Package 'msmsTests'

November 12, 2026

Type Package

Title LC-MS/MS Differential Expression Tests

Version 1.50.0

Date 2013-10-02

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Depends R (>= 3.0.1), MSnbase, msmsEDA

Imports edgeR, qvalue

Suggests xtable

Description Statistical tests for label-free LC-MS/MS data by spectral counts, to discover differentially expressed proteins between two biological conditions. Three tests are available: Poisson GLM regression, quasi-likelihood GLM regression, and the negative binomial of the edgeR package.The three models admit blocking factors to control for nuissance variables.To assure a good level of reproducibility a post-test filter is available, where we may set the minimum effect size considered biologicaly relevant, and the minimum expression of the most abundant condition.

License GPL-2

biocViews ImmunoOncology, Software, MassSpectrometry, Proteomics

git_url https://git.bioconductor.org/packages/msmsTests

git_branch RELEASE_3_23

git_last_commit c41561b

git_last_commit_date 2026-04-28

Repository Bioconductor 3.23

Date/Publication 2026-11-11

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`msmsTests-package` *LC-MS/MS Differential Expression Tests*

Description

Statistical tests for label-free LC-MS/MS data by spectral counts, to discover differentially expressed proteins between two biological conditions. Three tests are available: Poisson GLM regression, quasi-likelihood GLM regression, and the negative binomial of the edgeR package. The three models admit blocking factors to control for nuissance variables. To assure a good level of reproducibility a post-test filter is available, where we may set the minimum effect size considered biologicaly relevant, and the minimum expression of the most abundant condition.

Details

Package:	msmsTests
Type:	Package
Version:	0.99.1
Date:	2013-07-26
License:	GPL-2

`msms.glm.pois`:	Poisson based GLM regression
`msms.glm.qlll`:	Quasi-likelihood GLMregression
`msms.edgeR`:	The binomial negative of edgeR
`pval.by.fc`:	Table of cumulative frequencies of features by p-values in bins of log fold change
`test.results`:	Multitest p-value adjustement and post-test filter
`res.volcanoplot`:	Volcanplot of the results

Author(s)

Josep Gregori, Alex Sanchez, and Josep Villanueva
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References

Josep Gregori, Laura Villareal, Alex Sanchez, Jose Baselga, Josep Villanueva (2013). An Effect Size Filter Improves the Reproducibility in Spectral Counting-based Comparative Proteomics. Journal of Proteomics, DOI http://dx.doi.org/10.1016/j.jprot.2013.05.030

msms.edgeR *Spectral counts differential expression by edgeR*

Description

Given a null and an alternative model, with a two level treatment factor as the two conditions to compare, executes the negative binomial test by edgeR functions to discover differentially expressed proteins between the two conditions. The null and alternative models may include blocking factors.The reference level of the main factor is considered to be the control condition

Usage

```
msms.edgeR(msnset,form1,form0,facs=NULL,div=NULL,fnm=NULL)
```

Arguments

msnset	A MSnSet object with spectral counts in the expression matrix.
form1	The alternative hypothesis model as an standard R formula, with the treatment factor of interest, and eventual blocking factors.
form0	The null hypothesis model as an standard R formula.It may be the standard null model (y~.) or contain one or multiple blocking factors.
facs	NULL or a data frame with the factors in its columns.
div	NULL or a vector with the divisors used to compute the offsets.
fnm	NULL or a character string with the treatment factor name, as used in the column names of the factors data frame, and in the formula.

Details

The right hand site of the formulas is expected to be "y~", with the combination of factors after the tilde. If facs is NULL the factors are taken as default from pData(msnset). If div is NULL all divisors are taken equal to one. If fnm is NULL it is taken to be the first factor in facs.

Value

A data frame with column names 'LogFC', 'LR', 'p.value', with the estimated log fold changes, likelihood ratio statistic and corresponding p-value as obtaimed from a call to glmLRT() from the edgeR package.

Author(s)

Josep Gregori i Font

References

Robinson MD, McCarthy DJ and Smyth GK (2010). edgeR: a Bioconductor package for differential expression analysis of digital gene expression data. Bioinformatics 26, 139-140

Robinson MD and Smyth GK (2007). Moderated statistical tests for assessing differences in tag abundance. Bioinformatics 23, 2881-2887

Robinson MD and Smyth GK (2008). Small-sample estimation of negative binomial dispersion, with applications to SAGE data. Biostatistics, 9, 321-332

Josep Gregori, Laura Villareal, Alex Sanchez, Jose Baselga, Josep Villanueva (2013). An Effect Size Filter Improves the Reproducibility in Spectral Counting-based Comparative Proteomics. Journal of Proteomics, DOI http://dx.doi.org/10.1016/j.jprot.2013.05.030

See Also

MSnSet, edgeR, glmLRT, msmsEDA

Examples

```
## Example
library(msmsTests)
data(msms.dataset)
e <- pp.msms.data(msms.dataset)
e

null.f <- "y~batch"
alt.f <- "y~treat+batch"
div <- apply(exprs(e),2,sum)
res <- msms.edgeR(e,alt.f,null.f,div=div,fnm="treat")

str(res)
head(res)
```

---

msms.glm.pois *Spectral counts differential expression by Poisson GLM*

Description

Given a null and an alternative model, with a two level treatment factor as the two conditions to compare, executes a Poisson based GLM regression to discover differentially expressed proteins between the two conditions. The null and alternative models may include blocking factors.The reference level of the main factor is considered to be the control condition.

Usage

```
msms.glm.pois(msnset,form1,form0,facs=NULL,div=NULL)
```

Arguments

msnset	A MSnSet object with spectral counts in the expression matrix.
form1	The alternative hypothesis model as an standard R formula, with the treatment factor of interest, and eventual blocking factors.
form0	The null hypothesis model as an standard R formula.It may be the standard null model (y~.) or contain one or multiple blocking factors.
facs	NULL or a data frame with the factors in its columns.
div	NULL or a vector with the divisors used to compute the offsets.

Details

The right hand site of the formulas is expected to be "y~", with the combination of factors after the tilde. If facs is NULL the factors are taken as default from pData(msnset). If div is NULL all divisors are taken equal to one.

Value

A data frame with the following columns:

LogFC	Log fold change estimated from the model parameters.
D	Residual deviance as statistic of the test.
p.value	The p-values obtained from the test.

Author(s)

Josep Gregori i Font

References

Agresti, A. (2002) Categorical Data Analysis, 2nd Edition, John Wiley & Sons, Inc., Hoboken, New Jersey

Thompson L.A. (2009) R (and S-PLUS) Manual to Accompany Agresti s Categorical Data Analysis (2002), 2nd edition https://home.comcast.net/~lthompson221/Splusdiscrete2.pdf

Dobson, A.J. (2002) An Introduction to Generalized Linear Models, 2nd Edition, Chapman & Hall/CRC, New York

Venables, W. N. and Ripley, B. D. (2002) Modern Applied Statistics with S. New York: Springer

Josep Gregori, Laura Villareal, Alex Sanchez, Jose Baselga, Josep Villanueva (2013). An Effect Size Filter Improves the Reproducibility in Spectral Counting-based Comparative Proteomics. Journal of Proteomics, DOI http://dx.doi.org/10.1016/j.jprot.2013.05.030

See Also

MSnSet, glm

Examples

```
library(msmsTests)
data(msms.dataset)
# Pre-process expression matrix
e <- pp.msms.data(msms.dataset)
# Factors
pData(e)
# Control condition
levels(pData(e)$treat)[1]
# Treatment condition
levels(pData(e)$treat)[2]

# Models and normalizing condition
null.f <- "y~batch"
alt.f <- "y~treat+batch"
div <- apply(exprs(e),2,sum)

#Test
res <- msms.glm.pois(e,alt.f,null.f,div=div)

str(res)
head(res)
```

`msms.glm.qlll` *Spectral counts differential expression by quasi-likelihood GLM*

Description

Given a null and an alternative model, with a two level treatment factor as the two conditions to compare, executes a quasi-likelihood based GLM regression to discover differentially expressed proteins between the two conditions. The null and alternative models may include blocking factors.The reference level of the main factor is considered to be the control condition.

Usage

```
msms.glm.qlll(msnset,form1,form0,facs=NULL,div=NULL)
```

Arguments

`msnset`	A MSnSet object with spectral counts in the expression matrix.
`form1`	The alternative hypothesis model as an standard R formula, with the treatment factor of interest, and eventual blocking factors.
`form0`	The null hypothesis model as an standard R formula.It may be the standard null model (y~.) or contain one or multiple blocking factors.
`facs`	NULL or a data frame with the factors in its columns.
`div`	NULL or a vector with the divisors used to compute the offsets.

Details

The right hand site of the formulas is expected to be "y~", with the combination of factors after the tilde. If `facs` is NULL the factors are taken as default from `pData(msnset)`. If `div` is NULL all divisors are taken equal to one.

Value

A data frame with the following columns:

`LogFC`	Log fold change estimated from the model parameters.
`D`	Residual deviance as statistic of the test.
`p.value`	The p-values obtained from the test.

Author(s)

Josep Gregori i Font

References

Agresti, A. (2002) *Categorical Data Analysis*, 2nd Edition, John Wiley & Sons, Inc., Hoboken, New Jersey

Thompson L.A. (2009) *R (and S-PLUS) Manual to Accompany Agresti s Categorical Data Analysis*, 2nd edition https://home.comcast.net/~lthompson221/Splusdiscrete2.pdf

Dobson, A.J. (2002) *An Introduction to Generalized Linear Models*, 2nd Edition, Chapman & Hall/CRC, New York

Venables, W. N. and Ripley, B. D. (2002) *Modern Applied Statistics with S*, New York: Springer

Li, M.; Gray, W.; Zhang, H.; Chung, C. H.; Billheimer, D.; Yarbrough, W. G.; Liebler, D. C.; Shyr, Y.; Slebos, R. J. C. *Comparative shotgun proteomics using spectral count data and quasi-likelihood modeling*, J Proteome Res 2010, 9, 4295-4305.

Josep Gregori, Laura Villareal, Alex Sanchez, Jose Baselga, Josep Villanueva (2013). *An Effect Size Filter Improves the Reproducibility in Spectral Counting-based Comparative Proteomics*, Journal of Proteomics, http://dx.doi.org/10.1016/j.jprot.2013.05.030

See Also

`MSnSet`, `glm`

Examples

```
library(msmsTests)
data(msms.dataset)
# Pre-process expression matrix
e <- pp.msms.data(msms.dataset)
# Factors
pData(e)
# Control condition
levels(pData(e)$treat)[1]
# Treatment condition
```

```
levels(pData(e)$treat)[2]

# Models and normalizing condition
null.f <- "y~batch"
alt.f <- "y~treat+batch"
div <- apply(exprs(e),2,sum)

#Test
res <- msms.glm.qlll(e,alt.f,null.f,div=div)

str(res)
head(res)
```

---

msms.spk *Yeast lisate samples spiked with human proteins*

Description

A MSnSet with a spectral counts in the expression matrix and a treatment factor in the phenoData slot.
The spectral counts matrix has samples in the columns, and proteins in the rows. Each sample consists in 500ng of standard yeast lisate spiked with 100, 200, 400 and 600fm of a mix of 48 equimolar human proteins (UPS1, Sigma-Aldrich). The dataset contains a different number of technical replicates of each sample.

Usage

```
data(msms.spk)
```

Format

A MSnSet

References

Josep Gregori, Laura Villareal, Alex Sanchez, Jose Baselga, Josep Villanueva (2013). An Effect Size Filter Improves the Reproducibility in Spectral Counting-based Comparative Proteomics. Journal of Proteomics, DOI http://dx.doi.org/10.1016/j.jprot.2013.05.030

Laurent Gatto and Kathryn S. Lilley, MSnbase - an R/Bioconductor package for isobaric tagged mass spectrometry data visualization, processing and quantitation, Bioinformatics 28(2), 288-289 (2012).

See Also

See MSnSet for detail on the class, and the exprs and pData accessors.

Examples

```
data(msms.spk)
msms.spk
dim(msms.spk)
table(pData(msms.spk))
head(exprs(msms.spk))
```

`pval.by.fc` *Table of cumulative frequencies of p-values by log fold change bins*

Description

Given the set of p-values and log fold changes that result from a test, computes a table of cumulative frequencies of features by p-values in bins of log fold changes.

Usage

```
pval.by.fc(pvals,lfc)
```

Arguments

`lfc`	The log fold changes estimated from the tests.
`pvals`	The p-values, adjusted or not, obtained from the tests.

Value

A matrix of cumulated frequencies with descriptive row and column names.

Author(s)

Josep Gregori i Font

References

Josep Gregori, Laura Villareal, Alex Sanchez, Jose Baselga, Josep Villanueva (2013). An Effect Size Filter Improves the Reproducibility in Spectral Counting-based Comparative Proteomics. Journal of Proteomics, DOI http://dx.doi.org/10.1016/j.jprot.2013.05.030

See Also

`test.results`

Examples

```
library(msmsTests)
data(msms.spk)
# Subset
treat <- pData(msms.spk)
jdx <- which(treat=="U200" | treat=="U600")
e <- msms.spk[,jdx]
pData(e)$treat <- treat[jdx,1,drop=TRUE]
# Pre-process expression matrix
e <- pp.msms.data(e)
# Models and normalizing condition
null.f <- "y~1"
alt.f <- "y~treat"
div <- apply(exprs(e),2,sum)
#Test
res <- msms.glm.pois(e,alt.f,null.f,div=div)
# Post-test filter
lst <- test.results(res,e,pData(e)$treat,"U600","U200",div,
                    alpha=0.05,minSpC=2,minLFC=1,
                    method="BH")

##  On all features, with multitest adjusted p-values
pval.by.fc(lst$tres$adjp, lst$tres$LogFC)

### On all features deemed significant and biologically relevant
flags <- lst$tres$DEP
pval.by.fc(lst$tres$adjp[flags], lst$tres$LogFC[flags])
```

---

res.volcanoplot *Volcanoplot*

Description

Given the data frame obtained from `test.results()` a volcano plot is drawn.The features are colored according to significance and relevance.

Usage

```
res.volcanoplot(tres, max.pval=0.05, min.LFC=1, maxx=3, maxy=10,
                ylbls=20)
```

Arguments

`tres`	The dataframe with test results as obtained from `test.results()`. Or a data frame with, at least, the following columns: `LogFC` with log fold changes, `adjp` with multitest adjusted p-values, and `DEP` with TRUE or FALSE as post test filter results, being the TRUE features both statistically significant and relevant for reproducibility.
`max.pval`	The maximum adjusted p-value considered as statistically significant.

min.LFC	The minimum absolute log fold change considered as biologically relevant.
maxx	The maximum value in abcissas (i.e. log2(fold change)).
maxy	The maximum value in ordinates (i.e. -log10(p.val))
ylbls	All features with -log10(p.val) above this value will be ploted with feature labels.

Details

Abscissas and ordinates may be limited giving a value other than NULL to the parameters maxx and maxy. All features deemed significant and relevant are ploted by a blue dot, all features deemed significant but not passing the post test filter are plotted by a red dot. The non-significant features are plotted as smaller black dots. All features deemed significant and relevant and with a -log10 p-value above ylbls are plotted with a label showing their row index in the test results dataframe. The borders limiting the values given by max.pval and min.LFC are ploted as dash-and-dot red lines.

Value

No return value.

Author(s)

Josep Gregori i Font

References

Josep Gregori, Laura Villareal, Alex Sanchez, Jose Baselga, Josep Villanueva (2013). An Effect Size Filter Improves the Reproducibility in Spectral Counting-based Comparative Proteomics. Journal of Proteomics, DOI http://dx.doi.org/10.1016/j.jprot.2013.05.030

See Also

test.results, volcanoplot

Examples

```
library(msmsTests)
data(msms.dataset)
# Pre-process expression matrix
e <- pp.msms.data(msms.dataset)
# Models and normalizing condition
null.f <- "y~batch"
alt.f <- "y~treat+batch"
div <- apply(exprs(e),2,sum)
#Test
res <- msms.glm.qlll(e,alt.f,null.f,div=div)
lst <- test.results(res,e,pData(e)$treat,"U600","U200",div,
                    alpha=0.05,minSpC=2,minLFC=log2(1.8),
                    method="BH")
# Plot
```

```
res.volcanoplot(lst$tres, max.pval=0.05, min.LFC=1, maxx=3, maxy=NULL,
                        ylbls=4)
```

`test.results` *Multitest p-value adjustment and post-test filter*

Description

Operates on the statistic tests results obtained from `msms.glm.pois()`, `msms.glm.qlll()` or `msms.edgeR()`. The following variables are computed: Raw expression mean values for each condition (control and treatment), log fold change based on these expression levels and taking into account the normalizing divisors (`div`), multitest adjusted p-values with FDR control, and a post test filter based on minimum spectral counts and minimum absolute log fold change as estimated by the statistic test. According to the results of this post-test filter, features are flagged as T or F depending on whether they result relevant or not, beyond their statistic signicance.

Usage

```
test.results(test, msnset, gpf, gp1, gp2, div, alpha = 0.05,
             minSpC = 2, minLFC = 1, method = "BH")
```

Arguments

`test`	The dataframe obtained from either `msms.glm.pois()`, `msms.glm.qlll()` or `msms.edgeR()`
`msnset`	A MSnSet object with spectral counts in the expression matrix.
`gpf`	The factor used in the tests.
`gp1`	The treatment level name.
`gp2`	The control level name. Should be the factor's reference level. See R function `relevel`.
`div`	The weights used as divisors (offsets) in the GLM model. Usually the sum of spectral counts of each sample.
`alpha`	The multi test adjusted p-value significance threshold.
`minSpC`	The minimum spectral counts considered as relevant in the most abundant condition. This filter aims at reaching good reproducibility.
`minLFC`	The minimum absolute log fold change considered both, relevant and biologically significant. This filter aims at assuring enough biological effect size and at reaching good reproducibility.
`method`	One among `BH` or `qval`. The p-values are FDR ajusted by the Benjamini-Hochberg method (`BH`) or by qvalue (`qval`).

Details

No feature is removed in the filter, but instead they are flagged as TRUE or FALSE depending on whether they are considered as differentially expressed or not, in the `DEP` column, taking into account statistic significance and reproducibility metrics.

Value

A data frame with the following columns:

`first column`	Column named as the treatment level with the mean raw spectral counts observed for this condition
`second column`	Column named as the control level with the mean raw spectral counts observed for this condition
`lFC.Av`	Log fold change computed from the mean expression levesl taking into account the given normalization factors.
`logFC`	Log fold change estimated by fitting the given GLM model. The reference level of the main factor is taken as control.
`D or LR`	The statistic obtained from the tests. The residual deviance `D` for Poisson and quasi-likelihood, or the likelihood ratio `LR` for edgeR.
`p.val`	The unadjusted p-values obtained from the tests.
`adjp`	The multitest adjusted p-values with FDR control.
`DEP`	A logical flagging the features considered both as statistically significant and relevant for reproducibility.

Author(s)

Josep Gregori i Font

References

Josep Gregori, Laura Villareal, Alex Sanchez, Jose Baselga, Josep Villanueva (2013). An Effect Size Filter Improves the Reproducibility in Spectral Counting-based Comparative Proteomics. Journal of Proteomics, DOI http://dx.doi.org/10.1016/j.jprot.2013.05.030

Benjamini, Y., and Hochberg, Y. (1995). Controlling the false discovery rate: a practical and powerful approach to multiple testing. Journal of the Royal Statistical Society Series B, 57, 289-300.

Alan Dabney, John D. Storey and with assistance from Gregory R. Warnes. qvalue: Q-value estimation for false discovery rate control. R package version 1.30.0.

See Also

`pval.by.fc`, `p.adjust`, `qvalue`,

Examples

```
library(msmsTests)
data(msms.dataset)
# Pre-process expression matrix
e <- pp.msms.data(msms.dataset)
# Factors
pData(e)
# Control condition
levels(pData(e)$treat)[1]
# Treatment condition
```

```
levels(pData(e)$treat)[2]
# Models and normalizing condition
null.f <- "y~batch"
alt.f <- "y~treat+batch"
div <- apply(exprs(e),2,sum)
#Test
res <- msms.glm.qlll(e,alt.f,null.f,div=div)
# Post-test filter
lst <- test.results(res,e,pData(e)$treat,"U600","U200",div,
                    alpha=0.05,minSpC=2,minLFC=1,
                    method="BH")
str(lst)
lst$cond
head(lst$tres)
rownames(lst$tres)[which(lst$tres$DEP)]
```

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