Package 'rTRM'

November 12, 2026

Type Package

Title Identification of Transcriptional Regulatory Modules from Protein-Protein Interaction Networks

Version 1.50.0

Date 2015-12-25

Author Diego Diez

Depends R (>= 2.10), igraph (>= 1.0)

Imports methods, AnnotationDbi, DBI, RSQLite

Suggests RUnit, BiocGenerics, MotifDb, graph, PWMEnrich, biomaRt, Biostrings, BSgenome.Mmusculus.UCSC.mm8.masked, org.Hs.eg.db, org.Mm.eg.db, ggplot2, BiocStyle, knitr, rmarkdown

Maintainer Diego Diez <diego10ruiz@gmail.com>

Description rTRM identifies transcriptional regulatory modules (TRMs) from protein-protein interaction networks.

License GPL-3

LazyLoad yes

ByteCompile yes

VignetteBuilder knitr

biocViews Transcription, Network, GeneRegulation, GraphAndNetwork

URL https://github.com/ddiez/rTRM

BugReports https://github.com/ddiez/rTRM/issues

git_url https://git.bioconductor.org/packages/rTRM

git_branch RELEASE_3_23

git_last_commit 26a52c0

git_last_commit_date 2026-04-28

Repository Bioconductor 3.23

Date/Publication 2026-11-11

Contents

rTRM-package . . . 2
annotateFreq . . . 3
annotateModule . . . 4
annotateTRM . . . 4
biogrid_hs . . . 5
biogrid_mm . . . 5
findTRM . . . 6
getAnnotations . . . 7
getBiogridData . . . 7
getConcentricList . . . 8
getLargestComp . . . 9
getMaps . . . 9
getMatrices . . . 10
getMotifsFromEntrezgene . . . 10
getMotifsFromSymbol . . . 11
getOrthologFromMatrix . . . 11
getOrthologs . . . 12
getOrthologsFromBiomart . . . 12
getSequencesFromGenome . . . 13
getSimilarityMatrix . . . 14
getTFclass . . . 14
getTFclassFromEntrezgene . . . 15
getTFterms . . . 15
initBiomart . . . 16
layout.arc . . . 16
layout.concentric . . . 17
plotDegree . . . 17
plotGraph . . . 18
plotTRM . . . 19
plotTRMlegend . . . 20
processBiogrid . . . 20
removeVertices . . . 21
writeTRMreport . . . 21

Index **23**

`rTRM-package` *Identification transcription regulatory modules (TRMs)*

Description

This package identifies transcriptional regulatory modules (TRMs) from PPI networks.

Details

Package:	rTRM
Type:	Package
Version:	1.0
Date:	2013-02-01
License:	GPL-3

Author(s)

Diego Diez

Maintainer: Diego Diez <diego10ruiz@gmail.com>

Examples

```
getAnnotations()
```

`annotateFreq` *Annotate a graph with frequency of nodes/edges in other graphs.*

Description

Returns an annotated graph with node size and edge width proportional at the number of occurrences of nodes/edges in a suplied list of graphs.

Usage

```
annotateFreq(g, graph_list)
```

Arguments

`g`	target graph to annotate.
`graph_list`	list of graph to extract information from.

Details

Commonly graph_list refers to a list of predicted TRMs (with findTRM) and g is the combined TRM. This function annotates the nodes/edges in g to known their frequency in the original list of graphs.

Author(s)

Diego Diez

annotateModule *Annotate a network module with information*

Description

Uses information about expression, enrichment and parent PPI network to annotate a subgraph.

Usage

```
annotateModule(g, enrich, trm, targets, ppi, exprs, tfs)
```

Arguments

g	graph to annotate in igraph format.
enrich	list of enriched transcription factors (or motifs).
trm	TRM to compare with (to identify bridges).
targets	list of target transcription factors (typically those with ChIP-seq data).
ppi	parent PPI network (to check membership of nodes).
exprs	list of entrezgene ids representing expressed genes.
tfs	

Author(s)

Diego Diez

annotateTRM *Annotate a network object with information about clusters.*

Description

This function takes a network object and includes cluster information as piecolor attribute, suitable to be plotted with plotTRM()

Usage

```
annotateTRM(g, target)
```

Arguments

g	a network object.
target	target node (from findTRM())

Author(s)

Diego Diez

biogrid_hs *Network dataset of class 'igraph'*

Description

Human protein-protein interaction (PPI) dataset from the BioGRID database release .

Usage

```
data(biogrid_hs)
```

Format

An igraph object.

Author(s)

Diego Diez

biogrid_mm *Network dataset of class 'igraph'*

Description

Mouse protein-protein interaction (PPI) dataset from the BioGRID database .

Usage

```
data(biogrid_mm)
```

Format

An igraph object.

Author(s)

Diego Diez

findTRM *Indentifies a TRM associated with a target node and one or more query nodes.*

Description

This the main function used to identify TRMs. It takes a graph object and use it to search in the neighborhood of a target node for query nodes that are separated a maximum distance (controlled by max.bridge parameter).

Usage

```
findTRM(g, target, query, method = "nsa", max.bridge = 1, extended = FALSE, strict = FALSE, type = "igrap
```

Arguments

g	the network used to identify TRMs (tipically a PPI network)
target	character variable with the name of a target node.
query	character vector with the list of query nodes.
method	method to use.
max.bridge	maximum number of nodes allowed between the target and query nodes.
extended	whether to allow distance restrictions to include both target and query nodes.
strict	whether to return a single component (using decompose.graph())
type	type of graph object to return, either an "igraph" (the default) or a "graphNEL"

Details

Currently only "first" and "nsa" methods are available. First is used for tests and returns the first neighborhood of the target node. Method "nsa" implements the TRM finding algorithm.

Value

A network in igraph format or NULL.

Author(s)

Diego Diez

Examples

```
# load example network.
load(system.file(package = "rTRM", "extra/example.rda"))

# define target and query nodes.
target = "N6"
query = c("N7", "N12", "N28")
```

```
# find TRM:
s = findTRM(g, target = target, query = query, method = "nsa", max.bridge = 1)
```

`getAnnotations` *Obtain the 'pwm' table fromt the database, containing PWM's annotations.*

Description

Obtain the 'pwm' table fromt the database, containing PWM's annotations.

Usage

```
getAnnotations(filter, dbname = NULL)
```

Arguments

`filter`	one or more PWM ids.
`dbname`	the location of the database (to load custom datbases).

Author(s)

Diego Diez

Examples

```
ann = getAnnotations()
```

`getBiogridData` *Downloads network data from BioGRID in TAB2 format.*

Description

This function is used to generate igraph network objects from BioGRID data. It downloads the database into a data.frame object that can be used later with processBiogrid()

Usage

```
getBiogridData(release)
```

Arguments

`release`	release of BioGRID to download.

Details

The release to download must be specified as currently there is no way to download automatically the latests release.

Value

An data.frame object.

Author(s)

Diego Diez

`getConcentricList` *Returns a list with nodes membership to be used in a graph with a concentric layout*

Description

Specify target and enriched motifs and returns a list with circle membership. This information is used by layout.concentric to position the nodes in plots.

Usage

```
getConcentricList(g, t, e, max.size = 60, order.by = "label")
```

Arguments

`g`	graph to layout (extract the nodes).
`t`	list of target nodes (will go in the center).
`e`	list of enriched nodes (will go in the periphery).
`max.size`	maximum number of nodes per layer.
`order.by`	ordering attribute for list before split.

Author(s)

Diego Diez

getLargestComp *Gets the largest connected component*

Description

Returns the largest connected component from a graph.

Usage

```
getLargestComp(g)
```

Arguments

g	an igraph object.

Author(s)

Diego Diez

getMaps *Obtain the mapping between PWM and Entrez Gene identifiers.*

Description

Obtain the mapping between PWM and Entrez Gene identifiers.

Usage

```
getMaps(filter, dbname = NULL)
```

Arguments

filter	vector of PWMs to filter results.
dbname	

Author(s)

Diego Diez

Examples

```
getMaps()
```

getMatrices *Obtain a list of PWMs.*

Description

Returns a list of PWMs, by default all the PWMs in the database. Alternativelly, filtered by the ids provided by filter.

Usage

```
getMatrices(filter, dbname = NULL)
```

Arguments

filter	list of PWMs to filter results.
dbname	

Author(s)

Diego Diez

Examples

```
pwms = getMatrices()
```

getMotifsFromEntrezgene *Retrieve PWMs associated with genes provided as entrezgene identifiers.*

Description

Retrieve PWMs associated with genes provided as entrezgene identifiers.

Usage

```
getMotifsFromEntrezgene(e, organism)
```

Arguments

e	vector of entrezgene identifiers to retrieve exiting PWMs.
organism	target organism.

Author(s)

Diego Diez

`getMotifsFromSymbol` *Retrieve PWMs associated with genes provided as symbol.*

Description

Retrieve PWMs associated with genes provided as symbol.

Usage

```
getMotifsFromSymbol(s, organism)
```

Arguments

`s`	vector of gene symbols.
`organism`	target organism.

Author(s)

Diego Diez

`getOrthologFromMatrix` *Obtain gene identifiers for a target organism associated with a list of PWMs.*

Description

Obtain gene identifiers for a target organism associated with a list of PWMs.

Usage

```
getOrthologFromMatrix(filter, organism = "human", dbname = NULL)
```

Arguments

`filter`	vector of matrices to filter results.
`organism`	target organism.
`dbname`	database- usually not need to specify.

Author(s)

Diego Diez

getOrthologs *Obtain the mapping to Entrez Gene identifiers in the given organism.*

Description

Obtain the mapping to Entrez Gene identifiers in the given organism.

Usage

```
getOrthologs(filter, organism, dbname = NULL)
```

Arguments

filter	entrezgene identifiers for the original mapping (PWM to gene). These can belong to diverse species and correspond to the "entrezgene" column obtained with getMaps() function.
organism	target organisms, currently supported "human" and "mouse"
dbname	

Details

If organism is not specified the entire table of orthologs (with all supported species) is returned.

Value

A data.frame object with ortholog information.

Author(s)

Diego Diez

Examples

```
getOrthologs(organism = "human")
```

getOrthologsFromBiomart *Returns ortholog genes for a target organism*

Description

Returns ortholog genes for a target organism

Usage

```
getOrthologsFromBiomart(eg, target_org, mart)
```

Arguments

eg	list of entrezgene ids to obtain orthologs.
target_org	target organism.
mart	mart object.

Author(s)

Diego Diez

---

getSequencesFromGenome

Retrieves a set of sequences from a BSgenome object and optionally appends a label to each sequence id.

Description

This is just a wrapper to getSeq() in package Biostrings that facilitates adding a label to each sequence.

Usage

```
getSequencesFromGenome(BED, genome, append.id)
```

Arguments

BED	file with peak locations in BED format.
genome	a BSgenome object (e.g. Mmusculus)
append.id	optional label to append to each sequence id.

Author(s)

Diego Diez

`getSimilarityMatrix` *Compute similarity matrix of list of graphs.*

Description

This function computes pair-wise similarity based on common nodes (default) or edges between the graphs passed as a list.

Usage

```
getSimilarityMatrix(g_list, type = "edges")
```

Arguments

`g_list`	list of graph objects.
`type`	type of similarity, either node or edge (default).

Author(s)

Diego Diez

`getTFclass` *Return the ontology in the TFclass database associated with an entrezgene identifier*

Description

Return the ontology in the TFclass database associated with an entrezgene identifier.

Usage

```
getTFclass(dbname = NULL)
```

Arguments

`dbname`	SQLite file to use as database.

Author(s)

Diego Diez

getTFclassFromEntrezgene

Applies getTFclass sequentially to a vector of entrezgene identifiers.

Description

Applies getTFclass sequentially to a vector of entrezgene identifiers.

Usage

```
getTFclassFromEntrezgene(x, subset = "Class", tfclass, dbname = NULL)
```

Arguments

x	vector of entrezgene identifiers.
subset	level in the ontology (subset in TFclass terminology. By default "Class")
tfclass	data.frame with tfclass data to pass to the recursive function.
dbname	SQLite file to use as database.

Author(s)

Diego Diez

getTFterms

Get terms associated with a specified TFclass subset.

Description

Returns a vector of names (not ids) with the members of a particular subset in the TFclass database. By default it returns the Class subset.

Usage

```
getTFterms(subset = "Class", dbname = NULL)
```

Arguments

subset	a subset in TFclass (default Class).
dbname	SQLite file to use as database.

Author(s)

Diego Diez

initBiomart *Initializes mart objects to identify ortholog genes*

Description

Initializes mart objects to identify ortholog genes

Usage

```
initBiomart(filter, biomart = "ensembl", host)
```

Arguments

filter	list of supported organisms
biomart	
host	

Author(s)

Diego Diez

layout.arc *Layouts a graph using arcs.*

Description

Generates a layout for graphs that places in the center the target transcription factors, in the sides the enriched transcription factors and in between of them the bridge proteins.

Usage

```
layout.arc(g, target, query)
```

Arguments

g	the graph object to layout.
target	list of target nodes (typically target transcription factors.)
query	list of query nodes (typically enriched transcription factors.)

Value

A matrix with the x and y locations of each node in the target graph.

Author(s)

Diego Diez

layout.concentric *Generates a concentric layout for graphs*

Description

Generates a matrix with x,y coordinates for each node in a target graph, which layouts the nodes using concentric circles.

Usage

```
layout.concentric(g, concentric = NULL, radius = NULL, order.by)
```

Arguments

g	graph (igraph) to layout.
concentric	list with the components of each layer.
radius	radious of each layer.
order.by	graph attributes to order nodes by.

Author(s)

Diego Diez

plotDegree *Plot degree distribution for network nodes*

Description

Plots the degree distribution and fits a power law, returning in the legend the values of the fitted parameters.

Usage

```
plotDegree(g)
```

Arguments

g	igraph object

Author(s)

Diego Diez

plotGraph *Plot an graph in igraph format.*

Description

This function plots graphs of the class igraph.

Usage

```
plotGraph(g, layout = layout.fruchterman.reingold, mar = .5, vertex.pch = 21, vertex.cex, vertex.col, v
```

Arguments

g	a network object.
layout	graph layout, either a function or the output of a layout function.
mar	plot margin.
vertex.pch	node size.
vertex.cex	node size.
vertex.col	node line color.
vertex.bg	node background color.
vertex.lwd	node line width.
edge.col	edge color.
edge.lwd	edge line width.
edge.lty	edge line type.
label	logical; whether to plot labels.
label.col	label color.
label.cex	label expansion.
label.pos	label position.
label.offset	label offset.
adjust.label.col	whether to automatically adjust label color depending on the luminance of the node's color/s.
normalize.layout	whether to apply layout.norm (with limits xmin=-1, xmax=1, ymin=-1, ymax=1) to the layout.

Author(s)

Diego Diez

plotTRM *Plot an annotated TRM.*

Description

This function plots the output findTRM() after it has been annotated with cluster information with annotateTRM() function. Cluster membership is plotted using a pie plot.

Usage

```
plotTRM(g, layout = layout.fruchterman.reingold, mar = .5, vertex.col, vertex.cex, vertex.lwd, edge.col
```

Arguments

g	a network object with cluster information (attribute piecolor).
layout	graph layout, either a function or the output of a layout function.
mar	plot margin.
vertex.col	node color.
vertex.cex	node size.
vertex.lwd	node border line width.
edge.col	edge color.
edge.lwd	edge line width.
edge.lty	edge line type.
label	logical; whether to plot labels.
label.cex	label expansion.
label.col	label color.
label.pos	label position.
label.offset	label offset.
adjust.label.col	whether to automatically adjust label color depending on the luminance of the node's color.
normalize.layout	whether to apply layout.norm (with limits xmin=-1, xmax=1, ymin=-1, ymax=1) to the layout.

Author(s)

Diego Diez

plotTRMlegend *Plot the legend of a TRM with information about the cluster families.*

Description

This function just plots a legend with the cluster membership of the provided list of genes. The legend includes de most prominent families of each cluster and there is some name polishing as well.

Usage

```
plotTRMlegend(x, title = NULL, cex = 1)
```

Arguments

x	list of family names or igraph object.
title	title for the legend.
cex	numeric value controlling the size of the legend's text.

Author(s)

Diego Diez

processBiogrid *Process a data.frame with BioGRID data into a network for a target organism*

Description

Process a data.frame with BioGRID data into a network for a target organism.

Usage

```
processBiogrid(dblist, org = "human", simplify = TRUE, type = "physical", mimic.old = FALSE)
```

Arguments

dblist	data.frame containing the BioGRID data.
org	target organism (default: "human")
simplify	whether to eliminate redundant edges (default TRUE)
type	type of interaction (physical or genetic) to include (default: "physical")
mimic.old	mimic old behavior of processBiogrid() when interactions for multiple species could be retrieved. Used only for testing.

Value

An igraph object.

Author(s)

Diego Diez

`removeVertices` *Remove nodes from a graph and returns the largest component*

Description

Remove nodes from a graph and returns the largest component

Usage

```
removeVertices(g, filter, keep.hanging = FALSE)
```

Arguments

`g`	graph to remove nodes.
`filter`	
`keep.hanging`	(logical) whether to return the largest component or not.

Author(s)

Diego Diez

`writeTRMreport` *Export a table with TRM nodes and associated information.*

Description

This function generates a data.frame with the nodes in the provided graph and associated annotations.

Usage

```
writeTRMreport(graph, file, organism, target, query, sort.by = "symbol")
```

Arguments

graph	a graph object.
file	file name.
organism	organisms for the annotations.
target	target transcription factor.
query	query transcription factors.
sort.by	order the columns of the data.frame by (default: "symbol").

Author(s)

Diego Diez

Index

∗ **datasets**
biogrid_hs, 5
biogrid_mm, 5
∗ **package**
rTRM-package, 2

annotateFreq, 3
annotateModule, 4
annotateTRM, 4

biogrid_hs, 5
biogrid_mm, 5

findTRM, 6

getAnnotations, 7
getBiogridData, 7
getConcentricList, 8
getLargestComp, 9
getMaps, 9
getMatrices, 10
getMotifsFromEntrezgene, 10
getMotifsFromSymbol, 11
getOrthologFromMatrix, 11
getOrthologs, 12
getOrthologsFromBiomart, 12
getSequencesFromGenome, 13
getSimilarityMatrix, 14
getTFclass, 14
getTFclassFromEntrezgene, 15
getTFterms, 15

initBiomart, 16

layout.arc, 16
layout.concentric, 17

plotDegree, 17
plotGraph, 18
plotTRM, 19
plotTRMlegend, 20

processBiogrid, 20

removeVertices, 21
rTRM *(*rTRM-package*)*, 2
rTRM-package, 2

writeTRMreport, 21