Package 'phenopath'

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`clvm` *Fit a CLVM Model*

Description

Fit a covariate latent variable model using coordinate ascent variational inference.

Usage

```
clvm(y, x, maxiter = 10000, elbo_tol = 1e-05, thin = 1, verbose = TRUE,
  z_init = 1, tau_q = 1, tau_mu = 1, tau_c = 1, a = 2, b = 2,
  tau_alpha = 1, a_beta = 10, b_beta = 1, q = rep(0, nrow(y)),
  model_mu = FALSE, scale_y = TRUE)
```

Arguments

`y`	A N-by-G (dynamic) input matrix
`x`	A N-by-P (static) input matrix
`maxiter`	Maximum number of CAVI iterations
`elbo_tol`	The (percent) change in the ELBO below which it is considered converged
`thin`	The number of iterations to wait each time before re-calculating the elbo
`verbose`	Print convergence messages
`z_init`	The initialisation of the latent trajectory. Should be one of 1. A positive integer describing which principal component of the data should be used for initialisation (default 1), *or* 2. A numeric vector of length number of samples to be used directly for initialisation, *or*

3. The text character "random", for random initialisation from a standard normal distribution.

tau_q	Hyperparameter tau_q
tau_mu	Hyperparameter tau_mu
tau_c	Hyperparameter tau_c
a	Hyperparameter a
b	Hyperparameter b
tau_alpha	Hyperparameter tau_alpha
a_beta	Hyperparameter a_beta
b_beta	Hyperparameter b_beta
q	Priors on the latent variables
model_mu	Logical - should a gene-specific intercept term be modelled?
scale_y	Logical - should the expression matrix be centre scaled?

Value

A list whose entries correspond to the converged values of the variational parameters along with the ELBO.

Examples

```
sim <- simulate_phenopath()
fit <- clvm(sim$y, matrix(sim$x))
```

---

interactions *Tidy summary of interactions*

Description

Computes a tidy data frame of the interaction effects, pathway scores, and significance for each gene and covariate interaction.

Usage

```
interactions(phenopath_fit, n = 3)
```

Arguments

phenopath_fit	An object returned by a call to phenopath
n	The number of standard deviations away from 0 the posterior of the interaction effect sizes should be to be designated as significant

Value

A data frame with the following columns:

- `feature` The feature (usually gene)
- `covariate` The covariate, specified from the order originally supplied to the call to `phenopath`
- `interaction_effect_size` The effect size of the interaction ($beta$ from the statistical model)
- `significant` Boolean for whether the interaction effect is significantly different from 0
- `chi` The precision of the ARD prior on $beta$
- `pathway_loading` The pathway loading $lambda$, showing the overall effect for each gene marginalised over the covariate

Examples

```
sim <- simulate_phenopath()
fit <- phenopath(sim$y, sim$x, elbo_tol = 1e-2)
interactions(fit)
```

`interaction_effects` *Get the interaction effect sizes*

Description

Get the interaction effect sizes

Usage

```
interaction_effects(phenopath_fit)
```

Arguments

`phenopath_fit`	An object of class `phenopath_fit`

Value

TODO

Examples

```
sim <- simulate_phenopath()
fit <- phenopath(sim$y, sim$x, elbo_tol = 1e-2)
beta <- interaction_effects(fit)
```

interaction_sds *Get the interaction standard deviations*

Description

Get the interaction standard deviations

Usage

```
interaction_sds(phenopath_fit)
```

Arguments

phenopath_fit	An object of class phenopath_fit

Value

TODO

Examples

```
sim <- simulate_phenopath()
fit <- phenopath(sim$y, sim$x, elbo_tol = 1e-2)
beta_sd <- interaction_sds(fit)
```

phenopath *PhenoPath - genomic trajectories with heterogeneous backgrounds*

Description

PhenoPath learns genomic trajectories in the presence of heterogenous environmental and genetic backgrounds. It takes input gene expression measurements that are modelled by a single unobserved factor (the "trajectory"). The regulation of genes along the trajectory is perturbed by an additional set of covariates (such as genetic or environmental status) allowing for the identification of covariate-trajectory interactions. The model is fitted using mean-field co-ordinate ascent variational inference.

Usage

```
phenopath(exprs_obj, x, sce_assay = "exprs", elbo_tol = 1e-05, z_init = 1,
  ...)
```

Arguments

exprs_obj	Input gene expression, either 1. An SummarizedExperiment object, *or* 2. A cell-by-gene matrix of normalised expression values in log form.
x	The covariate vector, either 1. The name of a column of `colData(exprs_obj)` if `exprs_obj` is an `SummarizedExperiment`, *or* 2. A numeric of factor vector of length equal to the number of cells, *or* 3. A formula from which to build a model matrix from `colData(exprs_obj)`, if `exprs_obj` is a SummarizedExperiment
sce_assay	The assay from `exprs_obj` to use as expression if `exprs_object` is a `SummarizedExperiment`
elbo_tol	The relative pct change in the ELBO below which is considered converged. See convergence section in details below.
z_init	The initialisation of the latent trajectory. Should be one of 1. A positive integer describing which principal component of the data should be used for initialisation (default 1), *or* 2. A numeric vector of length number of samples to be used directly for initialisation, *or* 3. The text character `"random"`, for random initialisation from a standard normal distribution.
...	Additional arguments to be passed to `clvm`. See description below for more details or call `?clvm`.

Details

Input expression

If an `SummarizedExperiment` is provided, `assay(exprs_obj, sce_assay)` is used. This is assumed to be in a form that is suitably normalised and approximately normal, such as the log of TPM values (plus a suitable offset) or similar.

Encoding covariates

See the vignette.

Convergence of variational inference

It is strongly recommended to call `plot_elbo(phenopath_fit)` after the fitting procedure to ensure the ELBO has approximately converged (though convergence metrics are printed during the fitting process). For a good introduction to variational inference see Blei, D.M., Kucukelbir, A. & McAuliffe, J.D., 2017. Variational Inference: A Review for Statisticians. Journal of the American Statistical Association.

Additional arguments

Addition arguments to `clvm` are passed via `...`. For full documentation, call `?clvm`. Some notable options:

- `thin` - The ELBO is expensive to compute for larger datasets. The model will compute the ELBO and compare convergence every `thin` iterations.

- q and tau_q - Priors (such as capture times) for the latent space. Note that model_mu should be true if q is non-zero.
- scale_y By default the input expression is centre-scaled for each gene. If scale_y is FALSE this does not happen - but note that model_mu should be TRUE in such a case.

Value

An S3 structure with the following entries:

- m_z The converged mean estimates of the trajectory
- s_z The converged standard deviation estimates of z
- m_beta A P-by-G matrix of interaction coefficients
- s_beta A P-by-G matrix of interaction standard deviations

See Also

clvm for the underlying CAVI function, trajectory to extract the latent trajectory, interaction_effects for the interaction effect sizes, significant_interactions for the results of Bayesian significance testing.

Examples

```
sim <- simulate_phenopath() # returns a list with gene expression in y and covariates in x
fit <- phenopath(sim$y, sim$x, elbo_tol = 1e-2)

# Extract the trajectory
z <- trajectory(fit)
```

---

plot_elbo *Plots the ELBO*

Description

Plots the evidence lower bound (ELBO) as a function of iterations

Usage

```
plot_elbo(fit)
```

Arguments

fit	An object returned by a call to phenopath

Value

A ggplot2 object of the ELBO against the number of iterations

Examples

```
sim <- simulate_phenopath()
fit <- phenopath(sim$y, sim$x)
plot_elbo(fit)
```

`print.phenopath_fit` *Print a PhenoPath fit*

Description

Print a PhenoPath fit

Usage

```
## S3 method for class 'phenopath_fit'
print(x, ...)
```

Arguments

`x`	A `phenopath_fit` returned by a call to `phenopath`
`...`	Additional arguments

Value

A string representation of a `phenopath_fit` object.

Examples

```
sim <- simulate_phenopath()
fit <- phenopath(sim$y, sim$x, elbo_tol = 1e-2)
print(fit)
```

`sample_fns` *Sample parameters for simulation*

Description

Sample parameters from de regime

Sample parameters from pseudotime regime

Sample parameters from pseudotime-interaction regime

Sample paramters from de-pseudotime-interaction regime

Usage

```
sample_de()

sample_pst()

sample_pst_beta()

sample_de_pst_beta()
```

Value

A length-3 vector of parameters corresponding to the particular simulation regime

---

`scale_vec` *Scale a vector to have mean 0 and variance 1*

Description

Scales vector to mean 0 variance 1 unless input standard deviation is 0 in which case original vector is returned

Usage

```
scale_vec(x)
```

Arguments

`x`	Input vector to scale

Value

Scaled vector

`significant_interactions` *Significance testing for interaction features*

Description

Given the results of `clvm`, decide which features show significant iteractions between the latent trajectory and covariates. Significant features are designated as those where the variational mean of the interaction coefficient falls outside the $n\sigma$ interval of 0.

Usage

```
significant_interactions(phenopath_fit, n = 3)
```

Arguments

phenopath_fit	The results of a call to clvm
n	The number of standard deviations away from 0 the posterior estimate of beta should be to be designated significant.

Value

A logical vector describing whether each feature passes the significance test.

Examples

```
sim <- simulate_phenopath()
fit <- phenopath(sim$y, sim$x, elbo_tol = 1e-2)
signints <- significant_interactions(fit)
```

---

simulate_one_gene *Simulate one gene*

Description

Simulate one gene from the model given paramters, z and covariates

Usage

```
simulate_one_gene(N, pst, x, alpha = 0, c = 0, beta = 0, tau = 1e+06)
```

Value

A length-N gene expression vector simulated with the PhenoPath mean function for the given parameters

---

simulate_phenopath *Simulate from phenopath model*

Description

Simulate synthetic data from the phenopath model, where each gene is sampled from one of four types (see details).

Usage

```
simulate_phenopath(N = 100, G = 40, G_de = NULL, G_pst = NULL,
  G_pst_beta = NULL, G_de_pst_beta = NULL)
```

Arguments

N	Number of samples to simulate
G	Number of genes to simulate. Should be divisible by 4
G_de	Number of genes to simulate from the *differential expression* regime
G_pst	Number of genes to simulate from the *pseudotime* regime
G_pst_beta	Number of genes to simulate from the *pseudotime* + *beta interactions* regime
G_de_pst_beta	Number of genes to simulate from the *differential expression* + *pseudotime* + *interactions* regime

Details

Will simulate data for a number of genes corresponding to one of four regimes:

1. de ('differential expression'), where the gene has no association to the latent trajectory and exhibits differential expression only
2. pst ('pseudotime'), the gene shows pseudotemporal regulation but no differential regulation
3. pst_beta ('pseudotime + beta interactions'), the gene shows pseudotemporal regulation that is modulated by covariate interactions
4. de_pst_beta ('differential expression + pseudotime + interactions), all of the above

Value

A list with four entries:

- parameters A tibble with an entry for each gene and a column for each parameter value and simulation regime (see details).
- y The N-by-G simulated gene expression matrix.
- x The N-length vector of covariates.
- z The N-length vector of pseudotimes.

Examples

```
sim <- simulate_phenopath()
```

---

trajectory *Get the latent PhenoPath trajectory*

Description

Get the latent PhenoPath trajectory

Usage

```
trajectory(phenopath_fit)
```

Arguments

phenopath_fit	An object of class phenopath_fit

Value

A vector of latent trajectory (pseudotime) values

Examples

```
sim <- simulate_phenopath()
fit <- phenopath(sim$y, sim$x, elbo_tol = 1e-2)
z <- trajectory(fit)
```

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