Package 'qsea'

November 12, 2026

Type Package

Title IP-seq data analysis and vizualization

Version 1.38.0

Date 2025-07-22

Description qsea (quantitative sequencing enrichment analysis) was developed as the successor of the MEDIPS package for analyzing data derived from methylated DNA immunoprecipitation (MeDIP) experiments followed by sequencing (MeDIP-seq). However, qsea provides several functionalities for the analysis of other kinds of quantitative sequencing data (e.g. ChIP-seq, MBD-seq, CMS-seq and others) including calculation of differential enrichment between groups of samples.

License GPL-2

biocViews Sequencing, DNAMethylation, CpGIsland, ChIPSeq, Preprocessing, Normalization, QualityControl, Visualization, CopyNumberVariation, ChipOnChip, DifferentialMethylation

Depends R (>= 4.3)

Imports Biostrings, graphics, gtools, methods, stats, utils, HMMcopy, rtracklayer, BSgenome, GenomicRanges, Rsamtools, IRanges, limma, Seqinfo, BiocGenerics, grDevices, zoo, BiocParallel, S4Vectors

VignetteBuilder knitr

Suggests BSgenome.Hsapiens.UCSC.hg19, MEDIPSData, testthat, BiocStyle, knitr, rmarkdown, BiocManager, MASS

ByteCompile no

NeedsCompilation yes

git_url https://git.bioconductor.org/packages/qsea

git_branch RELEASE_3_23

git_last_commit 05161de

git_last_commit_date 2026-04-28

Repository Bioconductor 3.23

Date/Publication 2026-11-11

Author Matthias Lienhard [aut, cre] (ORCID:
<https://orcid.org/0000-0002-2549-3142>),
Lukas Chavez [aut] (ORCID: <https://orcid.org/0000-0002-8718-8848>),
Ralf Herwig [aut] (ORCID: <https://orcid.org/0000-0002-9335-1760>)

Maintainer Matthias Lienhard <lienhard@molgen.mpg.de>

Contents

qsea-package . . . 2
addCNV . . . 3
addContrast . . . 4
addCoverage . . . 6
addEnrichmentParameters . . . 7
addLibraryFactors . . . 8
addNewSamples . . . 9
addOffset . . . 10
addPatternDensity . . . 11
addSeqPref . . . 12
createQseaSet . . . 13
fitNBglm . . . 14
getExampleQseaSet . . . 16
getPCA . . . 17
isSignificant . . . 18
makeTable . . . 19
normMethod . . . 21
plotCNV . . . 22
plotCoverage . . . 23
plotEnrichmentProfile . . . 25
plotPCA . . . 26
qseaGLM-class . . . 28
qseaPCA-class . . . 28
qseaSet-class . . . 29
regionStats . . . 30

Index **32**

`qsea-package` *QSEA: Quantitative sequencing enrichment analysis and visualization*

Description

QSEA (quantitative sequencing enrichment analysis) was developed as the successor of the MEDIPS package for analyzing data derived from methylated DNA immunoprecipitation (MeDIP) experiments followed by sequencing (MeDIP-seq). However, qsea provides functionality for the analysis of other kinds of quantitative sequencing data (e.g. ChIP-seq, MBD-seq, CMS-seq and others) including calculation of differential enrichment between groups of samples.

Author(s)

Matthias Lienhard, Lukas Chavez and Ralf Herwig

Maintainer: Matthias Lienhard <lienhard@molgen.mpg.de>

References

Lienhard M, Grimm C, Morkel M, Herwig R, Chavez L., (Bioinformatics, 2014): MEDIPS: genome-wide differential coverage analysis of sequencing data derived from DNA enrichment experiments.

`addCNV` *estimate CNV information and add to qseaSet object*

Description

This function adds information on Copy Number Variation (CNV) to the qseaSet object, which is used for normalization. Sample wise CNV information can either be provided, or estimated from input or enrichment sequencing data, by incorporating functions of the HMMcopy package.

Usage

```
addCNV(qs,file_name, window_size=1000000, paired=FALSE, fragment_length,cnv,
mu=log2(c(1/2, 2/3, 1, 3/2,2,3)), normal_idx, plot_dir, MeDIP=FALSE,
parallel=FALSE)
```

Arguments

`qs`	the qseaSet object
`cnv`	pre-computed CNV information for each sample. If provided, the following parameters are ignored
`file_name`	column name of the sample table for the sequencing files, from which CNV information are computed
`window_size`	window size for CNV analysis
`paired`	are files in file_name column paired end
`fragment_length`	for single end sequencing, provide the average fragment length
`mu`	a priori CNV levels of different states, parameter passed to HMMcopy
`normal_idx`	index of samples which are assumed to be CNV free. The median of these samples serves as "normal" CNV reference level, and CNV are computed relative to this reference level. By default, QSEA looks for samples with "normal" or "control" in its name.
`plot_dir`	If provided, detail CNV plots for each chromosome and each sample are created in the provided directory

MeDIP	If set TRUE, QSEA assumes that provided files are methylation enriched sequencing data. In this case, only fragments without CpG dinucleotides are considered. This option allows QSEA to infer CNV information from MeDIP or MDB seq experiments directly
parallel	Switch for parallel computing, using BiocParallel

Value

The qseaSet object, extended by the CNV information

Author(s)

Mathias Lienhard

See Also

HMMsegment

Examples

```
library("BSgenome.Hsapiens.UCSC.hg19")

bam_hESCs_1 = system.file("extdata",
    "hESCs.MeDIP.Rep1.chr22.bam", package="MEDIPSData")
bam_hESCs_2 = system.file("extdata",
    "hESCs.MeDIP.Rep2.chr22.bam", package="MEDIPSData")
sample_table=data.frame(sample_name=paste0("hESCs_", 1:2),
    file_name=c(bam_hESCs_1,bam_hESCs_2),
    group=rep("hESC",2), stringsAsFactors=FALSE)
qseaSet=createQseaSet(sampleTable=sample_table,
        BSgenome="BSgenome.Hsapiens.UCSC.hg19",
        chr.select="chr22",
        window_size=500)

#this is an example for computing CNVs from MeDIP data. A very limited example
#however, since the samples do not contain CNVs.
qseaSet=addCNV(qseaSet, fragment_length=300, file_name="file_name", MeDIP=TRUE,
window_size=1000000)
```

---

addContrast *fit GLMs to reduced model and test for significance*

Description

This function fits negative binomial GLMs to reduced models defined either by the "contrast" parameter, or by one or several model coefficients (specified by "coef" parameter) set to zero. Subsequently, a likelihood ratio test is applied, to identify windows significantly dependent on the tested coefficient.

Usage

```
addContrast(qs,glm,contrast,coef,name,verbose=TRUE, nChunks = NULL,
    parallel = FALSE )
```

Arguments

qs	a qseaSet object
glm	a qseaGLM object
contrast	numeric vector specifying a contrast of the model coefficients. This contrast can for example be defined using limma::makeContrasts()
coef	alternatively defines the contrast by coefficient(s) of the model tested to be equal to zero.
name	short descriptive name for the contrast (as "TvsN"), used for examples in columns of result tables
verbose	more messages that document the process
nChunks	fit GLMs in multiple chunks
parallel	use multicore processing

Value

This function returns the qseaGLM object, extended by the fitted coefficients of the reduced GLMs, as well as the test statistics. Note that one qseaGLM object can contain several contrasts.

Author(s)

Mathias Lienhard

See Also

limma::makeContrasts(), fitNBglm(), isSignificant()

Examples

```
qs=getExampleQseaSet()
design=model.matrix(~group, getSampleTable(qs))
TvN_glm=fitNBglm(qs, design, norm_method="beta")
TvN_glm=addContrast(qs,TvN_glm, coef=2, name="TvN")
```

addCoverage *Import sequencing data*

Description

This function imports the alignment files (in sam/bam format) and counts the reads per genomic window or directly imports coverage files (in wiggle/bigwiggle format)

Usage

```
addCoverage(qs, fragment_length, uniquePos=TRUE, minMapQual=1, paired=FALSE,
parallel=FALSE)
```

Arguments

qs	qseaSet object, e.g. produced by the createQseaSet() function
fragment_length	For single end data, provide the expected fragment length
paired	If set to TRUE, data is considered to be paired end sequencing, and the actual fragments size is used.
uniquePos	If set to TRUE, fragments with same position and orientation are considered to be PCR duplicates and replaced by one representative.
minMapQual	The minimal mapping quality for reads to be considered. Note that the definition of mapping quality depends on the alignment tool.
parallel	Switch for parallel computing, using BiocParallel

Details

The coverage is imported from the files specified in the file_name column of the sample table, provided for the createQseaSet() function. In case of alignment files, the reads are counted for the window at the center of the sequencing fragment. For single end data, Filetypes is detected automatically from the file suffix.

Value

The function returns the qseaSet object, extended by the number of reads per window for all samples

Author(s)

Mathias Lienhard

See Also

crateQseaSet

Examples

```
library("BSgenome.Hsapiens.UCSC.hg19")

bam_hESCs_1 = system.file("extdata",
    "hESCs.MeDIP.Rep1.chr22.bam", package="MEDIPSData")
bam_hESCs_2 = system.file("extdata",
    "hESCs.MeDIP.Rep2.chr22.bam", package="MEDIPSData")
sample_table=data.frame(sample_name=paste0("hESCs_", 1:2),
    file_name=c(bam_hESCs_1,bam_hESCs_2),
    group=rep("hESC",2), stringsAsFactors=FALSE)
qseaSet=createQseaSet(sampleTable=sample_table,
        BSgenome="BSgenome.Hsapiens.UCSC.hg19",
        chr.select="chr22",
        window_size=500)
qseaSet=addCoverage(qseaSet, fragment_length=300)
```

---

addEnrichmentParameters

Enrichment analysis

Description

This function analyses the dependency of enrichment on a sequence pattern, based on a subset of windows for which the signal is known.

Usage

```
addEnrichmentParameters(qs, enrichmentPattern, signal, windowIdx,
min_wd=5,bins=seq(.5,40.5,1))
```

Arguments

qs	The qseaSet object
enrichmentPattern	The name of the pattern, on which the enrichment depends on (usually CpG for methylation analysis). This name must correspond to the name specified in addPatternDensity()
windowIdx	vector of window indices, for which "true" values are known (or can be estimated)
signal	Matrix containing the known (or estimated) values for all samples and all specified windows, as a numeric matrix. These values are expected to be between 0 and 1.
bins	For the enrichment analysis, windows are binned according to pattern density. This parameter specifies the bins.
min_wd	minimal number of windows per bin to be considered

Value

The function returns the qseaSet object, extended by the parameters of the enrichment profiles for all samples

Author(s)

Mathias Lienhard

See Also

plotEnrichmentProfile, addPatternDensity

Examples

```
qs=getExampleQseaSet(enrichmentAnalysis=FALSE)
#this procedure assumes that regions with low CpG density is 80% methylated
#on average, and regions within CpG islands are 25% methylated on average.
wd=which(getRegions(qs)$CpG_density>1 &
    getRegions(qs)$CpG_density<15)
signal=(15-getRegions(qs)$CpG_density[wd])*.55/15+.25
signal=matrix(signal,nrow=length(signal),ncol=length(getSampleNames(qs)))
qs=addEnrichmentParameters(qs, enrichmentPattern="CpG",
    windowIdx=wd, signal=signal)
```

---

addLibraryFactors *Estimate effective library size*

Description

Normalization factors for effective library size are computed using the trimmed mean of m-values approach (TMM).

Usage

```
addLibraryFactors(qs, factors,...)
```

Arguments

qs	The qseaSet object
factors	In case normalization factors have been pre-computed by the user, they can be passed with this parameter. In this case QSEA adds this factors to the qseaSet object and does not compute normalization factors.
...	Further parameters used for the TMM normalization (see details)

Details

The user can specify the TMM normalization by setting the following additional parameters, which are passed to the internal functions. \trimA [default: c(.5,.99)] lower and upper quantiles for trimming of A values \trimM [default: c(.1,.9)] lower and upper quantiles for trimming of M values \doWeighting [default: TRUE] computes a weighted TMM \ref [default: 1] the index of the reference sample \plot [default: FALSE] if set to TRUE, MvsA plots depicting the TMM normalization are created. \nReg [default: 500000] Number of regions to be analyzed for normalization. Regions are drawn uniformly over the whole genome.

Value

This function returns the qseaSet object, containing effective library size normalization factors.

Author(s)

Mathias Lienhard

See Also

edgeR::calcNormFactors

Examples

```
qs=getExampleQseaSet(expSamplingDepth=500*10^(1:5), repl=5)
#in this case, the first sample has only view reads, so it is important to set
#the reference sample
qs=addLibraryFactors(qs, plot=TRUE, ref="Sim5N")
```

---

addNewSamples *Extends an exisiting qseaSet by new samples*

Description

This function allows the qseaSet to be extended by new samples, provided in the sample table.

Usage

```
addNewSamples(qs, sampleTable, force=FALSE, parallel=FALSE)
```

Arguments

qs	The qseaSet object to be extended
sampleTable	data.frame, describing the samples. Must be in same format as getSampleTable(qs)
force	force adding of new samples, even if existing CNV or enrichment information requires recomputation
parallel	parallel processing of alignment files

Value

An object of class qseaSet, including the new samples.

Author(s)

Mathias Lienhard

Examples

```
library("BSgenome.Hsapiens.UCSC.hg19")
data(samplesNSCLC, package="MEDIPSData")
path=system.file("extdata", package="MEDIPSData")
samples_NSCLC$file_name=paste0(path,"/",samples_NSCLC$file_name )
originalQseaSet=createQseaSet(sampleTable=samples_NSCLC[1:4,],
        BSgenome="BSgenome.Hsapiens.UCSC.hg19", chr.select="chr22",
        window_size=500)
originalQseaSet=addCoverage(originalQseaSet, uniquePos=TRUE, paired=TRUE)
qseaSet=addNewSamples(originalQseaSet, samples_NSCLC)
```

---

addOffset *Estimate background reads*

Description

This function sets the background reads offset parameters for the qseaSet object, either by estimating offset reads, or by setting user provided values.

Usage

```
addOffset(qs,enrichmentPattern , maxPatternDensity=0.01,offset)
```

Arguments

qs	the qseaSet object
enrichmentPattern	name of the enrichment pattern, as specified in addPatternDensity
maxPatternDensity	Maximum pattern density, at which the window is treated as pattern free.
offset	This parameter alternatively allows to specify the amount of background reads for each sample manually. In this case, please provide average background reads for CNV free windows in rpkm scale.

Value

The function returns the qseaSet object, extended by the estimated amount of background reads for all samples

Author(s)

Mathias Lienhard

See Also

addPatternDensity, getOffset

Examples

```
#simulate data with varing background fractions
qs=getExampleQseaSet(expSamplingDepth=5e4, repl=5,bgfraction=seq(0,.8,.2))
#estimate the background in simulated data
addOffset(qs, "CpG", maxPatternDensity=0.7)
#return the background on different scales
getOffset(qs, scale="fraction") #estimated fraction of total reads
getOffset(qs, scale="rpw") #average background reads per CNV free window
```

---

addPatternDensity *Infer sequence pattern density values and add to qseaSet object*

Description

This function estimates the average occurrences of a sequence pattern (such as CpG dinucleotides) within the overlapping sequencing fragments for each genomic window

Usage

```
addPatternDensity(qs, pattern,name, fragment_length, fragment_sd,
patternDensity, fixed=TRUE, masks=c("AGAPS","AMB", "RM", "TRF")[1:2])
```

Arguments

qs	a qseaSet object
pattern	actual sequence of the pattern (e.g. "CG")
,	
name	a name for the sequence pattern(e.g. "CpG")
,	
fragment_length	the average fragment length to be assumed for pattern density estimation. If omitted, this parameter is taken from the qseaSet object.
fragment_sd	the standard deviation of fragment length to be assumed for pattern density estimation. If omitted, this parameter is taken from the qseaSet object.

patternDensity	this parameter alternatively allows to specify the pattern density manually. In this case, please provide a numerical vector, containing a value (greater than 0) for each genomic window.
fixed	if FALSE, an IUPAC ambiguity code in the pattern can match any letter in the reference genome that is associated with the code, and vice versa.
masks	names of the BSgenome masks to be active.

Value

The function returns the qseaSet object, extended by the pattern density for all genomic windows

Author(s)

Mathias Lienhard

Examples

```
library("BSgenome.Hsapiens.UCSC.hg19")
bam_hESCs_1 = system.file("extdata",
    "hESCs.MeDIP.Rep1.chr22.bam", package="MEDIPSData")
bam_hESCs_2 = system.file("extdata",
    "hESCs.MeDIP.Rep2.chr22.bam", package="MEDIPSData")
sample_table=data.frame(sample_name=paste0("hESCs_", 1:2),
    file_name=c(bam_hESCs_1,bam_hESCs_2),
    group=rep("hESC",2), stringsAsFactors=FALSE)
qseaSet=createQseaSet(sampleTable=sample_table,
        BSgenome="BSgenome.Hsapiens.UCSC.hg19",
        chr.select="chr22",
        window_size=500)
qseaSet=addPatternDensity(qseaSet, "CG", name="CpG", fragment_length=300)
```

---

addSeqPref *Add sequence preference to qseaSet object*

Description

This function allows to add window specific sequencing preference, that can be used by the normalization procedure. This preference can be defined by the user, or estimated from sequencing of input libraries.

Usage

```
addSeqPref(qs, seqPref,file_name, fragment_length, paired=FALSE,
uniquePos=TRUE, alpha=0.05, pseudocount=5, cut=3)
```

Arguments

qs	a qseaSet object
seqPref	A vector with predefined sequencing preference for each window. Values are interpreted as log2 ratios relative to normal/average sequencing preference.
file_name	alternatively, the sequencing preference can be estimated from input sequencing. In this case, provide the column of the sample table that contains the file names for input sequencing alignment or coverage files.
fragment_length	for single end data, provide the expected fragment length
paired	if set to TRUE, data is considered to be paired end sequencing, and the actual fragments size is used.
uniquePos	if set to TRUE, fragments with same position and orientation are considered to be PCR duplicates and replaced by one representative.
alpha	currently ignored
pseudocount	this value is added to the coverage of each window, to smooth the estimates.
cut	absolute log2 value threshold for windows to be excluded from later analysis due to extreme preference values.

Value

the function returns the qseaSet object, extended by the sequencing preference for all genomic windows.

Author(s)

Mathias Lienhard

---

createQseaSet *Prepares a qseaSet Object*

Description

This method prepares the qseaSet object, and prepares genome wide bins. Coverage and normalization parameters are added in succeeding functions.

Usage

```
createQseaSet(sampleTable,BSgenome, chr.select,Regions, window_size=250 )
```

Arguments

BSgenome	name of BSgenome package
Regions	GRanges object. If specified, only selected regions are processed
chr.select	If specified, only selected chromosomes are processed
sampleTable	data.frame, containing at least 3 columns: the sample names (sample_name), paths to alignment or coverage file in sam/bam/wiggle/bigwig format (file_name), and one or more test condition(s) (group). Optionally it may contain a column with alignment or coverage files for CNV analysis, and further information in the samples that are of interest for the analysis.
window_size	size for the genome wide bins in base pairs

Value

An object of class qseaSet, containing the sample and genome information.

Author(s)

Mathias Lienhard

Examples

```
library("BSgenome.Hsapiens.UCSC.hg19")
bam_hESCs_1 = system.file("extdata", "hESCs.MeDIP.Rep1.chr22.bam",
    package="MEDIPSData")
bam_hESCs_2 = system.file("extdata", "hESCs.MeDIP.Rep2.chr22.bam",
    package="MEDIPSData")
samplesTable=data.frame(sample_name=paste0("hESCs_", 1:2),
file_name=c(bam_hESCs_1,bam_hESCs_2),
group=rep("hESC",2),stringsAsFactors=FALSE)
qs=createQseaSet(samplesTable, BSgenome="BSgenome.Hsapiens.UCSC.hg19",
    chr.select="chr22", window_size=500)
```

---

fitNBglm *Fit GLM for each window*

Description

This function fits a negative binomial GLM for each genomic window, according to the design matrix.

Usage

```
fitNBglm(qs,design,link="log",keep, disp_method="region_wise",
norm_method="rpkm",init_disp=0.5 ,verbose=TRUE, minRowSum=10, pseudocount=1,
    disp_iter = 3, nChunks = NULL, parallel = FALSE)
```

Arguments

qs	a qseaSet object
design	the design matrix for the GLMs
link	name of the link function. Currently, only the canonical dQuotelog link function is implemented.
keep	indices of windows to be included in the analysis.
disp_method	method to estimate dispersion parameters. Allowed values are dQuoteregion_wise for independent window wise estimates, dQuotecommon for a single estimate for all windows, dQuotecutAtQuantiles for window wise estimates trimmed at the 25% and 75% quantiles, or dQuoteinitial for using the dispersion parameters provided with the init_disp parameter.
norm_method	normalization method, as defined by normMethod() function
init_disp	initial estimate for dispersion parameter. Either a single parameter for all regions, or a vector with window wise parameters.
verbose	more messages that document the process
minRowSum	filter out windows with less than minRowSum reads over all samples
pseudocount	this value is added to the read counts
disp_iter	number of iterations for dispersion estimation
nChunks	fit GLMs in multiple chunks
parallel	use multicore processing

Value

This function returns a qseaGLM object, containing the fitted coefficients of the GLMs.

Author(s)

Mathias Lienhard

See Also

addContrast()

Examples

```
#tbd
qs=getExampleQseaSet()
design=model.matrix(~group, getSampleTable(qs))
TvN_glm=fitNBglm(qs, design, norm_method="beta")
```

`getExampleQseaSet` *Simulation of MeDIP seq QSEA set*

Description

Creates a example qseaSet object by sampling reads for simulated Tumor and Normal samples. Number of replicates, sequencing depth and fraction of background reads can be specified.

Usage

```
getExampleQseaSet(CpG=TRUE,CNV=TRUE,repl=2,
    doSampling=TRUE,enrichmentAnalysis=TRUE, expSamplingDepth=50000,
    bgfraction=.1)
```

Arguments

`CpG`	if TRUE CpG density is added to the object
`CNV`	if TRUE CNV are emulated for the tumor samples
`repl`	number of replicates for tumor and normal samples
`doSampling`	if TRUE, read counts are sampled and added to the object
`enrichmentAnalysis`	if TRUE, parameters for enrichment profiles are added
`expSamplingDepth`	expected value of sequencing depth
`bgfraction`	fraction of background reads

Details

The function creates an example and test qseaSet object for an toy example genome (one chromosome, 50kb) with 500 bases windows.

Value

The qseaSet object

Author(s)

Mathias Lienhard

See Also

createQseaSet()

Examples

```
qs=getExampleQseaSet()
```

getPCA *Principle Component Analysis (PCA) in QSea*

Description

The getPCA() function performs a Principle Component Analysis (PCA) of the coverage profiles from a qsea object for exploratory data analysis.

Usage

```
getPCA(qs, chr= getChrNames(qs),ROIs, minRowSum=20, keep ,
    norm_method=normMethod(logRPM =
    c("log", "library_size", "cnv", "preference", "psC10")), topVar=1000,
    samples=getSampleNames(qs), minEnrichment = 0)
```

Arguments

qs	DIPSset (mandatory)
chr	chromosomes to consider
ROIs	If specified, only windows overlapping ROIs are considered.
minRowSum	minimal number of total read counts per window over all samples
keep	windows to consider
norm_method	name of predefined normalization (e.g. "beta"), or user defined normalization by calling normMethod() function
topVar	only the top variable windows are considered
samples	names of samples to be considered
minEnrichment	for transformation to absolute methylation level, you can specify the minimal number of expected reads for a fully methylated window. This avoids inaccurate estimates, due to low enrichment.

Details

The principle component analysis is calculated using the singular value decomposition (svd).

Value

getPCA() returns a list object, containing the svd and information on the selected windows.

Author(s)

Mathias Lienhard

See Also

plotPCA

Examples

```
qs=getExampleQseaSet( repl=5)
pca=getPCA(qs, norm_method="beta")
colors=c(rep("red", 5), rep("green", 5))
plotPCA(pca, bgColor=colors)
#plotPCAfactors is more interesting, if ROIs have been specified in getPCA
plotPCAfactors(pca)
```

---

isSignificant *Finds Significant Regions*

Description

This function looks for regions, where the test statistic is below the defined thresholds

Usage

```
isSignificant(glm, contrast = NULL, fdr_th = NULL, pval_th = NULL,
            absLogFC_th = NULL, direction = "both")
```

Arguments

`glm`	A qseaGLM object (mandatory)
`contrast`	name of contrast to be used
`fdr_th`	a threshold for the false discovery rate
`pval_th`	a p value threshold
`absLogFC_th`	the threshold for the absolute value of logFC
`direction`	direction of change: either "both", "loss", or "gain"

Details

If a threshold is NULL, it is ignored.

For the direction parameter, the following synonyms are valid:

"loss" == "less" == "hypo"

"gain" == "more" == "hyper"

Value

A vector with indices of significant windows, which can be passed to keep parameter of makeTable() function

Author(s)

Mathias Lienhard

See Also

makeTable

Examples

```
qs=getExampleQseaSet()
design=model.matrix(~group, getSampleTable(qs))
TvN_glm=fitNBglm(qs, design, norm_method="beta")
TvN_glm=addContrast(qs,TvN_glm, coef=2, name="TvN")
sig=isSignificant(TvN_glm, fdr_th=0.01)
```

---

makeTable *Create a Results Table*

Description

This function creates a table from the qsea objects qseaSet and qseaTvN_glm

Usage

```
makeTable(qs,glm,norm_methods="counts",samples,groupMeans, keep, ROIs,
    annotation, minPvalSummarize, CNV=FALSE, verbose=TRUE, minEnrichment=3,
    chunksize=1e5)
```

Arguments

qs	a qseaSet object (mandatory)
glm	a list of one or more qseaGLM objects (optional)
norm_methods	ether a character vector of pre-defined normalization combinations, or a list defining normalization combinations. This affects both individual and mean values.
samples	The indices of the samples for which individual values are to be written out in the specified order
groupMeans	a named list of indices vectors, defining groups for which mean values are to be written out
keep	a vector of indices of the windows that are considered (as created by isSignificant)
ROIs	A GRanges object, containing regions of interest (ROIs). Only windows overlapping ROIs are considered.
annotation	a named list of GRange objects, containing annotations (e.g. genes, CpG islands, ...) that are added to the table.
minPvalSummarize	If ROIs are given, you can specify a QseaTvN_glm object. For each ROI the window with the most significant differential coverage is written out

CNV	If set TRUE, the CNV logFC for the samples specified by samples are written out.
verbose	verbosity level
minEnrichment	for transformation to absolute methylation level, you can specify the minimal number of expected reads for a fully methylated window. This avoids inaccurate estimates, due to low enrichment.
chunksize	For efficient memory usage, the table is built up in chunks. With this parameter, the maximum number of windows processed in one chunk is specified.

Details

Note that, if overlapping ROIs are specified, windows might emerge in the table several times.

Value

A result table containing the specified normalized values for the selected windows and samples/groups

Author(s)

Mathias Lienhard

See Also

isSignificant

Examples

```
#create example set
qs=getExampleQseaSet()
design=model.matrix(~group, getSampleTable(qs))
TvN_glm=fitNBglm(qs, design, norm_method="beta")
TvN_glm=addContrast(qs,TvN_glm, coef=2, name="TvN")
sig=isSignificant(TvN_glm, fdr_th=0.01)

##Table containing all significant windows
tab1=makeTable(qs=qs, glm=TvN_glm,
    keep=sig, samples=getSampleNames(qs))
##additional CNV logFC for the selected samples
tab2=makeTable(qs=qs, glm=TvN_glm,
    keep=sig, samples=getSampleNames(qs), CNV=TRUE)
##explicit selection of normalization:
##counts (i.e. no normalization, only counts)
tab3=makeTable(qs=qs, glm=TvN_glm, keep=sig,
    samples=getSampleNames(qs), norm_method="counts")

##counts AND %methylation values for individual samples and group means
tab4=makeTable(qs=qs, glm=TvN_glm, keep=sig,
    samples=getSampleNames(qs), groupMeans=getSampleGroups(qs),
    norm_method=c("counts", "beta"))
```

`normMethod` *Definition of normalization procedure*

Description

This function allows to define normalization methods by specifying components.

Usage

```
normMethod(methods, ...)
```

Arguments

`methods`	names of predefined normalization methods (for a list of predefined methods, see details)
`...`	sets of normalization components, that can be combined to user defined normalization methods

Details

Predefined normalization methods:

"counts": no normalization, simply raw count values

"reads": same as counts

"rpm": reads per million mappable reads

"nrpm": CNV normalized reads per million mappable reads

"beta": transformation to % methylation, posterior mean point estimator

"logitbeta": logit transformed beta values

"betaLB": 2.5 lower bound for the point estimator

"betaUB": 97.5 upper bound for the point estimator

Allowd components for user defined normalization methods:

"library_size": scale by effective library size

"region_length": scale by window size

"preference": scale by positional sequencing preference

"cnv": scale by CNV ratio

"enrichment": use enrichment profiles for transformation to absolute methylation level

"qXY": quantile estimator for transformation to absolute methylation level. XY must be replaced by the quantile (see example with self defined lower and upper bound)

"offset": consider background reads

WARNING: not all combinations are allowed (eg qXY requires enrichment) and not all allowed combinations are meaningful. Inexperienced users should stick to predefined normalization methods.

Value

a list object, containing the components for the specified normalization procedure

Author(s)

Mathias Lienhard

See Also

makeTable

Examples

```
#simply raw counts
nm=normMethod("counts")
#beta-values (% methylation) including lower and upper bounds
nm=normMethod(c("beta", "betaLB", "betaUB"))
#self defined lower and upper bound: 10% and 90% quantile
nm=normMethod("beta",
betaLB_10=c("enrichment", "cnv", "library_size",
    "region_length", "preference","q10", "offset"),
betaUB_90=c("enrichment", "cnv", "library_size",
    "region_length", "preference","q90", "offset")
)
```

---

plotCNV *Plots a Heatmap-like Overview of the CNVs*

Description

This function plots the Copy Number Variations (CNVs) of the samples in a heatmap like representation. Amplified regions are depicted in red, whereas deletions are depicted green, and CNV free regions blue. The samples are ordered by an hierarchical clustering.

Usage

```
plotCNV(qs, dist = c("euclid", "cor")[1], clust_method = "complete",
            chr = getChrNames(qs), samples =getSampleNames(qs),
            cex = 1, labels = c(TRUE, TRUE,TRUE, TRUE), naColor = "darkgrey",
            indicateLogFC = TRUE )
```

Arguments

qs	a qseaSet object (mandatory)
dist	distance measure for clustering. dQuoteeuclidian or dQuotecorrelation based (1-cor)
clust_method	method to be passed to hclust

chr	vector of chromosomes to be depicted
samples	samples for which CNVs are depicted
cex	font size of labels
labels	Boolean vector of length four (bottom, left, top, right), specifying the sides of the map to be labeled
naColor	Color for regions without CNV information
indicateLogFC	indicate the CNV logFC values in the legend

Value

This function returns the pairwise distances of the CNV profiles, on which the clustering is based on.

Author(s)

Mathias Lienhard

Examples

```
qs=getExampleQseaSet()
plotCNV(qs, labels=c(FALSE, TRUE, TRUE, FALSE))
```

---

plotCoverage *Plots a genome-browser-like image of a region*

Description

This function plots the normalized coverage of specified samples in a specified region, together with annotations, in a genome-browser-like fashion

Usage

```
plotCoverage(qs,test_results, chr, start, end, samples,samples2,
norm_method="nrpkm", yoffset, xlab="Position",
ylab="MeDIP seq", col="black", main, reorder="non", indicate_reorder=TRUE,
distfun=dist, clustmethod="complete", scale=TRUE, steps=TRUE, space=0.05,
baselines=TRUE, scale_val, scale_unit=NULL, logFC_pc=.1, cex=1, smooth_width,
smooth_function=mean, regions, regions_lwd=1, regions_col,
regions_offset, regions_names, regions_dash=0.1)
```

Arguments

qs	a qseaSet object
chr	the chromosome of the region to be depicted
start	the start position of the region to be depicted
end	the end position of the region to be depicted
samples	the indices of the samples to be depicted
samples2	if specified, used to calculated logFC (samples/samples2) profiles, must be of same length as samples
logFC_pc	if samples2 is specified and logFC are calculated, this parameter specifies the pseudocount to avoid division by zero
norm_method	a vector of normalization methods to be combined
yoffset	horizontal offset, used to adjust the space between the profiles
xlab	title for the x axis
ylab	title for the y axis
main	an overall title for the plot
col	color vector for the samples (is recycled)
reorder	indicate whether, and if yes how, the samples are reordered. Valid values are "non", "clust", "max", "minP", or a genomic position within the range that is depicted
test_results	a qseaGLM object, used to find the region with minimal p value (only if reorder="minP")
indicate_reorder	indicate the window that has been used for reordering by an arrow.
distfun	if reorder="clust": for hierarchical clustering for reordering
clustmethod	if reorder="clust": for hierarchical clustering for reordering
scale	if set TRUE, print a bar scale
scale_val	length of the bar scale
scale_unit	unit of the bar scale
steps	plot the coverage as step function (steps=TRUE), or as lines
space	fraction of the plot set aside for sample names etc.
baselines	depict the baselines (zero) of the coverage profiles
cex	font size
smooth_width	number of windows to be considered for sliding window smoothing
smooth_function	function to be applied on the sliding windows for smoothing
regions	named list of GenomicRanges objects, containing annotation (eg exons) to be depicted below the coverage profiles
regions_lwd	vector of line width for the
regions_col	vector of colors for the regions
regions_offset	offset value, defining the space between the regions
regions_names	vector of column names, that store the names of the regions
regions_dash	vector, specifying the length of the end dashes of the regions

Value

list containing a table containing the plotted coverage values, the position that has been used for ordering, and the image coordinates

Author(s)

Mathias Lienhard

Examples

```
qs=getExampleQseaSet(repl=5)
colors=c(rep("red", 5), rep("green", 5))
plot(1)
plotCoverage(qs,samples=getSampleNames(qs),
    chr="chr1", start=1960001, end=1970001,col=colors,
    norm_method="beta", yoffset=1,space=.2, reorder=1964500)
plotCoverage(qs,samples=getSampleNames(qs),
    chr="chr1", start=1960001, end=1970001,col=colors,
    norm_method="beta", yoffset=1,space=.2, reorder="clust")
```

---

plotEnrichmentProfile *Plotting functions for enrichment profiles*

Description

Plots the estimated sequence pattern dependent enrichment profile for one or several samples as a matrix of plots

Usage

```
plotEnrichmentProfile(qs,sample, sPoints=seq(0,30,1),
    fitPar=list(lty=2, col="green"),cfPar=list(lty=1), densityPar, meanPar,... )
plotEPmatrix(qs, sa=getSampleNames(qs),nrow=ceiling(sqrt(length(sa))),
    ncol=ceiling(length(sa)/nrow), ...)
```

Arguments

qs	The qseaSet object
sample	The index of the sample for which the enrichment profile should be depicted
sPoints	The values at which the enrichment profile function is evaluated
fitPar	List of parameters for depiction of the fitted enrichment profile function (see details)
cfPar	List of parameters for depiction of the empirical enrichment profile (see details)
densityPar	List of parameters for depiction high density scatterplot of coverage and pattern density (see details)

meanPar	List of parameters for depiction of the mean coverage per pattern density bin (see details)
sa	vector of samples to be depicted in matrix plot
nrow	number of rows in matrix plot
ncol	number of columns in matrix plot
...	Further graphical parameters may also be supplied

Details

Parameter lists for lines in the plot (e.g. fitPar, cfPar and meanPar) are passed to graphics::lines(), densityPar are passed to graphics::smoothScatter() function.

Value

plotEnrichmentProfile returns the coordinates of the enrichment profile. plotEPmatrix returns enrichment profile coordinates for all depicted samples.

Author(s)

Mathias Lienhard

See Also

addEnrichmentParameters

Examples

```
#create example object with different sequencing depth
qs=getExampleQseaSet(expSamplingDepth=50*10^(1:4), repl=4)
#enrichment profile for one sample
plotEnrichmentProfile(qs, "Sim4T")
#enrichment profile for all samples
plotEPmatrix(qs)
```

---

plotPCA *Plots for Principle Component Analysis (PCA) in QSEA*

Description

The principle components can be depicted using the plotting methods plotPCA and plotPCAfactors

Usage

```
## S4 method for signature 'qseaPCA'
plotPCA(object,plotComponents=c(1,2), fgColor="black",
    bgColor = "white", legend, plotLabels=TRUE, radius=5, labelOffset=.5,
    labelPos=1, labelAdj, labelColor="black", cex=1, ...)

## S4 method for signature 'qseaPCA'
plotPCAfactors(object,plotComponents=c(1,2),
    fgColor="black",bgColor = "white", plotTopLabels=100, labelsOfInterest,
    radius=1, labelOffset=.5,labelPos=1,labelColor="black", cex=1, ...)
```

Arguments

`object`	the qseaPCA object, resulting from the getPCA function
`plotComponents`	vector of the two components of the PCA
`fgColor`	vector of foreground colors for the circles
`bgColor`	vector of background colors for the circles
`legend`	add a legend to the plot
`plotLabels`	if set TRUE, the labels of the samples are written in the plot
`radius`	defines the size of the plotted circles
`labelOffset`	defines the offset of the labels to the circles
`labelPos`	specify position of the labels in the plot (see graphics::text)
`labelAdj`	alternative way to specify position of the labls in the plot (see graphics::text)
`labelColor`	a vector of colors for the labels
`cex`	font size of the labels
`plotTopLabels`	labels of factors with strongest contribution to plotted components are shown
`labelsOfInterest`	vector of factor names that are highlighted and labeled in the plot
`...`	further graphical parameters

Value

The functions return a list with the coordinates of the depicted components

Author(s)

Mathias Lienhard

See Also

plotPCA

Examples

```
qs=getExampleQseaSet( repl=5)
pca=getPCA(qs, norm_method="beta")
colors=c(rep("red", 5), rep("green", 5))

plotPCA(pca, bgColor=colors)
#plotPCAfactors is more interesting, if ROIs have been specified in getPCA
plotPCAfactors(pca)
```

`qseaGLM-class` *qseaGLM class and its methods*

Description

The qseaGLM class is used in qsea to store fitted coefficients of the GLM.

Slots

`fullModelDesign`: design matrix of full model

`fullModel` : list containing parameters and fitted coefficients of full model

`parameters`: list of parameters used to create the object

`contrast`: list of lists containing parameters and the fitted model coeficients of the reduced models

`windows`: vector of window indices, for which GLMs have been fitted

Author(s)

Matthias Lienhard

Examples

```
showClass("qseaGLM")
```

`qseaPCA-class` *qseaPCA class and its methods*

Description

The qseaPCA class is used in qsea to store results of the principle component analysis.

Slots

`svd`: singular value decomposition

`sample_names` : names of the samples

`factor_names`: names of the genomic windows involved

Author(s)

Matthias Lienhard

Examples

```
showClass("qseaPCA")
```

---

qseaSet-class *qseaSet class and its methods*

Description

The qseaSet class is used in qsea to store information about the coverage, the dependent organism, the chromosomes included in the input file, the length of the included chromosomes (automatically loaded), the number of regions, and optionally CNV information.

Slots

`sampleTable`**:** Object of class `"data.frame"`: the sample table

`count_matrix`**:** Object of class `"matrix"`: matrix containing the coverage for all samples

`zygosity`**:** Object of class `"matrix"`: matrix containing the zygosity for all chromosomes and all samples

`regions`**:** Object of class `"GenomicRanges"`: the genomic regions for the coverage matrix

`parameters`**:** Object of class `"list"`: the parameter list used to create this object

`cnv`**:** Object of class `"GenomicRanges"`: CNV ranges and logFCs

`enrichment` **:** Object of class `"list"`: parameters of the sequence pattern enrichment analysis

`libraries` **:** Object of class `"matrix"`: parameters of the sequencing libraries

Methods

getSampleTable `signature(object = "qseaSet")`: extracts the sample table of a qsea set

getSampleNames `signature(object = "qseaSet")`: extracts the sample names of a qsea set

getSampleGroups `signature(object = "qseaSet")`: extracts the sample groups of a qsea set

getChrNames `signature(object = "qseaSet")`: returns the analysed chromosomes

getCounts `signature(object = "qseaSet")`: extracts the count matrix a qsea set

getRegions `signature(object = "qseaSet")`: extracts the regions object of a qsea set

getParameters `signature(object = "qseaSet")`: extracts the parameter list of a qsea set

getLibSize `signature(object = "qseaSet")`: extracts the library size (eg the total number of read counts per sample)

getNormFactors `signature(object = "qseaSet")`: extracts the list with the different normalization factors

hasCNV `signature(object = "qseaSet")`: TRUE if CNV information is present, FALSE otherwise

getCNV `signature(object = "qseaSet")`: extracts the CNV regions and logFCs

getOffset `signature(object = "qseaSet")`: extracts offset of rpkm scaled background reads

getWindowSize `signature(object = "qseaSet")`: returns the window size of the object

getZyosity `signature(object = "qseaSet")`: returns the zygosity matrix of the object

setZygosity `signature(object = "qseaSet", zygosityMatrix)`: sets the zygosity matrix, and resets CNV

Author(s)

Matthias Lienhard

Examples

```
showClass("qseaSet")
```

`regionStats` *Counts the Windows in Regions of Interest*

Description

This function takes a list of window indices and a list of ROIs and counts the number of overlapping windows

Usage

```
regionStats(qs, subsets = list(covered = which(rowSums(getCounts(qs)) >= 20)),
 ROIs = list(), minoverlap = 0, maxgap = -1)
```

Arguments

`qs`	A qsea Set object
`subsets`	A list of window indices
`ROIs`	A list of Regions of Interest
`minoverlap`	Passed to findOverlaps
`maxgap`	Passed to findOverlaps

Value

a matrix, containing the total number of windows overlapping the ROIs and the numbers of windows from the subset list overlapping ROIs

Author(s)

Mathias Lienhard

See Also

findOverlaps

Examples

```
qs=getExampleQseaSet()
#as an example, we analyze the fraction of reads covered by at least 10
#or at least 20 reads, for bins of CpG density
ROIs=list()
regs=getRegions(qs)
cpg=getRegions(qs)$CpG_density
bins=seq(0,30,5)
for(i in 1:(length(bins)-1)){
    n=paste0(bins[i],"-",bins[i+1]," CpGs")
    ROIs[[n]]=regs[which(cpg>=bins[i] & cpg < bins[i+1])]
}
subsets = list(
    ">10" = which(rowSums(getCounts(qs)) >= 10),
    ">20" = which(rowSums(getCounts(qs)) >= 20))
coverage_stats=regionStats(qs, subsets, ROIs)
coverage_stats_rel=coverage_stats[,-1]/coverage_stats[,1]
x=barplot(t(coverage_stats_rel)*100,ylab="fraction of windows[%]",
    beside=TRUE, legend=TRUE, las=2, args.legend=list(x="topleft"),
    main="Covered Windows")
```

Index

∗ **classes**
qseaGLM-class, 28
qseaPCA-class, 28
qseaSet-class, 29
∗ **package**
qsea-package, 2

addCNV, 3
addContrast, 4
addCoverage, 6
addEnrichmentParameters, 7
addLibraryFactors, 8
addNewSamples, 9
addOffset, 10
addParameters,qseaGLM-method *(qseaGLM-class)*, 28
addParameters,qseaSet-method *(qseaSet-class)*, 29
addPatternDensity, 11
addSeqPref, 12

createQseaSet, 13

fitNBglm, 14

getChrNames *(qseaSet-class)*, 29
getChrNames,qseaSet-method *(qseaSet-class)*, 29
getCNV *(qseaSet-class)*, 29
getCNV,qseaSet-method *(qseaSet-class)*, 29
getCounts *(qseaSet-class)*, 29
getCounts,qseaSet-method *(qseaSet-class)*, 29
getExampleQseaSet, 16
getLibSize *(qseaSet-class)*, 29
getLibSize,qseaSet-method *(qseaSet-class)*, 29
getNormFactors *(qseaSet-class)*, 29
getNormFactors,qseaSet-method *(qseaSet-class)*, 29
getOffset *(qseaSet-class)*, 29
getOffset,qseaSet-method *(qseaSet-class)*, 29
getParameters *(qseaSet-class)*, 29
getParameters,qseaGLM-method *(qseaGLM-class)*, 28
getParameters,qseaSet-method *(qseaSet-class)*, 29
getPCA, 17
getRegions *(qseaSet-class)*, 29
getRegions,qseaSet-method *(qseaSet-class)*, 29
getSampleGroups *(qseaSet-class)*, 29
getSampleGroups,qseaSet-method *(qseaSet-class)*, 29
getSampleNames *(qseaSet-class)*, 29
getSampleNames,qseaGLM-method *(qseaGLM-class)*, 28
getSampleNames,qseaPCA-method *(qseaPCA-class)*, 28
getSampleNames,qseaSet-method *(qseaSet-class)*, 29
getSampleTable *(qseaSet-class)*, 29
getSampleTable,qseaSet-method *(qseaSet-class)*, 29
getWindowSize *(qseaSet-class)*, 29
getWindowSize,qseaSet-method *(qseaSet-class)*, 29
getZygosity *(qseaSet-class)*, 29
getZygosity,qseaSet-method *(qseaSet-class)*, 29

hasCNV *(qseaSet-class)*, 29
hasCNV,qseaSet-method *(qseaSet-class)*, 29

isSignificant, 18

makeTable, 19

normMethod, 21

plotCNV, 22
plotCoverage, 23
plotEnrichmentProfile, 25
plotEPmatrix *(*plotEnrichmentProfile*)*, 25
plotPCA, 26
plotPCA,qseaPCA-method *(*plotPCA*)*, 26
plotPCAfactors *(*plotPCA*)*, 26
plotPCAfactors,qseaPCA-method
*(*plotPCA*)*, 26

QSEA *(*qsea-package*)*, 2
qsea *(*qsea-package*)*, 2
qsea-package, 2
qseaGLM *(*qseaGLM-class*)*, 28
qseaGLM-class, 28
qseaPCA *(*qseaPCA-class*)*, 28
qseaPCA-class, 28
qseaSet *(*qseaSet-class*)*, 29
qseaSet-class, 29

regionStats, 30

setZygosity *(*qseaSet-class*)*, 29
setZygosity,qseaSet-method
*(*qseaSet-class*)*, 29
show,qseaGLM-method *(*qseaGLM-class*)*, 28
show,qseaPCA-method *(*qseaPCA-class*)*, 28
show,qseaSet-method *(*qseaSet-class*)*, 29