Package 'splicelogic'

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Title splicelogic: differential transcripts to splice events

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Description Translate differential transcript usage results into discrete splice events.

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https://thelovelab.github.io/splicelogic

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`splicelogic-package` *splicelogic: differential transcripts to splice events*

Description

For more details on the features of splicelogic, read the vignette: `browseVignettes(package = "splicelogic")`

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See Also

Useful links:

- https://github.com/thelovelab/splicelogic
- https://thelovelab.github.io/splicelogic
- Report bugs at https://github.com/thelovelab/splicelogic/issues

create_mock_data *Create mock GRanges data for splicing event testing*

Description

Create mock GRanges data for splicing event testing

Usage

```
create_mock_data(
  n_genes = 1,
  n_tx_per_gene = 2,
  n_exons_per_tx = 5,
  coef_range = c(-1, 1)
)
```

Arguments

n_genes	Number of genes to simulate
n_tx_per_gene	Number of transcripts per gene
n_exons_per_tx	Number of exons per transcript
coef_range	Range of coefficient values to sample from

Value

A GRanges object with simulated transcripts and exons

Examples

```
# create mock data with 2 genes, 4 transcripts
# per gene, and 4 exons per transcript
gr <- create_mock_data(n_genes = 2, n_tx_per_gene = 4, n_exons_per_tx = 4)
```

find_events *Find splice events from annotated exons*

Description

Functions to find different types of alternative splicing events from preprocessed GRanges exon data. Events include skipped exon (se), included exon (ie), mutatualy exclusive exons (mxe), retained intron (ri), and alternative 5' and 3' splice sites (a5ss / a3ss).

Usage

```
find_se(gr, type = c("boundary", "over", "in"), inverse = FALSE)

find_ie(gr, type = c("boundary", "over", "in"))

find_mxe(gr, type = c("boundary", "in", "over"))

find_ri(gr)

find_a5ss(gr)

find_a3ss(gr)

find_all_events(gr, type = c("boundary", "over", "in"), verbose = TRUE)
```

Arguments

gr	A GRanges object with exon annotations, including 'tx_id', 'exon', and 'coef_col' metadata columns and preprocessed with preprocess().
type	The type of overlap to consider when identifying events.
inverse	If TRUE, identifies included exons instead of skipped exons.
verbose	If TRUE, prints progress messages. Default TRUE.

Value

A GRanges object with the detected exon ranges and the following additional metadata columns:

`event_type` The type of splicing event detected (e.g. `"se"`, `"ie"`, `"mxe"`, `"ri"`, `"a5ss"`, `"a3ss"`).

`event_tx_id` Transcript ID of the paired transcript involved in the event.

`event_estimate` DTU coefficient of the paired transcript.

`event_<col>` One column per name in `metadata(gr)$additional_columns`, prefixed with `event_`, carrying the corresponding value from the paired transcript.

`find_se()`: skipped exons

`find_ie()`: included exons

`find_mxe()`: mutually exclusive exons

`find_ri()`: retained introns

`find_a5ss()`: alternative 5' splice sites

`find_a3ss()`: alternative 3' splice sites

`find_all_events()`: all detected events

Examples

```
# make some mock data and run the function
gr <- create_mock_data(n_genes = 2, n_tx_per_gene = 4, n_exons_per_tx = 4) |>
  preprocess(coef_col = "estimate") |>
  generate_se(n_events = 1)

# this should find the skipped exon events we generated
find_se(gr, type = "boundary")

find_ie(gr, type = "boundary")


# detect mutually exclusive exons
gr_mx <- create_mock_data(
  n_genes = 2, n_tx_per_gene = 4, n_exons_per_tx = 4
) |>
  preprocess(coef_col = "estimate") |>
  generate_mxe(n_events = 1)

find_mxe(gr_mx, type = "boundary")


# detect retained introns
gr_ri <- create_mock_data(
  n_genes = 2, n_tx_per_gene = 4, n_exons_per_tx = 4
) |>
  preprocess(coef_col = "estimate") |>
  generate_ri(n_events = 1)

find_ri(gr_ri)


# detect alternative 5' splice sites
gr_a5 <- create_mock_data(
  n_genes = 2, n_tx_per_gene = 4, n_exons_per_tx = 4
) |>
  preprocess(coef_col = "estimate") |>
  generate_a5ss(n_events = 1)

find_a5ss(gr_a5)


# detect alternative 3' splice sites
gr_a3 <- create_mock_data(
  n_genes = 2, n_tx_per_gene = 4, n_exons_per_tx = 4
) |>
  preprocess(coef_col = "estimate") |>
  generate_a3ss(n_events = 1)
find_a3ss(gr_a3)


# detect all event types at once
```

```
gr_all <- create_mock_data(
  n_genes = 2, n_tx_per_gene = 4, n_exons_per_tx = 4
) |>
  preprocess(coef_col = "estimate") |>
  generate_se(n_events = 1)

find_all_events(gr_all, type = "boundary", verbose = FALSE)
```

`generate_events` *Generate mock splice events in a GRanges object*

Description

Functions to introduce specific types of alternative splicing events into mock GRanges data for testing purposes.

Usage

```
generate_se(gr, n_events = 1)

generate_mxe(gr, n_events = 1)

generate_ri(gr, n_events = 1)

generate_a5ss(gr, n_events = 1)

generate_a3ss(gr, n_events = 1)
```

Arguments

`gr`	A GRanges object with metadata columns: 'exon_rank', 'gene_id', 'tx_id', and 'estimate'.
`n_events`	Number of events to generate

Value

`generate_se()`: A GRanges object with skipped exon events introduced

`generate_mxe()`: A GRanges object with mutually exclusive exon events introduced

`generate_ri()`: A GRanges object with retained intron events introduced

`generate_a5ss()`: A GRanges object with alternative 5' splice site events introduced

`generate_a3ss()`: A GRanges object with alternative 3' splice site events introduced

Examples

```
gr <- create_mock_data(
  n_genes = 2, n_tx_per_gene = 4, n_exons_per_tx = 4
)
generate_se(gr, n_events = 1)


gr <- create_mock_data(
  n_genes = 2, n_tx_per_gene = 4, n_exons_per_tx = 4
)
generate_mxe(gr, n_events = 1)


gr <- create_mock_data(
  n_genes = 2, n_tx_per_gene = 4, n_exons_per_tx = 4
)
generate_ri(gr, n_events = 1)


gr <- create_mock_data(
  n_genes = 2, n_tx_per_gene = 4, n_exons_per_tx = 4
)
generate_a5ss(gr, n_events = 1)


gr <- create_mock_data(
  n_genes = 2, n_tx_per_gene = 4, n_exons_per_tx = 4
)
generate_a3ss(gr, n_events = 1)
```

---

prepare_exons *Prepare exon ranges from a TxDb and DTU results table*

Description

Extracts exon ranges from a TxDb object, merges them with differential transcript usage (DTU) results, and returns a flat GRanges ready for `preprocess`.

Usage

```
prepare_exons(
  txdb,
  dtu_table,
  coef_col,
  tx_id_col = "tx_id",
  gene_id_col = "gene_id",
  verbose = TRUE
)
```

Arguments

txdb	A TxDb object (from GenomicFeatures).
dtu_table	A data.frame or tibble with DTU results. Must contain columns for transcript ID, gene ID, and a coefficient.
coef_col	Column name in dtu_table with the coefficient / effect size values.
tx_id_col	Column name in dtu_table with transcript IDs matching the TxDb transcript names. Default "tx_id".
gene_id_col	Column name in dtu_table with gene IDs. Default "gene_id".
verbose	Whether to print progress messages. Default TRUE.

Value

A GRanges object with metadata columns: gene_id, tx_id, exon_rank, the coefficient column, and any additional columns from dtu_table.

Examples

```
library(AnnotationHub)
library(AnnotationDbi)
library(GenomicFeatures)
library(tibble)

ah <- AnnotationHub()
txdb <- ah[["AH84134"]] # fly TxDb (Drosophila melanogaster)

# build a simulated DTU table from the TxDb transcripts
txps <- txdb |>
  AnnotationDbi::select(
    keys(txdb, "TXID"), c("TXNAME", "GENEID"), "TXID"
  ) |>
  tibble::as_tibble() |>
  dplyr::select(tx_id = TXNAME, gene_id = GENEID)|>
  dplyr::filter(!is.na(gene_id))

sim_dtu_table <- txps |>
  dplyr::mutate(
    padj = runif(dplyr::n()),
    effect_est = rnorm(dplyr::n())
  )

fly_exons <- prepare_exons(
  txdb, sim_dtu_table, coef_col = "effect_est", verbose = TRUE
)
```

prepare_exons_by_partition *Prepare exons from two transcript partitions*

Description

Combines two transcript partitions (up- and down-regulated) and assigns an `estimate` coefficient: `+1` to `up` and `-1` to `down`. Accepts either GRanges objects or character vectors of transcript IDs (in which case `txdb` is required to look up exon coordinates). The result is ready to pass to `preprocess` with `coef_col = "estimate"`.

Usage

```
prepare_exons_by_partition(
  up,
  down,
  txdb = NULL,
  tx_id_col = "TXNAME",
  verbose = TRUE
)
```

Arguments

`up`	A GRanges object or character vector of transcript IDs for the upregulated partition (assigned `estimate = +1`).
`down`	A GRanges object or character vector of transcript IDs for the downregulated partition (assigned `estimate = -1`).
`txdb`	A `TxDb` object (from GenomicFeatures). Required when `up` and `down` are character vectors of transcript IDs.
`tx_id_col`	The keytype in `txdb` matching the transcript IDs in `up` and `down`. Default `"TXNAME"`. Only used when `up` and `down` are character vectors. See `AnnotationDbi::keytypes(txdb)` for available options.
`verbose`	Whether to print progress messages. Default `TRUE`.

Details

When `up` and `down` are GRanges, both must have `exon_rank`, `gene_id`, and `tx_id` metadata columns. Extra columns are kept; if one object lacks a column present in the other, those entries receive `NA`.

Value

A combined GRanges object with an `estimate` column (`+1` for `up`, `-1` for `down`), ready for `preprocess`.

Examples

```
# GRanges input
gr <- create_mock_data(n_genes = 1, n_tx_per_gene = 4, n_exons_per_tx = 4)
gr <- generate_se(gr, n_events = 1)
gr_down <- gr[gr$estimate < 0]
gr_up <- gr[gr$estimate > 0]
prepare_exons_by_partition(gr_up, gr_down) |>
  preprocess(coef_col = "estimate")
```

`preprocess` *Preprocess input GRanges object for splicing event calculation*

Description

This function checks that the input is a valid GRanges object with required metadata columns, then adds a unique key, the number of exons per transcript, and an 'internal' flag for each exon.

Usage

```
preprocess(gr, coef_col, method_string = NULL, additional_columns = NULL)
```

Arguments

`gr`	A GRanges object with metadata columns: 'exon_rank', 'gene_id', 'tx_id', 'coef'.
`coef_col`	The name of the metadata column indicating upregulated (+1) and downregulated (-1) exons.
`method_string`	The Differential Transcript Usage (DTU) method used to obtain the coef_col, for annotation purposes (optional).
`additional_columns`	A character vector of metadata column names to record for downstream use. Stored in `metadata(result)$additional_columns` (optional).

Value

A GRanges object with added 'key', 'nexons', and 'internal' columns.

Examples

```
# create mock data and run preprocessing
gr <- create_mock_data(n_genes = 2, n_tx_per_gene = 4, n_exons_per_tx = 4) |>
  preprocess(coef_col = "estimate", method_string = "mock_method")
```

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