meyer [ctb],
Lutz Marlene [ctb],
Schiller Chiara [ctb],
Ibañez Victor [ctb]

Maintainer Daniel Schulz <daniel.schulz@uzh.ch>

Contents

aggregateNeighbors . . . 3
binAcrossPixels . . . 4
buildSpatialGraph . . . 6
countInteractions . . . 8
detectCommunity . . . 11
detectSpatialContext . . . 13
distToCells . . . 15
filterPixels . . . 17
filterSpatialContext . . . 18
findBorderCells . . . 20
patchDetection . . . 21
patchSize . . . 24
plotInteractions . . . 25
plotSpatial . . . 27
plotSpatialContext . . . 31
plotSpotHeatmap . . . 33
readImagefromTXT . . . 35
readSCEfromTIFF . . . 37
readSCEfromTXT . . . 38
read_cpout . . . 40
read_steinbock . . . 42
show_cpout_features . . . 45
testInteractions . . . 46

Index **50**

aggregateNeighbors *Function to aggregate all neighbors of each cell.*

Description

Function to summarize categorical or expression values of all neighbors of each cell.

Usage

```
aggregateNeighbors(
  object,
  colPairName,
  aggregate_by = c("metadata", "expression"),
  count_by = NULL,
  proportions = TRUE,
  assay_type = NULL,
  subset_row = NULL,
  statistic = c("mean", "median", "sd", "var"),
  name = NULL
)
```

Arguments

object	a SingleCellExperiment or SpatialExperiment object
colPairName	single character indicating the colPair(object) entry containing the neighbor information.
aggregate_by	character specifying whether the neighborhood should be summarized by cellular features stored in colData(object) (aggregate_by = "metdata") or by marker expression of the neighboring cells (aggregate_by = "expression").
count_by	for summarize_by = "metadata", a single character specifying the colData(object) entry containing the cellular metadata that should be summarized across each cell's neighborhood.
proportions	single logical indicating whether aggregated metadata should be returned in form of proportions instead of absolute counts.
assay_type	for summarize_by = "expression", single character indicating the assay slot to use.
subset_row	for summarize_by = "expression", an integer, logical or character vector specifying the features to use. If NULL, defaults to all features.
statistic	for summarize_by = "expression", a single character specifying the statistic to be used for summarizing the expression values across all neighboring cells. Supported entries are "mean", "median", "sd", "var". Defaults to "mean" if not specified.
name	single character specifying the name of the data frame to be saved in the colData(object). Defaults to "aggregatedNeighbors" when summarize_by = "metadata" or "statistic_aggregatedExpression" when summarize_by = "expression".

Value

returns an object of `class(object)` containing the aggregated values in form of a `DataFrame` object in `colData(object)[[name]]`.

Author(s)

Daniel Schulz (<daniel.schulz@uzh.ch>)

Examples

```
library(cytomapper)
data(pancreasSCE)

sce <- buildSpatialGraph(pancreasSCE, img_id = "ImageNb",
                         type = "knn", k = 3)

# Aggregating neighboring cell-types
sce <- aggregateNeighbors(sce, colPairName = "knn_interaction_graph",
                         aggregate_by = "metadata",
                         count_by = "CellType")
sce$aggregatedNeighbors

# Aggregating neighboring expression values
sce <- aggregateNeighbors(sce, colPairName = "knn_interaction_graph",
                         aggregate_by = "expression",
                         assay_type = "exprs",
                         statistic = "mean")
sce$mean_aggregatedExpression
```

---

binAcrossPixels *Aggregate consecutive pixels per single-metal spot*

Description

Helper function for estimating the spillover matrix. Per metal spot, consecutive pixels a aggregated (default: summed).

Usage

```
binAcrossPixels(
  object,
  bin_size,
  spot_id = "sample_id",
  assay_type = "counts",
  statistic = "sum",
  ...
)
```

Arguments

object	a SingleCellExperiment object containing pixel intensities for all channels. Individual pixels are stored as columns and channels are stored as rows.
bin_size	single numeric indicating how many consecutive pixels per spot should be aggregated.
spot_id	character string indicating which colData(object) entry stores the isotope names of the spotted metal.
assay_type	character string indicating which assay to use.
statistic	character string indicating the statistic to use for aggregating consecutive pixels.
...	additional arguments passed to aggregateAcrossCells

Value

returns the binned pixel intensities in form of a SingleCellExperiment object

Author(s)

Nils Eling (<nils.eling@dqbm.uzh.ch>)

See Also

aggregateAcrossCells for the aggregation function

Examples

```
path <- system.file("extdata/spillover", package = "imcRtools")
# Read in .txt files
sce <- readSCEfromTXT(path)
dim(sce)

# Visualizes heatmap before aggregation
plotSpotHeatmap(sce)

# Sum consecutive pixels
sce <- binAcrossPixels(sce, bin_size = 10)
dim(sce)

# Visualizes heatmap after aggregation
plotSpotHeatmap(sce)
```

buildSpatialGraph *Builds an interaction graph based on the cells' locations*

Description

Function to define cell-cell interactions via distance-based expansion, delaunay triangulation or k nearest neighbor detection.

Usage

```
buildSpatialGraph(
  object,
  img_id,
  type = c("expansion", "knn", "delaunay"),
  k = NULL,
  directed = TRUE,
  max_dist = NULL,
  threshold = NULL,
  coords = c("Pos_X", "Pos_Y"),
  name = NULL,
  BNPARAM = KmknnParam(),
  BPPARAM = SerialParam(),
  ...
)
```

Arguments

object	a SingleCellExperiment or SpatialExperiment object
img_id	single character indicating the colData(object) entry containing the unique image identifiers.
type	single character specifying the type of graph to be build. Supported entries are "expansion" (default) to find interacting cells via distance thresholding; "delaunay" to find interactions via delaunay triangulation; "knn" to find the k nearest neighboring cells.
k	(when type = "knn") single numeric integer defining the number of nearest neighbors to search for.
directed	(when type = "knn") should the returned graph be directed? (see details).
max_dist	(when type = "knn" or type = "delaunay") the maximum distance at which to consider neighboring cells. All neighbors within a distance larger than max_dist will be excluded from graph construction.
threshold	(when type = "expansion") single numeric specifying the maximum distance for considering neighbors
coords	character vector of length 2 specifying the names of the colData (for a SingleCellExperiment object) or the spatialCoords entries of the cells' x and y locations.
name	single character specifying the name of the graph.

BNPARAM	a BiocNeighborParam object defining the algorithm to use.
BPPARAM	a BiocParallelParam-class object defining how to parallelize computations.
...	additional parameters passed to the findNeighbors function (type = "expansion"), the triangulate function (type = "delaunay") or the findKNN function (type = "knn")).

Value

returns a SpatialExperiment or SingleCellExperiment containing the graph in form of a SelfHits object in colPair(object, name). The object is grouped by entries in the img_id slot.

Building an interaction graph

This function defines interacting cells in different ways. They are based on the cells' centroids and do not incorporate cell shape or area.

1. When type = "expansion", all cells within the radius threshold are considered interacting cells.

2. When type = "delaunay", interacting cells are found via a delaunay triangulation of the cells' centroids.

3. When type = "knn", interacting cells are defined as the k nearest neighbors in the 2D spatial plane.

The directed parameter only affects graph construction via k nearest neighbor search. For directed = FALSE, each interaction will be stored as mutual edge (e.g. node 2 is connected to node 10 and vise versa). For type = "expansion" and type = "delaunay", each edge is stored as mutual edge by default.

The graph is stored in form of a SelfHits object in colPair(object, name). This object can be regarded as an edgelist and coerced to an igraph object via graph_from_edgelist(as.matrix(colPair(object, name))).

Choosing the graph construction method

When finding interactions via expansion or knn, the findNeighbors or findKNN functions are used, respectively. Both functions accept the BNPARAM parameter via which the graph construction method can be defined (default KmknnParam). For an overview on the different algorithms, see BiocNeighborParam. Within the BiocNeighborParam object, distance can be set to "Euclidean" (default), "Manhattan" or "Cosine".

Ordering of the output object

The buildSpatialGraph function operates on individual images. Therefore the returned object is grouped by entries in img_id. This means all cells of a given image are grouped together in the object. The ordering of cells within each individual image is the same as the ordering of these cells in the input object.

Author(s)

Nils Eling (<nils.eling@dqbm.uzh.ch>)

See Also

findNeighbors for the function finding interactions via expansion

findKNN for the function finding interactions via nearest neighbor search

triangulate for the function finding interactions via delaunay triangulation

plotSpatial for visualizing spatial graphs

Examples

```
path <- system.file("extdata/mockData/steinbock", package = "imcRtools")
spe <- read_steinbock(path)

# Constructing a graph via expansion
spe <- buildSpatialGraph(spe, img_id = "sample_id",
                         type = "expansion", threshold = 10)
colPair(spe, "expansion_interaction_graph")

# Constructing a graph via delaunay triangulation
spe <- buildSpatialGraph(spe, img_id = "sample_id",
                         type = "delaunay")
colPair(spe, "delaunay_interaction_graph")

# Constructing a graph via k nearest neighbor search
spe <- buildSpatialGraph(spe, img_id = "sample_id",
                         type = "knn", k = 5)
colPair(spe, "knn_interaction_graph")
```

---

countInteractions *Summarizes cell-cell interactions within grouping levels (e.g. images)*

Description

Function to calculate the average number of neighbors B that a cell of type A has using different approaches.

Usage

```
countInteractions(
  object,
  group_by,
  label,
  colPairName,
  method = c("classic", "conditional", "patch", "interaction"),
  patch_size = NULL
)
```

Arguments

object	a SingleCellExperiment or SpatialExperiment object .
group_by	a single character indicating the colData(object) entry by which interactions are grouped. This is usually the image ID or patient ID.
label	single character specifying the colData(object) entry which stores the cell labels. These can be cell-types labels or other metadata.
colPairName	single character indicating the colPair(object) entry containing cell-cell interactions in form of an edge list.
method	which cell-cell interaction counting method to use (see details)
patch_size	if method = "patch", a single numeric specifying the minimum number of neighbors of the same type to be considered a patch (see details)

Value

a DataFrame containing one row per group_by entry and unique label entry combination (from_label, to_label). The ct entry stores the interaction count as described in the details. NA is returned if a certain label is not present in this grouping level.

Counting and summarizing cell-cell interactions

In principle, the countInteractions function counts the number of edges (interactions) between each set of unique entries in colData(object)[[label]]. Simplified, it counts for each cell of type A the number of neighbors of type B. This count is averaged within each unique entry colData(object)[[group_by]] in four different ways:

1. method = "classic": The count is divided by the total number of cells of type A. The final count can be interpreted as "How many neighbors of type B does a cell of type A have on average?"

2. method = "conditional": Formerly named "histocat". The count is divided by the number of cells of type A that have at least one neighbor of type B. The final count can be interpreted as "How many neighbors of type B has a cell of type A on average, given it has at least one neighbor of type B?".

3. method = "patch": For each cell, the count is binarized to 0 (less than patch_size neighbors of type B) or 1 (more or equal to patch_size neighbors of type B). The binarized counts are averaged across all cells of type A. The final count can be interpreted as "What fraction of cells of type A have at least a given number of neighbors of type B?"

4. method = "interaction": The count is divided by the total number of interactions from cell type A. The final count can be interpreted as "What fraction of interactions of cell type A occur with cell type B."

Author(s)

Vito Zanotelli

Jana Fischer

adapted by Nils Eling (<nils.eling@dqbm.uzh.ch>)

adapted by Marlene Lutz (<marlene.lutz@uzh.ch>)

References

Schulz, D. et al., Simultaneous Multiplexed Imaging of mRNA and Proteins with Subcellular Resolution in Breast Cancer Tissue Samples by Mass Cytometry., Cell Systems 2018 6(1):25-36.e5

Schapiro, D. et al., histoCAT: analysis of cell phenotypes and interactions in multiplex image cytometry data, Nature Methods 2017 14, p. 873–876

See Also

testInteractions for testing cell-cell interactions per grouping level.

Examples

```
library(cytomapper)
data(pancreasSCE)

pancreasSCE <- buildSpatialGraph(pancreasSCE, img_id = "ImageNb",
                                 type = "knn", k = 3)

# Classic style calculation
(out <- countInteractions(pancreasSCE,
                                group_by = "ImageNb",
                                label = "CellType",
                                method = "classic",
                                colPairName = "knn_interaction_graph"))

# Conditional style calculation
(out <- countInteractions(pancreasSCE,
                                group_by = "ImageNb",
                                label = "CellType",
                                method = "conditional",
                                colPairName = "knn_interaction_graph"))

# Patch style calculation
(out <- countInteractions(pancreasSCE,
                                group_by = "ImageNb",
                                label = "CellType",
                                method = "patch",
                                patch_size = 3,
                                colPairName = "knn_interaction_graph"))

# Interaction style calculation
(out <- countInteractions(pancreasSCE,
                                group_by = "ImageNb",
                                label = "CellType",
                                method = "interaction",
                                colPairName = "knn_interaction_graph"))
```

`detectCommunity` *Detect the spatial community of each cell*

Description

Function to detect the spatial community of each cell as proposed by Jackson et al., The single-cell pathology landscape of breast cancer, Nature, 2020. Each cell is clustered based on its interactions as defined by a spatial object graph.

Usage

```
detectCommunity(
  object,
  colPairName,
  size_threshold = 0,
  group_by = NULL,
  name = "spatial_community",
  cluster_fun = "louvain",
  BPPARAM = SerialParam()
)
```

Arguments

`object`	a `SingleCellExperiment` or `SpatialExperiment` object
`colPairName`	single character indicating the `colPair(object)` entry containing the neighbor information.
`size_threshold`	single positive numeric that specifies the minimum number of cells per community. Defaults to 0.
`group_by`	single character indicating that spatial community detection will be performed separately for all unique entries to `colData(object)[,group_by]`.
`name`	single character specifying the name of the output saved in `colData(object)`. Defaults to "spatial_community".
`cluster_fun`	single character specifying the function to use for community detection. Options are all strings that contain the suffix of an `igraph` community detection algorithm (e.g. "walktrap"). Defaults to "louvain".
`BPPARAM`	a `BiocParallelParam-class` object defining how to parallelize computations. Applicable when `group_by` is specified and defaults to `SerialParam()`. For reproducibility between runs, we recommend defining `RNGseed` in the `BiocParallelParam-class` object.

Value

returns an object of `class(object)` containing a new column entry to `colData(object)[[name]]`.

Spatial community detection procedure

1. Create an igraph object from the edge list stored in `colPair(object, colPairName)`.
2. Perform community detection using the specified `cluster_fun` algorithm.
3. Store the community IDs in a vector and replace all communities with a size smaller than `size_threshold` by NA.

Optional steps: Specify `group_by` to perform spatial community detection separately for all unique entries to `colData(object)[,group_by]` e.g. for tumor and non-tumor cells.

Author(s)

Lasse Meyer (<lasse.meyer@uzh.ch>)

References

Jackson et al., The single-cell pathology landscape of breast cancer, Nature, 2020

Examples

```
library(cytomapper)
library(BiocParallel)
data(pancreasSCE)

sce <- buildSpatialGraph(pancreasSCE, img_id = "ImageNb",
                         type = "expansion",
                         name = "neighborhood",
                         threshold = 20)

## Detect spatial community
set.seed(22)
sce <- detectCommunity(sce,
                       colPairName = "neighborhood",
                       size_threshold = 10)

plotSpatial(sce,
            img_id = "ImageNb",
            node_color_by = "spatial_community",
            scales = "free")

## Detect spatial community - specify group_by
sce <- detectCommunity(sce,
                        colPairName = "neighborhood",
                        group_by = "CellType",
                        size_threshold = 10,
                        BPPARAM = SerialParam(RNGseed = 22))

plotSpatial(sce,
            img_id = "ImageNb",
            node_color_by = "spatial_community",
            scales = "free")
```

`detectSpatialContext` *Detect the spatial context of each cell based on its neighborhood*

Description

Function to detect the spatial context (SC) of each cell. Based on its sorted (high-to-low) cellular neighborhood (CN) fractions in a spatial interaction graph, the SC of each cell is assigned as the set of CNs that cumulatively exceed a user-defined fraction threshold.

The term was coined by Bhate S. et al., Tissue schematics map the specialization of immune tissue motifs and their appropriation by tumors, Cell Systems, 2022 and describes tissue regions in which distinct CNs may be interacting.

Usage

```
detectSpatialContext(
  object,
  entry = "aggregatedNeighbors",
  threshold = 0.9,
  name = "spatial_context"
)
```

Arguments

`object`	a `SingleCellExperiment` or `SpatialExperiment` object
`entry`	single character specifying the `colData(object)` entry containing the `aggregateNeighbors` DataFrame output. Defaults to "aggregatedNeighbors".
`threshold`	single numeric between 0 and 1 that specifies the fraction threshold for SC assignment. Defaults to 0.9.
`name`	single character specifying the name of the output saved in `colData(object)`. Defaults to "spatial_context".

Value

returns an object of `class(object)` containing a new column entry to `colData(object)[[name]]`

Spatial context background

The function relies on CN fractions for each cell in a spatial interaction graph (originally a k-nearest neighbor (KNN) graph).

We can retrieve the CN fractions using the buildSpatialGraph and aggregateNeighbors functions.

The window size (k for KNN) for buildSpatialGraph should reflect a length scale on which biological signals can be exchanged and depends, among others, on cell density and tissue area. In view of their divergent functionality, we recommend to use a larger window size for SC (interaction between local processes) than for CN (local processes) detection.

Subsequently, the CN fractions are sorted from high-to-low and the SC of each cell is assigned the minimal combination of SCs that additively surpass a user-defined threshold. The default threshold of 0.9 aims to represent the dominant CNs, hence the most prevalent signals, in a given window.

For more details, please refer to: Bhate S. et al., Tissue schematics map the specialization of immune tissue motifs and their appropriation by tumors, Cell Systems, 2022.

Author(s)

Lasse Meyer (<lasse.meyer@uzh.ch>)

References

Bhate S. et al., Tissue schematics map the specialization of immune tissue motifs and their appropriation by tumors, Cell Systems, 2022

See Also

`filterSpatialContext` for the function to filter spatial contexts

`plotSpatialContext` for the function to plot spatial context graphs

Examples

```
set.seed(22)
library(cytomapper)
data(pancreasSCE)

## 1. Cellular neighborhood (CN)
sce <- buildSpatialGraph(pancreasSCE, img_id = "ImageNb",
                         type = "knn",
                         name = "knn_cn_graph",
                         k = 5)

sce <- aggregateNeighbors(sce, colPairName = "knn_cn_graph",
                          aggregate_by = "metadata",
                          count_by = "CellType",
                          name = "aggregatedCellTypes")

cur_cluster <- kmeans(sce$aggregatedCellTypes, centers = 3)
sce$cellular_neighborhood <- factor(cur_cluster$cluster)

plotSpatial(sce, img_id = "ImageNb",
            colPairName = "knn_cn_graph",
            node_color_by = "cellular_neighborhood",
            scales = "free")

## 2. Spatial context (SC)
sce <- buildSpatialGraph(sce, img_id = "ImageNb",
                         type = "knn",
                         name = "knn_sc_graph",
                         k = 15)
```

```
sce <- aggregateNeighbors(sce, colPairName = "knn_sc_graph",
                          aggregate_by = "metadata",
                          count_by = "cellular_neighborhood",
                          name = "aggregatedNeighborhood")

# Detect spatial context
sce <- detectSpatialContext(sce, entry = "aggregatedNeighborhood",
                            threshold = 0.9)

plotSpatial(sce, img_id = "ImageNb",
            colPairName = "knn_sc_graph",
            node_color_by = "spatial_context",
            scales = "free")
```

`distToCells` *Function to calculate distance to cells of interest*

Description

Function to return the min, max, mean or median distance to the cells of interest for each cell in the data. In the case of patched/clustered cells negative distances are returned by default which indicate the distance of the cells of interest to the cells that are not of the type of cells of interest.

Usage

```
distToCells(
  object,
  x_cells,
  img_id,
  name = "distToCells",
  coords = c("Pos_X", "Pos_Y"),
  statistics = "min",
  return_neg = TRUE,
  BPPARAM = SerialParam()
)
```

Arguments

`object`	a `SingleCellExperiment` or `SpatialExperiment` object
`x_cells`	logical vector of length equal to the number of cells contained in `object`. `TRUE` entries define the cells to which distances will be calculated.
`img_id`	single character indicating the `colData(object)` entry containing the unique image identifiers.
`name`	character specifying the name of the `colData` entry to safe the distances in.
`coords`	character vector of length 2 specifying the names of the `colData` (for a `SingleCellExperiment` object) or the `spatialCoords` entries of the cells' x and y locations.

statistics	one of "min", "max", "mean" or "meadian" specifying the distance statistics to use when computing the distances.
return_neg	logical indicating whether negative distances are to be returned for the distances of patched/spatially clustered cells.
BPPARAM	a BiocParallelParam-class object defining how to parallelize computations.

Value

returns an object of class(object) containing a new column entry to colData(object)[[name]]. Cells in the object are grouped by entries in img_id.

Ordering of the output object

The minDistToCells function operates on individual images. Therefore the returned object is grouped by entries in img_id. This means all cells of a given image are grouped together in the object. The ordering of cells within each individual image is the same as the ordering of these cells in the input object.

Author(s)

Daniel Schulz & Bruno Palau (<daniel.schulz@uzh.ch>)

Examples

```
library(cytomapper)
data(pancreasSCE)

# Build interaction graph
pancreasSCE <- buildSpatialGraph(pancreasSCE, img_id = "ImageNb",
type = "expansion",threshold = 20)

# Detect patches of "celltype_B" cells
pancreasSCE <- patchDetection(pancreasSCE,
                              img_id = "ImageNb",
                              patch_cells = pancreasSCE$CellType == "celltype_B",
                              colPairName = "expansion_interaction_graph",
                              min_patch_size = 20,
                              expand_by = 1)

plotSpatial(pancreasSCE,
            img_id = "ImageNb",
            node_color_by = "patch_id",
            scales = "free")

# Distance to celltype_B patches
pancreasSCE <- distToCells(pancreasSCE,
                              x_cells = !is.na(pancreasSCE$patch_id),
                              coords = c("Pos_X","Pos_Y"),
                              statistics = "min",
                              img_id = "ImageNb")
```

```
plotSpatial(pancreasSCE,
            img_id = "ImageNb",
            node_color_by = "distToCells",
            scales = "free")
```

`filterPixels` *Filter pixels based on their assigned masses*

Description

Helper function for estimating the spillover matrix. After assigning each pixel to a spotted mass, this function will filter incorrectly assigned pixels and remove small pixel sets.

Usage

```
filterPixels(
  object,
  bc_id = "bc_id",
  spot_mass = "sample_mass",
  minevents = 40,
  correct_pixels = TRUE
)
```

Arguments

`object`	a `SingleCellExperiment` object containing pixel intensities per channel. Individual pixels are stored as columns and channels are stored as rows.
`bc_id`	character string indicating which `colData(object)` entry stores the estimated mass
`spot_mass`	character string indicating which `colData(object)` entry stores the true isotope mass of the spotted metal.
`minevents`	single numeric indicating the threshold under which pixel sets are excluded from spillover estimation.
`correct_pixels`	logical indicating if incorrectly assigned pixels should be excluded from spillover estimation.

Value

returns a SingleCellExperiment object in which `colData(object)$bc_id` has been adjusted based on the filter criteria.

Author(s)

Vito Zanotelli, adapted by Nils Eling (<nils.eling@dqbm.uzh.ch>)

Examples

```
path <- system.file("extdata/spillover", package = "imcRtools")
sce <- readSCEfromTXT(path)
assay(sce, "exprs") <- asinh(counts(sce)/5)

# Pre-process via CATALYST
library(CATALYST)

bc_key <- as.numeric(unique(sce$sample_mass))
sce <- assignPrelim(sce, bc_key = bc_key)
sce <- estCutoffs(sce)
sce <- applyCutoffs(sce)

sce <- filterPixels(sce)

table(sce$sample_mass, sce$bc_id)
```

---

filterSpatialContext *Filter spatial contexts*

Description

Function to filter detected spatial contexts (SCs) based on a user-defined threshold for number of group entries and/or cells.

Usage

```
filterSpatialContext(
  object,
  entry = "spatial_context",
  group_by = "sample_id",
  group_threshold = NULL,
  cells_threshold = NULL,
  name = "spatial_context_filtered"
)
```

Arguments

object	a SingleCellExperiment or SpatialExperiment object
entry	a single character specifying the colData(object) entry containing the detectSpatialContext output. Defaults to "spatial_context".
group_by	a single character indicating the colData(object) entry by which SCs are grouped. This is usually the image or patient ID. Defaults to "sample_id".
group_threshold	a single numeric specifying the minimum number of group entries in which a SC is detected.

`cells_threshold`
: a single numeric specifying the minimum total number of cells in a SC.

`name`
: a single character specifying the name of the output saved in `colData(object)`. Defaults to "spatial_context_filtered".

Value

returns an object of `class(object)` containing a new column entry to `colData(object)[[name]]` and a new `data.frame` entry to `metadata(object)[["filterSpatialContext"]]` containing the group and cell counts per SC.

Author(s)

Lasse Meyer (<lasse.meyer@uzh.ch>)

References

Bhate S. et al., Tissue schematics map the specialization of immune tissue motifs and their appropriation by tumors, Cell Systems, 2022

See Also

`detectSpatialContext` for the function to detect spatial contexts

`plotSpatialContext` for the function to plot spatial context graphs

Examples

```
set.seed(22)
library(cytomapper)
data(pancreasSCE)

## 1. Cellular neighborhood (CN)
sce <- buildSpatialGraph(pancreasSCE, img_id = "ImageNb",
                         type = "knn",
                         name = "knn_cn_graph",
                         k = 5)

sce <- aggregateNeighbors(sce, colPairName = "knn_cn_graph",
                          aggregate_by = "metadata",
                          count_by = "CellType",
                          name = "aggregatedCellTypes")

cur_cluster <- kmeans(sce$aggregatedCellTypes, centers = 3)
sce$cellular_neighborhood <- factor(cur_cluster$cluster)

plotSpatial(sce, img_id = "ImageNb",
            colPairName = "knn_cn_graph",
            node_color_by = "cellular_neighborhood",
            scales = "free")

## 2. Spatial context (SC)
```

```
sce <- buildSpatialGraph(sce, img_id = "ImageNb",
                         type = "knn",
                         name = "knn_sc_graph",
                         k = 15)

sce <- aggregateNeighbors(sce, colPairName = "knn_sc_graph",
                          aggregate_by = "metadata",
                          count_by = "cellular_neighborhood",
                          name = "aggregatedNeighborhood")

# Detect spatial context
sce <- detectSpatialContext(sce, entry = "aggregatedNeighborhood",
                            threshold = 0.9)

plotSpatial(sce, img_id = "ImageNb",
            colPairName = "knn_sc_graph",
            node_color_by = "spatial_context",
            scales = "free")

# Filter spatial context
# By group
sce <- filterSpatialContext(sce, group_by = "ImageNb",
                            group_threshold = 2)

plotSpatial(sce, img_id = "ImageNb",
             colPairName = "knn_sc_graph",
             node_color_by = "spatial_context_filtered",
             scales = "free")

# By cells
sce <- filterSpatialContext(sce, group_by = "ImageNb",
                            cells_threshold = 15)

plotSpatial(sce, img_id = "ImageNb",
            colPairName = "knn_sc_graph",
            node_color_by = "spatial_context_filtered",
            scales = "free")
```

`findBorderCells` *Find cells at the image border*

Description

Detection of cells close to the image border for subsequent exclusion from downstream analyses.

Usage

```
findBorderCells(object, img_id, border_dist, coords = c("Pos_X", "Pos_Y"))
```

Arguments

object	a SingleCellExperiment or SpatialExperiment object.
img_id	single character indicating the colData(object) entry containing the unique image identifiers.
border_dist	single numeric defining the distance to the image border. The image border here is defined as the minimum and maximum among the cells' x and y location.
coords	character vector of length 2 specifying the names of the colData (for a SingleCellExperiment object) or the spatialCoords entries indicating the cells' x and y locations.

Value

an object of class(object) containing the logical border_cells entry in the colData slot.

Examples

```
library(cytomapper)
data("pancreasSCE")

sce <- findBorderCells(pancreasSCE, img_id = "ImageNb",
                       border_dist = 10)

plotSpatial(sce,
            img_id = "ImageNb",
            node_color_by = "border_cells",
            scales = "free")
```

---

patchDetection *Function to detect patches containing defined cell types*

Description

Function to detect spatial clusters of defined types of cells. By defining a certain distance threshold, all cells within the vicinity of these clusters are detected as well.

Usage

```
patchDetection(
  object,
  patch_cells,
  colPairName,
  min_patch_size = 1,
  name = "patch_id",
  expand_by = 0,
  coords = c("Pos_X", "Pos_Y"),
  convex = FALSE,
  img_id = NULL,
  BPPARAM = SerialParam()
)
```

Arguments

object	a SingleCellExperiment or SpatialExperiment object
patch_cells	logical vector of length equal to the number of cells contained in object. TRUE entries define the cells to consider for patch detection (see Details).
colPairName	single character indicating the colPair(object) entry containing the neighbor information.
min_patch_size	single integer indicating the minimum number of connected cells that make up a patch before expansion.
name	single character specifying the colData entry storing the patch IDs in the returned object.
expand_by	single numeric indicating in which vicinity range cells should be considered as belonging to the patch (see Details).
coords	character vector of length 2 specifying the names of the colData (for a SingleCellExperiment object) or the spatialCoords entries of the cells' x and y locations.
convex	should the convex hull be computed before expansion? Default: the concave hull is computed.
img_id	single character indicating the colData(object) entry containing the unique image identifiers.
BPPARAM	a BiocParallelParam-class object defining how to parallelize computations.

Value

An object of class(object) containing a patch ID for each cell in colData(object)[[name]]. If expand_by > 0, cells in the output object are grouped by entries in img_id.

Detecting patches of defined cell types

This function works as follows:

1. Only cells defined by patch_cells are considered for patch detection.

2. Patches of connected cells are detected. Here, cell-to-cell connections are defined by the interaction graph stored in colPair(object, colPairName). At this point, patches that contain fewer than min_patch_size cells are removed.

3. If expand_by > 0, a concave (default) or convex hull is constructed around each patch. This is is then expanded by expand_by and cells within the expanded hull are detected and assigned to the patch. This expansion only works if a patch contains at least 3 cells.

The returned object contains an additional entry colData(object)[[name]], which stores the patch ID per cell. NA indicate cells that are not part of a patch.

Ordering of the output object

If expand_by > 0, the patchDetection function operates on individual images. Therefore the returned object is grouped by entries in img_id. This means all cells of a given image are grouped together in the object. The ordering of cells within each individual image is the same as the ordering of these cells in the input object.

If expand_by = 0, the ordering of cells in the output object is the same as in the input object.

Author(s)

Tobias Hoch

adapted by Nils Eling (<nils.eling@dqbm.uzh.ch>)

References

Hoch, T. et al., Multiplexed Imaging Mass Cytometry of Chemokine Milieus in Metastatic Melanoma Characterizes Features of Response to Immunotherapy., bioRxiv 2021

Examples

```
library(cytomapper)
data(pancreasSCE)

# Visualize cell types
plotSpatial(pancreasSCE,
            img_id = "ImageNb",
            node_color_by = "CellType",
            scales = "free")

# Build interaction graph
pancreasSCE <- buildSpatialGraph(pancreasSCE, img_id = "ImageNb",
                                 type = "expansion", threshold = 20)

# Detect patches of "celltype_B" cells
pancreasSCE <- patchDetection(pancreasSCE,
                              patch_cells = pancreasSCE$CellType == "celltype_B",
                              colPairName = "expansion_interaction_graph")

plotSpatial(pancreasSCE,
            img_id = "ImageNb",
            node_color_by = "patch_id",
            scales = "free")

# Include cells in vicinity
pancreasSCE <- patchDetection(pancreasSCE,
                              patch_cells = pancreasSCE$CellType == "celltype_B",
                              colPairName = "expansion_interaction_graph",
                              expand_by = 20,
                              img_id = "ImageNb")

plotSpatial(pancreasSCE,
            img_id = "ImageNb",
            node_color_by = "patch_id",
            scales = "free")
```

patchSize *Function to compute the area of c3ll patches*

Description

This function constructs polygons around patch cells and computes their area.

Usage

```
patchSize(
  object,
  patch_name = "patch_id",
  coords = c("Pos_X", "Pos_Y"),
  convex = FALSE
)
```

Arguments

object	a SingleCellExperiment or SpatialExperiment object
patch_name	single character indicating the colData(object) entry containing the patch cell identifiers.
coords	character vector of length 2 specifying the names of the colData (for a SingleCellExperiment object) or the spatialCoords entries of the cells' x and y locations.
convex	should the convex hull be computed to construct the polygon? Default: the concave hull is computed.

Value

A DataFrame object containing the patch identifier, the constructed polygon and the polygon size.

Author(s)

Nils Eling (<nils.eling@dqbm.uzh.ch>)

Examples

```
library(cytomapper)
data(pancreasSCE)

# Build interaction graph
pancreasSCE <- buildSpatialGraph(pancreasSCE, img_id = "ImageNb",
                                 type = "expansion", threshold = 20)

# Detect patches of "celltype_B" cells
pancreasSCE <- patchDetection(pancreasSCE,
                              patch_cells = pancreasSCE$CellType == "celltype_B",
                              expand_by = 5, img_id = "ImageNb",
                              colPairName = "expansion_interaction_graph")
```

```
# Compute the patch area
patchSize(pancreasSCE)
```

`plotInteractions` *Plot interaction graph*

Description

Function to plot directed interaction graphs based on symbolic edge-lists and vertex metadata. The user can specify node, node_label and edge aesthetics using dedicated arguments. The resulting plot can be further refined with ‘ggplot2‘ for node styling and ‘ggraph‘ for edge-specific customization.

Usage

```
plotInteractions(
  out,
  object,
  label,
  group_by,
  node_color_by = NULL,
  node_size_by = NULL,
  node_color_fix = NULL,
  node_size_fix = NULL,
  node_label_repel = TRUE,
  node_label_color_by = NULL,
  node_label_color_fix = NULL,
  edge_color_by = NULL,
  edge_color_fix = NULL,
  edge_width_by = NULL,
  edge_width_fix = NULL,
  draw_edges = TRUE,
  return_data = FALSE,
  graph_layout = "circle"
)
```

Arguments

`out`	a data frame, usually the output from `countInteractions` or `testInteractions`, representing an edge list with columns `"group_by"`, `"from_label"` and `"to_label"`. Additional columns may be included to specify edge attributes (weight or color).
`object`	a `SingleCellExperiment` or `SpatialExperiment` object.
`label`	single character specifying the `colData(object)` entry which stores the cell labels. These can be cell-types labels or other metadata entries.

group_by — a single character indicating the colData(object) entry by which interactions are grouped. This is usually the image or patient ID. a single character indicating the colData(object)

node_color_by — single character either NULL, "name","n_cells", "n_group" by which the nodes should be colored.

node_size_by — single character either NULL, "n_cells","n_group" by which the size of the nodes are defined.

node_color_fix — single character specifying the color of all nodes.

node_size_fix — single numeric specifying the size of all nodes.

node_label_repel — should nodes be labelled? Defaults to TRUE.

node_label_color_by — single character either NULL, "name","n_cells","n_group" by which the node labels should be colored.

node_label_color_fix — single character specifying the color of all node labels.

edge_color_by — single character indicating the name of the column of "out" used represent edge colors. This column is usually newly added by the user and must assign a unique value to each 'from_label'–'to_label' pair. Typically, these values could encode the direction of significantly interacting cell type pairs.

edge_color_fix — single character specifying the color of all edges.

edge_width_by — single character indicating the name of the column of "out" used to scale edge widths. The values in this column are averaged for each 'from_label'–'to_label' pair. Typically, this could be the 'ct' column from of "out" or a newly added column representing an interaction feature.

edge_width_fix — single numeric specifying the width of all edges.

draw_edges — should edges be drawn between nodes? Defaults to TRUE.

return_data — should the edge list and vertex metadata for graph construction be returned as a list of two data.frames?

graph_layout — single character of "circle", "chord", "linear", "fr", "kk", "drl", "stress", "graphopt","lgl", "tree", "sugiyama", "star", "nicely", "manual", "grid", "mds", "sphere","randomly", "gem", "dh" which defines the graph layout. Defaults to "circle". For more information, see ggraph.

Value

returns a ggplot object or a list of two data.frames.

Author(s)

Marlene Lutz (<marlene.lutz@uzh.ch>)

See Also

countInteractions for counting (but not testing) cell-cell interactions per grouping level. testInteractions for testing cell-cell interactions per grouping level.

Examples

```
set.seed(22)
library(cytomapper)
library(BiocParallel)
data(pancreasSCE)

## 1. countInteractions or testInteractions
sce  <- buildSpatialGraph(pancreasSCE, img_id = "ImageNb", type = "knn", k = 3)

count_out <- countInteractions(sce,
                               group_by = "ImageNb",
                               label = "CellType",
                    method = "classic", # choose from c("classic", "histocat", "patch", "interaction")
                               colPairName = "knn_interaction_graph")

test_out <- testInteractions(sce,
                             group_by = "ImageNb",
                             label = "CellType",
                   method = "classic", # choose from c("classic", "histocat", "patch", "interaction")
                             colPairName = "knn_interaction_graph",
                             iter = 100,
                             p_threshold = 0.5,
                             BPPARAM = SerialParam(RNGseed = 123))

## 2. Plot interactions

# default
plotInteractions(count_out, sce, "CellType", "ImageNb")

# adjust node aesthetics
plotInteractions(count_out, sce, "CellType", "ImageNb",
                 node_color_by = "name",
                 node_size_by = "n_cells")

# adjust edge aesthetics
plotInteractions(test_out, sce, "CellType", "ImageNb",
                 edge_width_by = "ct")

# Plot interactions - return data
plotInteractions(test_out, sce, "CellType", "ImageNb",
                 return_data = TRUE)
```

`plotSpatial` *Visualizes the spatial locations and interactions of cells*

Description

A general function to plot spatial locations of cells while specifying color, shape, size. Cell-cell interactions can be visualized in form of edges between points.

Usage

```
plotSpatial(
  object,
  img_id,
  coords = c("Pos_X", "Pos_Y"),
  node_color_by = NULL,
  node_shape_by = NULL,
  node_size_by = NULL,
  node_color_fix = NULL,
  node_shape_fix = NULL,
  node_size_fix = NULL,
  assay_type = NULL,
  draw_edges = FALSE,
  directed = TRUE,
  edge_color_by = NULL,
  edge_width_by = NULL,
  edge_color_fix = NULL,
  edge_width_fix = NULL,
  arrow = NULL,
  end_cap = NULL,
  colPairName = NULL,
  nodes_first = TRUE,
  ncols = NULL,
  nrows = NULL,
  scales = "fixed",
  flip_x = FALSE,
  flip_y = TRUE,
  aspect_ratio = "auto"
)
```

Arguments

object	a SingleCellExperiment or SpatialExperiment object.
img_id	single character indicating the colData(object) entry containing the unique image identifiers.
coords	character vector of length 2 specifying the names of the colData (for a SingleCellExperiment object) or the spatialCoords entries indicating the the cells' x and y locations.
node_color_by	single character indicating the colData(object) entry or marker name by which the nodes (cell locations) should be colored.
node_shape_by	single character indicating the colData(object) entry by which the shape of the nodes are defined.
node_size_by	single character indicating the colData(object) entry by which the size of the nodes are defined.
node_color_fix	single character or numeric specifying the color of all nodes.
node_shape_fix	single numeric or character specifying the shape of all nodes.
node_size_fix	single numeric specifying the size of all nodes

assay_type	single character indicating the assay slot from which to extract the expression data when node_color_by is set to one of rownames(object).
draw_edges	should cell-cell interactions be drawn as edges between nodes?
directed	should cell-cell interactions be handled as a directed graph?
edge_color_by	single character indicating by which to color the edges. See details for more information.
edge_width_by	single character determining the size of the edges. See details for more information.
edge_color_fix	single character or numeric specifying the color of all edges.
edge_width_fix	single numeric specifying the size of all edges.
arrow	an arrow object specifying how to draw arrows between cells.
end_cap	a geometry object specifying how long the edges are. This only takes effect when drawing arrows. Default: end_cap = circle(0.1, 'cm')
colPairName	single character specifying the colPair(object) slot to retrieve the cell-cell pairings.
nodes_first	should the nodes be plotted first and then the edges?
ncols	number of columns of the grid to arrange individual images.
nrows	number of rows of the grid to arrange individual images.
scales	one of "free", "fixed", "free_x" or "free_y" indicating if x- and y-axis ranges should be fixed across all images. Defaults to "fixed" to match physical units on the x- and y-axis.
flip_x	flip the x-axis?
flip_y	flip the y-axis?
aspect_ratio	single numeric, "auto" or NULL to define the relative ratio between the physical units of the x and y axis. If "auto" (default), the physical units match between the x and y axis if scales = "fixed". If scales = "free", the default aspect ratio is set to 1. Ignore setting the aspect ratio with aspect_ratio = NULL.

Value

returns a ggplot object.

Visualizing cell locations and cell-cell interactions

By default, the cells' locations are visualized in form of points (here also referred to as "nodes") on a 2-dimensional plane. The cells' coordinates are extracted either from colData(object) slot (for a SingleCellExperiment input object) or from the spatialCoords(object) slot (for a SpatialExperiment input object). Node aesthetics are controlled by setting node_color_by, node_shape_by and node_size_by for associating the aesthetics with variables. If node aesthetics should be the same for all nodes, node_color_fix, node_shape_fix and node_size_fix can be set.

When draw_edges = TRUE, cell-cell interactions are visualized in form of edges between nodes. For this, object needs to contain column pairings in colPair(object, colPairName). Edge color and size can be set by specifying either an entry in mcols(colPair(object, colPairName)) (edge

attributes) or in colData(object). In the latter case, edges are colored by attributes associated to the "from" node. Variable aesthetics can be set using edge_color_by and edge_width_by. If all edges should have the same width or color, edge_color_fix and edge_width_fix can be set.

Arrows for displaying directed graphs can be drawn by supplying a arrow object. Arrow attributes can be set within this class. To cap the edge before it reaches the next node, the end_cap parameter can be used.

Author(s)

Nils Eling (<nils.eling@dqbm.uzh.ch>)

See Also

buildSpatialGraph for constructing interaction graphs

ggraph for handling graph aesthetics

Examples

```
library(cytomapper)
data(pancreasSCE)

sce <- buildSpatialGraph(pancreasSCE, img_id = "ImageNb",
                         type = "knn", k = 3, directed = FALSE)

# Only nodes
plotSpatial(sce, img_id = "ImageNb",
            node_color_by = "CellType",
            node_shape_by = "ImageNb",
            node_size_by = "Area",
            scales = "free")

# With edges and nodes colored by expression
plotSpatial(sce, img_id = "ImageNb",
            node_color_by = "PIN",
            assay_type = "exprs",
            node_shape_by = "ImageNb",
            node_size_by = "Area",
            draw_edges = TRUE,
            colPairName = "knn_interaction_graph",
            edge_color_by = "Pattern",
            scales = "free")

# With arrows
plotSpatial(sce, img_id = "ImageNb",
            node_color_by = "CellType",
            node_shape_by = "ImageNb",
            node_size_by = "Area",
            draw_edges = TRUE,
            colPairName = "knn_interaction_graph",
            edge_color_fix = "green",
            arrow = grid::arrow(length = grid::unit(0.1, "inch")),
```

```
              end_cap = ggraph::circle(0.2, "cm"),
              scales = "free")
```

---

plotSpatialContext *Plot spatial context graph*

Description

Function to plot directed spatial context graphs based on symbolic edge-lists and vertex metadata, which operates on the cohort-level. The user can specify node, node_label and edge aesthetics.

Usage

```
plotSpatialContext(
  object,
  entry = "spatial_context",
  group_by = "sample_id",
  node_color_by = NULL,
  node_size_by = NULL,
  node_color_fix = NULL,
  node_size_fix = NULL,
  node_label_repel = TRUE,
  node_label_color_by = NULL,
  node_label_color_fix = NULL,
  draw_edges = TRUE,
  edge_color_fix = NULL,
  return_data = FALSE
)
```

Arguments

object	a SingleCellExperiment or SpatialExperiment object.
entry	single character specifying the colData(object) entry containing the detectSpatialContext output. Defaults to "spatial_context".
group_by	a single character indicating the colData(object) entry by which SCs are grouped. This is usually the image or patient ID. Defaults to "sample_id".
node_color_by	single character either NULL, "name","n_cells", "n_group" by which the nodes should be colored.
node_size_by	single character either NULL, "n_cells","n_group" by which the size of the nodes are defined.
node_color_fix	single character specifying the color of all nodes.
node_size_fix	single numeric specifying the size of all nodes.
node_label_repel	should nodes be labelled? Defaults to TRUE.

node_label_color_by
: single character either NULL, "name","n_cells","n_group" by which the node labels should be colored.

node_label_color_fix
: single character specifying the color of all node labels.

draw_edges
: should edges be drawn between nodes? Defaults to TRUE.

edge_color_fix
: single character specifying the color of all edges.

return_data
: should the edge list and vertex metadata for graph construction be returned as a list of two data.frames?

Value

returns a ggplot object or a list of two data.frames.

Author(s)

Lasse Meyer (<lasse.meyer@uzh.ch>)

References

Bhate S. et al., Tissue schematics map the specialization of immune tissue motifs and their appropriation by tumors, Cell Systems, 2022

See Also

detectSpatialContext for the function to detect spatial contexts

filterSpatialContext for the function to filter spatial contexts

Examples

```
set.seed(22)
library(cytomapper)
data(pancreasSCE)

## 1. Cellular neighborhood (CN)
sce <- buildSpatialGraph(pancreasSCE, img_id = "ImageNb",
                         type = "knn",
                         name = "knn_cn_graph",
                         k = 5)

sce <- aggregateNeighbors(sce, colPairName = "knn_cn_graph",
                          aggregate_by = "metadata",
                          count_by = "CellType",
                          name = "aggregatedCellTypes")

cur_cluster <- kmeans(sce$aggregatedCellTypes, centers = 3)
sce$cellular_neighborhood <- factor(cur_cluster$cluster)

plotSpatial(sce, img_id = "ImageNb",
            colPairName = "knn_cn_graph",
```

```
            node_color_by = "cellular_neighborhood",
            scales = "free")

## 2. Spatial context (SC)
sce <- buildSpatialGraph(sce, img_id = "ImageNb",
                         type = "knn",
                         name = "knn_sc_graph",
                         k = 15)

sce <- aggregateNeighbors(sce, colPairName = "knn_sc_graph",
                          aggregate_by = "metadata",
                          count_by = "cellular_neighborhood",
                          name = "aggregatedNeighborhood")

# Detect spatial context
sce <- detectSpatialContext(sce, entry = "aggregatedNeighborhood",
                            threshold = 0.9)

plotSpatial(sce, img_id = "ImageNb",
            colPairName = "knn_sc_graph",
            node_color_by = "spatial_context",
            scales = "free")

# Plot spatial context - default
plotSpatialContext(sce, group_by = "ImageNb")

# Plot spatial context - adjust aesthetics
plotSpatialContext(sce, group_by = "ImageNb",
                   node_color_by = "name",
                   node_size_by = "n_cells",
                   node_label_color_by = "name")

plotSpatialContext(sce, group_by = "ImageNb",
                   node_color_by = "n_cells",
                   node_size_by = "n_group")

# Plot spatial context - return data
plotSpatialContext(sce, group_by = "ImageNb",
                  return_data = TRUE)
```

---

plotSpotHeatmap *Summarizes and visualizes the pixel intensities per spot and channel*

Description

Helper function for estimating the spillover matrix. This function visualizes the median pixel intensities per spot (rows) and per channel (columns) in form of a heatmap.

Usage

```
plotSpotHeatmap(
  object,
  spot_id = "sample_id",
  channel_id = "channel_name",
  assay_type = "counts",
  statistic = "median",
  log = TRUE,
  threshold = NULL,
  order_metals = TRUE,
  color = viridis(100),
  breaks = NA,
  legend_breaks = NA,
  cluster_cols = FALSE,
  cluster_rows = FALSE,
  ...
)
```

Arguments

object	a SingleCellExperiment object containing pixel intensities per channel. Individual pixels are stored as columns and channels are stored as rows.
spot_id	character string indicating which colData(object) entry stores the isotope names of the spotted metal. Entries should be of the form (mt)(mass) (e.g. Sm152 for Samarium isotope with the atomic mass 152).
channel_id	character string indicating which rowData(object) entry contains the isotope names of the acquired channels.
assay_type	character string indicating which assay to use (default counts).
statistic	the statistic to use when aggregating channels per spot (default median)
log	should the aggregated pixel intensities be log10(x + 1) transformed?
threshold	single numeric indicating a threshold after pixel aggregation. All aggregated values larger than threshold will be labeled as 1.
order_metals	should the metals be ordered based on spotted mass?
color	see parameter in pheatmap
breaks	see parameter in pheatmap
legend_breaks	see parameter in pheatmap
cluster_cols	see parameter in pheatmap
cluster_rows	see parameter in pheatmap
...	other arguments passed to pheatmap.

Value

a pheatmap object

Quality control for spillover estimation

Visualizing the aggregated pixel intensities serves two purposes:

1. Small median pixel intensities (< 200 counts) might hinder the robust estimation of the channel spillover. In that case, consecutive pixels can be summed (see `binAcrossPixels`).
2. Each spotted metal (row) should show the highest median pixel intensity in its corresponding channel (column). If this is not the case, either the naming of the .txt files was incorrect or the incorrect metal was spotted.

By setting the `threshold` parameter, the user can easily identify spots where pixel intensities are too low for robust spillover estimation.

Author(s)

Nils Eling (<nils.eling@dqbm.uzh.ch>)

See Also

`pheatmap` for visual modifications

`aggregateAcrossCells` for the aggregation function

Examples

```
path <- system.file("extdata/spillover", package = "imcRtools")
# Read in .txt files
sce <- readSCEfromTXT(path)

# Visualizes heatmap
plotSpotHeatmap(sce)

# Visualizes thresholding results
plotSpotHeatmap(sce, log = FALSE, threshold = 200)
```

---

readImagefromTXT *Reads one or multiple .txt files into a CytoImageList object*

Description

Reader function to generate `Image` objects in form of a `CytoImageList` container from .txt files.

Usage

```
readImagefromTXT(
  path,
  pattern = ".txt$",
  channel_pattern = "[A-Za-z]{1,2}[0-9]{2,3}Di",
  index_names = c("X", "Y"),
  BPPARAM = SerialParam()
)
```

Arguments

path	Full path to where the individual .txt files are located. This is usualy the path where the .mcd file is located.
pattern	pattern to select which files should be read in (default ".txt$").
channel_pattern	regular expression to select the channel names from the files.
index_names	exact names of the columns storing the x and y coordinates of the image
BPPARAM	parameters for parallelized reading in of images. This is only recommended for very large images.

Value

returns a CytoImageList object containing one Image object per .txt file.

Imaging mass cytometry .txt files

As part of the raw data folder, the Hyperion imaging system writes out one .txt file per acquisition. These files store the ion counts per pixel and channel.

This function reads these .txt files into a single CytoImageList object for downstream analysis. The pattern argument allows selection of all .txt files or a specific subset of files. The channelNames of the CytoImageList object are determined by the channel_pattern argument.

Author(s)

Nils Eling (<nils.eling@dqbm.uzh.ch>)

See Also

CytoImageList for the container

MulticoreParam for parallelized processing

Image for the multi-channel image object

vignette("cytomapper") for visualization of multi-channel images

Examples

```
path <- system.file("extdata/mockData/raw", package = "imcRtools")

# Read in all images
x <- readImagefromTXT(path)
x

# Read in specific files
y <- readImagefromTXT(path, pattern = "ROI_002")
y

# Read in other channelNames
z <- readImagefromTXT(path, channel_pattern = "[A-Za-z]{2}[0-9]{3}")
z
```

readSCEfromTIFF *Generates a SingleCellExperiment from .tiff files*

Description

Helper function to process .tiff files created with the steinbock pipeline into a SingleCellExperiment object. This function is mainly used to read-in data generated from a "spillover slide" from the new XTi generation of IMC machines. Here, each .tiff file contains the measurements of multiple pixels for a single stain across all open channels.

Usage

```
readSCEfromTIFF(x, image_df_path, panel_df_path, verbose = TRUE)
```

Arguments

x	has to be a path to a folder containing .tiff files.
image_df_path	has to be a path to a images.csv file, generated with the steinbock pipeline.
panel_df_path	has to be a path to a panel.csv file, generated with the steinbock pipeline.
verbose	logical indicating if additional information regarding the spotted and acquired masses should be shown.

Value

returns a SCE object where pixels are stored as columns and acquired channels are stored as rows.

Reading in .tiff files for spillover correction

As described in the original publication, single metal spots are acquired using the Hyperion imaging system. Each acquisition corresponds to one spot. All acquisitions are stored in a single .mcd file and individual acquisitions are stored in single .tiff files after extraction with the steinbock pipeline.

This function aggregates these measurements into a single SingleCellExperiment object:

x is a path: All .tiff files are read in from the specified path. Here, the path should indicate the location of the spillover slide measurement. Additionally, the images.csv and panel.csv files generated with the steinbock pipeline must be passed. The column acquisition_description in images.csv as well as the column channel in panel.csv must contain the spotted metal isotope name in the format (mt)(mass) (e.g. Sm152 for Samarium isotope with the atomic mass 152).

Author(s)

Victor Ibañez (<victor.ibanez@uzh.ch>)

References

Chevrier, S. et al. 2017. "Compensation of Signal Spillover in Suspension and Imaging Mass Cytometry." Cell Systems 6: 612–20. Windhager, J. et al. 2023. "An end-to-end workflow for multiplexed image processing and analysis." Nature Protocols 18: 3565–3613.

Examples

```
# Read files from path
path <- system.file("extdata/spillover_tiff/img", package = "imcRtools")
image_df_path <- system.file("extdata/spillover_tiff/images.csv", package = "imcRtools")
panel_df_path <- system.file("extdata/spillover_tiff/panel.csv", package = "imcRtools")

sce <- readSCEfromTIFF(path, image_df_path, panel_df_path)
sce
```

`readSCEfromTXT` *Generates a SingleCellExperiment from .txt files*

Description

Helper function to process raw .txt files acquired by the Hyperion imaging system into a `SingleCellExperiment` object. This function is mainly used to read-in data generated from a "spillover slide". Here, each .txt file contains the measurements of multiple pixels for a single stain across all open channels.

Usage

```
readSCEfromTXT(
  x,
  pattern = ".txt$",
  metadata_cols = c("Start_push", "End_push", "Pushes_duration", "X", "Y", "Z"),
  verbose = TRUE,
  read_metal_from_filename = TRUE
)
```

Arguments

`x`	input can be of different types: **A path** Full path to where the single stain .txt files are located. **A list object** A named list object where each entry is a `data.frame` or coercible to one. The names of each entry indicate the spotted metals (see details).
`pattern`	pattern to select which files should be read in (default `".txt$"`). Only used when `x` is a path.
`metadata_cols`	character vector indicating which column entries of the .txt files should be saved in the `colData(sce)` slot.
`verbose`	logical indicating if additional information regarding the spotted and acquired masses should be shown.
`read_metal_from_filename`	should the sample metal and mass be extracted from the file/object names?

Value

returns a SCE object where pixels are stored as columns and acquired channels are stored as rows.

Reading in .txt files for spillover correction

As described in the original publication, single metal spots are acquired using the Hyperion imaging system. Each acquisition corresponds to one spot. All acquisitions are stored in a single .mcd file and individual acquisitions are stored in single .txt files.

This function aggregates these measurements into a single `SingleCellExperiment` object. For this, two inputs are possible:

1. `x` is a path: By default all .txt files are read in from the specified path. Here, the path should indicate the location of the spillover slide measurement. The file names of the .txt file must contain the spotted metal isotope name in the format `(mt)(mass)` (e.g. `Sm152` for Samarium isotope with the atomic mass 152). Internally, the last occurrence of such a pattern is read in as the metal isotope name and stored in the `colData(sce)$sample_id` slot.
2. `x` is a named list: If there are issues with reading in the metal isotope names from the .txt file names, the user can provide a list for which each entry contains the contents of a single .txt file. The names of the list must indicate the spotted metal in the format `(mt)(mass)`. These names will be stored in the `colData(sce)$sample_id` slot.

When `read_metal_from_filename = FALSE`, the function will not attempt to read in the spotted metal isotopes from the file or list names. Therefore, only the `sample_id` will be set based on the file/list names.

Author(s)

Nils Eling (<nils.eling@dqbm.uzh.ch>)

References

Chevrier, S. et al. 2017. "Compensation of Signal Spillover in Suspension and Imaging Mass Cytometry." Cell Systems 6: 612–20.

Examples

```
# Read files from path
path <- system.file("extdata/spillover", package = "imcRtools")

sce <- readSCEfromTXT(path)
sce

# Read files as list
cur_file_names <- list.files(path, pattern = ".txt", full.names = TRUE)
cur_files <- lapply(cur_file_names, read.delim)
names(cur_files) <- sub(".txt", "", basename(cur_file_names))

sce <- readSCEfromTXT(cur_files)
sce
```

read_cpout *Reads in single-cell data generated by the ImcSegmentationPipeline*

Description

Reader function to generate a SpatialExperiment or SingleCellExperiment object from single-cell data obtained by the ImcSegmentationPipeline pipeline.

Usage

```
read_cpout(
  path,
  object_file = "cell.csv",
  image_file = "Image.csv",
  panel_file = "panel.csv",
  graph_file = "Object relationships.csv",
  object_feature_file = "var_cell.csv",
  intensities = "Intensity_MeanIntensity_FullStack",
  extract_imgid_from = "ImageNumber",
  extract_cellid_from = "ObjectNumber",
  extract_coords_from = c("Location_Center_X", "Location_Center_Y"),
 extract_cellmetadata_from = c("AreaShape_Area", "Neighbors_NumberOfNeighbors_8",
  "AreaShape_Eccentricity", "AreaShape_MajorAxisLength", "AreaShape_MinorAxisLength",
    "AreaShape_MeanRadius"),
  extract_imagemetadata_from = c("Metadata_acname", "Metadata_acid",
    "Metadata_description"),
  extract_graphimageid_from = "First Image Number",
  extract_graphcellids_from = c("First Object Number", "Second Object Number"),
  extract_metal_from = "Metal Tag",
  scale_intensities = TRUE,
  extract_scalingfactor_from = "Scaling_FullStack",
  return_as = c("spe", "sce")
)
```

Arguments

path	full path to the CellProfiler output folder.
object_file	single character indicating the file name storing the object/cell-specific intensities and metadata.
image_file	single character indicating the file name storing meta data per image (can be NULL).
panel_file	single character indicating the file name storing the panel information (can be NULL).
graph_file	single character indicating the file name storing the object/cell interaction information (can be NULL).

`object_feature_file`
: single character indicating the file name storing object/cell feature information.

`intensities`
: single character indicating which column entries of the `object_file` contain the intensity features of interest. See details.

`extract_imgid_from`
: single character indicating which column entries of the `object_file` and `image_file` contain the image integer ID.

`extract_cellid_from`
: single character indicating which column entry of the `object_file` contains the object/cell integer ID.

`extract_coords_from`
: character vector indicating which column entries of the `object_file` contain the x and y location of the objects/cells.

`extract_cellmetadata_from`
: character vector indicating which additional object/cell specific metadata to extract from the `object_file`.

`extract_imagemetadata_from`
: character vector indicating which additional image specific metadata to extract from the `image_file`. These will be stored in the `colData(x)` slot as object/cell-specific entries.

`extract_graphimageid_from`
: single character indicating which column entries of the `graph_file` contain the image integer ID.

`extract_graphcellids_from`
: character vector indicating which column entries of the `graph_file` contain the first and second object/cell integer IDs. These will be stored as the `from` and `to` entry of the `SelfHits` object in colPair(x, "neighborhood").

`extract_metal_from`
: single character indicating which column entry of the `panel_file` contains the metal isotopes of the used antibodies. This entry is used to match the panel information to the acquired channel information.

`scale_intensities`
: single logical. Should the measured intensity features be scaled by `extract_scalingfactor_from`.

`extract_scalingfactor_from`
: single character indicating which column entries of the `image_file` contain the image specific scaling factor.

`return_as`
: should the object be returned as `SpatialExperiment` (`return_as = "spe"`) or `SingleCellExperiment` (`return_as = "sce"`).

Value

returns a `SpatialExperiment` or `SingleCellExperiment` object with markers in rows and cells in columns.

The returned data container

In the case of both containers `x`, intensity features (as selected by the `intensities` parameter) are stored in the `counts(x)` slot. Cell metadata (e.g morphological features) are stored in the `colData(x)` slot. The interaction graphs are stored as `SelfHits` object in the `colPair(x, "neighborhood")` slot.

Intensity features are extracted via partial string matching. Internally, the `read_cpout` function checks if per channel a single intensity feature is read in (by checking the `_cXY` ending where `XY` is the channel number).

In the case of a returned `SpatialExperiment` object, the cell coordinates are stored in the `spatialCoords(x)` slot.

In the case of a returned `SingleCellExperiment` object, the cell coordinates are stored in the `colData(x)` slot named as `Pos_X` and `Pos_Y`.

Author(s)

Tobias Hoch

Nils Eling (<nils.eling@dqbm.uzh.ch>)

See Also

https://github.com/BodenmillerGroup/ImcSegmentationPipeline for the pipeline

`read_steinbock` for reading in single-cell data as produced by the steinbock pipeline

`colPair` for information on how to work with the cell-cell interaction graphs

Examples

```
path <- system.file("extdata/mockData/cpout", package = "imcRtools")

# Read in as SpatialExperiment object
x <- read_cpout(path, graph_file = "Object_relationships.csv")
x

# Read in as SingleCellExperiment object
x <- read_cpout(path, graph_file = "Object_relationships.csv",
                return_as = "sce")
x
```

---

read_steinbock *Reads in single-cell data generated by the steinbock pipeline*

Description

Reader function to generate a `SpatialExperiment` or `SingleCellExperiment` object from single-cell data obtained by the steinbock pipeline.

Usage

```
read_steinbock(
  path,
  intensities_folder = "intensities",
  regionprops_folder = "regionprops",
  graphs_folder = "neighbors",
  pattern = NULL,
  extract_cellid_from = "Object",
  extract_coords_from = c("centroid-1", "centroid-0"),
  image_file = "images.csv",
  extract_imagemetadata_from = c("width_px", "height_px"),
  panel_file = "panel.csv",
  extract_names_from = "name",
  return_as = c("spe", "sce"),
  BPPARAM = SerialParam()
)
```

Arguments

path	full path to the steinbock output folder
intensities_folder	name of the folder containing the intensity measurements per image
regionprops_folder	name of the folder containing the cell-specific morphology and spatial measurements per image. Can be set to NULL to exclude reading in morphology measures.
graphs_folder	name of the folder containing the spatial connectivity graphs per image. Can be set to NULL to exclude reading in graphs.
pattern	regular expression specifying a subset of files that should be read in.
extract_cellid_from	single character indicating which column entry in the intensity files contains the integer cell id.
extract_coords_from	character vector indicating which column entries in the regionprops files contain the x (first entry) and y (second entry) coordinates.
image_file	single character indicating the file name storing meta data per image (can be NULL).
extract_imagemetadata_from	character vector indicating which additional image specific metadata to extract from the image_file. These will be stored in the colData(x) slot as object/cell-specific entries.
panel_file	single character containing the name of the panel file. This can either be inside the steinbock path (recommended) or located somewhere else.
extract_names_from	single character indicating the column of the panel file containing the channel names.

return_as should the object be returned as SpatialExperiment (return_as = "spe") or SingleCellExperiment (return_as = "sce").

BPPARAM parameters for parallelised processing.

Value

returns a SpatialExperiment or SingleCellExperiment object markers in rows and cells in columns.

The returned data container

In the case of both containers x, intensity features are stored in the counts(x) slot. Morphological features are stored in the colData(x) slot. The graphs are stored as SelfHits object in the colPair(x, "neighborhood") slot.

In the case of a returned SpatialExperiment object, the cell coordinates are stored in the spatialCoords(x) slot.

In the case of a returned SingleCellExperiment object, the cell coordinates are stored in the colData(x) slot named as Pos_X and Pos_Y.

Author(s)

Nils Eling (<nils.eling@dqbm.uzh.ch>)

See Also

https://github.com/BodenmillerGroup/steinbock for the pipeline

read_cpout for reading in single-cell data as produced by the ImcSegmentationPipeline

SingleCellExperiment and SpatialExperiment for the constructor functions.

colPair for information on how to work with the cell-cell interaction graphs

bpparam for the parallelised backend

Examples

```
path <- system.file("extdata/mockData/steinbock", package = "imcRtools")

# Read in as SpatialExperiment object
x <- read_steinbock(path)
x

# Read in as SingleCellExperiment object
x <- read_steinbock(path, return_as = "sce")
x

# Read in a subset of files
x <- read_steinbock(path, pattern = "mockData1")
x

# Only read in intensities
x <- read_steinbock(path, graphs_folder = NULL, regionprops_folder = NULL)
```

```
x

# Parallelisation
x <- read_steinbock(path, BPPARAM = BiocParallel::bpparam())
```

`show_cpout_features` *Display all features measured by CellProfiler.*

Description

Searchable datatable object of cell and image features as extracted by CellProfiler.

Usage

```
show_cpout_features(
  path,
  display = c("cell_features", "image_features"),
  cell_features = "var_cell.csv",
  image_features = "var_Image.csv"
)
```

Arguments

`path`	full path to the CellProfiler output folder
`display`	single character indicating which features to display. Accepted entries are `cell_features` to display extracted single-cell features or `image_features` to display extracted image-level features.
`cell_features`	single character indicating the name of the file storing the extracted cell features.
`image_features`	single character indicating the name of the file storing the extracted image features.

Value

a `datatable` object

Author(s)

Nils Eling (<nils.eling@dqbm.uzh.ch>)

See Also

`read_cpout` for the CellProfiler reader function

Examples

```
path <- system.file("extdata/mockData/cpout", package = "imcRtools")

# Display cell features
show_cpout_features(path)

# Display image features
show_cpout_features(path, display = "image_features")
```

`testInteractions` *Tests if cell types interact more or less frequently than random*

Description

Cell-cell interactions are summarized in different ways and the resulting count is compared to a distribution of counts arising from random permutations.

Usage

```
testInteractions(
  object,
  group_by,
  label,
  colPairName,
  method = c("classic", "conditional", "patch", "interaction"),
  patch_size = NULL,
  iter = 1000,
  p_threshold = 0.01,
  return_samples = FALSE,
  tolerance = sqrt(.Machine$double.eps),
  BPPARAM = SerialParam()
)
```

Arguments

`object`	a `SingleCellExperiment` or `SpatialExperiment` object.
`group_by`	a single character indicating the `colData(object)` entry by which interactions are grouped. This is usually the image ID or patient ID.
`label`	single character specifying the `colData(object)` entry which stores the cell labels. These can be cell-types labels or other metadata entries.
`colPairName`	single character indicating the `colPair(object)` entry containing cell-cell interactions in form of an edge list.
`method`	which cell-cell interaction counting method to use (see details)
`patch_size`	if `method = "patch"`, a single numeric specifying the minimum number of neighbors of the same type to be considered a patch (see details)

iter	single numeric specifying the number of permutations to perform
p_threshold	single numeric indicating the empirical p-value threshold at which interactions are considered to be significantly enriched or depleted per group.
return_samples	single logical indicating if the permuted interaction counts of all iterations should be returned.
tolerance	single numeric larger than 0. This parameter defines the difference between the permuted count and the actual counts at which both are regarded as equal. Default taken from all.equal.
BPPARAM	parameters for parallelized processing.

Counting and summarizing cell-cell interactions

In principle, the countInteractions function counts the number of edges (interactions) between each set of unique entries in colData(object)[[label]]. Simplified, it counts for each cell of type A the number of neighbors of type B. This count is averaged within each unique entry colData(object)[[group_by]] in four different ways:

1. method = "classic": The count is divided by the total number of cells of type A. The final count can be interpreted as "How many neighbors of type B does a cell of type A have on average?"

2. method = "conditional": Formerly named "histocat". The count is divided by the number of cells of type A that have at least one neighbor of type B. The final count can be interpreted as "How many neighbors of type B has a cell of type A on average, given it has at least one neighbor of type B?".

3. method = "patch": For each cell, the count is binarized to 0 (less than patch_size neighbors of type B) or 1 (more or equal to patch_size neighbors of type B). The binarized counts are averaged across all cells of type A. The final count can be interpreted as "What fraction of cells of type A have at least a given number of neighbors of type B?"

4. method = "interaction": The count is divided by the total number of interactions from cell type A. The final count can be interpreted as "What fraction of interactions of cell type A occur with cell type B."

Testing for significance

Within each unique entry to colData(object)[[group_by]], the entries of colData(object)[[label]] are randomized iter times. For each iteration, the interactions are counted as described above. The result is a distribution of the interaction count under spatial randomness. The observed interaction count is compared against this Null distribution to derive empirical p-values:

p_gt: fraction of perturbations equal or greater than the observed count

p_lt: fraction of perturbations equal or less than the observed count

Based on these empirical p-values, the interaction score (attraction or avoidance), overall p value and significance by comparison to p_treshold (sig and sigval) are derived.

Author(s)

Vito Zanotelli

Jana Fischer

adapted by Nils Eling (<nils.eling@dqbm.uzh.ch>)

adapted by Marlene Lutz (<marlene.lutz@uzh.ch>)

adapted by Chiara Schiller (<chiara.schiller@uni-heidelberg.de>)

References

Schulz, D. et al., Simultaneous Multiplexed Imaging of mRNA and Proteins with Subcellular Resolution in Breast Cancer Tissue Samples by Mass Cytometry., Cell Systems 2018 6(1):25-36.e5

Schapiro, D. et al., histoCAT: analysis of cell phenotypes and interactions in multiplex image cytometry data, Nature Methods 2017 14, p. 873–876

Schiller, C. et al., Comparison and Optimization of Cellular Neighbor Preference Methods for Quantitative Tissue Analysis, biorRxiv 2025.03.31.646289

See Also

countInteractions for counting (but not testing) cell-cell interactions per grouping level.

bpparam for the parallelised backend

Examples

```
library(cytomapper)
library(BiocParallel)
data(pancreasSCE)

pancreasSCE <- buildSpatialGraph(pancreasSCE, img_id = "ImageNb",
                                 type = "knn", k = 3)

# Classic style calculation - setting the seed inside SerialParam for reproducibility
(out <- testInteractions(pancreasSCE,
                         group_by = "ImageNb",
                         label = "CellType",
                         method = "classic",
                         colPairName = "knn_interaction_graph",
                         iter = 1000,
                         BPPARAM = SerialParam(RNGseed = 123)))

# Conditional style calculation
(out <- testInteractions(pancreasSCE,
                         group_by = "ImageNb",
                         label = "CellType",
                         method = "conditional",
                         colPairName = "knn_interaction_graph",
                         iter = 1000,
                         BPPARAM = SerialParam(RNGseed = 123)))

# Patch style calculation
(out <- testInteractions(pancreasSCE,
                         group_by = "ImageNb",
                         label = "CellType",
                         method = "patch",
```

```
                          patch_size = 3,
                          colPairName = "knn_interaction_graph",
                          iter = 1000,
                          BPPARAM = SerialParam(RNGseed = 123)))

# Interaction style calculation
(out <- testInteractions(pancreasSCE,
                          group_by = "ImageNb",
                          label = "CellType",
                          method = "interaction",
                          colPairName = "knn_interaction_graph"))
```

Index

aggregateAcrossCells, *5*, *35*
aggregateNeighbors, 3, *13*
arrow, *29, 30*

binAcrossPixels, 4, *35*
BiocNeighborParam, *7*
bpparam, *44*, *48*
buildSpatialGraph, 6, *13*, *30*

channelNames, *36*
colPair, *42*, *44*
countInteractions, 8, *26*, *47, 48*
CytoImageList, *35, 36*

datatable, *45*
detectCommunity, 11
detectSpatialContext, 13, *19*, *31, 32*
distToCells, 15

filterPixels, 17
filterSpatialContext, *14*, 18, *32*
findBorderCells, 20
findKNN, *7, 8*
findNeighbors, *7, 8*

geometry, *29*
ggraph, *26*, *30*

Image, *35, 36*

KmknnParam, *7*

MulticoreParam, *36*

patchDetection, 21
patchSize, 24
pheatmap, *34, 35*
plotInteractions, 25
plotSpatial, *8*, 27
plotSpatialContext, *14*, *19*, 31
plotSpotHeatmap, 33

read_cpout, 40, *44, 45*
read_steinbock, *42*, 42
readImagefromTXT, 35
readSCEfromTIFF, 37
readSCEfromTXT, 38

SelfHits, *42*, *44*
show_cpout_features, 45
SingleCellExperiment, *37, 38*, *40–42*, *44*
SpatialExperiment, *40–42*, *44*

testInteractions, *10*, *26*, 46
triangulate, *7, 8*