# Package 'tomoda'

November 12, 2026

**Title** Tomo-seq data analysis

**Version** 1.22.0

**Description** This package provides many easy-to-use methods to analyze and visualize tomo-seq data. The tomo-seq technique is based on cryosectioning of tissue and performing RNA-seq on consecutive sections. (Reference: Kruse F, Junker JP, van Oudenaarden A, Bakkers J. Tomo-seq: A method to obtain genome-wide expression data with spatial resolution. Methods Cell Biol. 2016;135:299-307. doi:10.1016/bs.mcb.2016.01.006)
The main purpose of the package is to find zones with similar transcriptional profiles and spatially expressed genes in a tomo-seq sample. Several visulization functions are available to create easy-to-modify plots.

**Depends** R (>= 4.0.0)

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# Contents

corHeatmap . . . . . 2
createTomo . . . . . 3
embedPlot . . . . . 5
expHeatmap . . . . . 6
findPeak . . . . . 7
findPeakGene . . . . . 7
geneCorHeatmap . . . . . 9
geneEmbedPlot . . . . . 10
hierarchClust . . . . . 11
kmeansClust . . . . . 12
linePlot . . . . . 13
normalizeTomo . . . . . 14
runPCA . . . . . 15
runTSNE . . . . . 16
runUMAP . . . . . 17
scaleTomo . . . . . 18
tomoMatrix . . . . . 18
tomoSummarizedExperiment . . . . . 19
zh.data . . . . . 20

**Index** **22**

---

`corHeatmap` *Correlation heatmap of sections*

---

## Description

Heatmap pf correlation coefficients between any two sections in a `SummarizedExperiment` object.

## Usage

```
corHeatmap(object, matrix = "scaled", max.cor = 0.5, cor.method = "pearson")
```

## Arguments

| | |
|---|---|
| `object` | A `SummarizedExperiment` object. |
| `matrix` | Character, must be one of `"count"`, `"normalized"`, or `"scaled"`. |
| `max.cor` | Numeric, correlation coefficients bigger than `max.cor` are set to `max.cor`. It is used to clearly show small correlation coefficients. |
| `cor.method` | Character, the method to calculate correlation coefficients. must be one of `"pearson"`, `"kendall"`, or `"spearman"`. |

## Value

A ggplot object.

## Examples

```
data(zh.data)
zh <- createTomo(zh.data)
corHeatmap(zh)

# Use Spearman correlation coefficients.
corHeatmap(zh, cor.method='spearman')

# Set max correlation coefficients to 0.3.
corHeatmap(zh, max.cor=0.3)
```

---

createTomo *Create an object representing tomo-seq data*

---

## Description

This is a generic function to create an object representing tomo-seq data. The input object can either be a matrix or a SummarizeExperiment.

## Usage

```
createTomo(object, ...)

## S4 method for signature 'SummarizedExperiment'
createTomo(
  object,
  min.section = 3,
  normalize = TRUE,
  normalize.method = "median",
  scale = TRUE
)

## S4 method for signature 'matrix'
createTomo(
  object,
  matrix.normalized = NULL,
  min.section = 3,
  normalize = TRUE,
  normalize.method = "median",
  scale = TRUE
)

## S4 method for signature 'missing'
```

```
createTomo(
  matrix.normalized = NULL,
  min.section = 3,
  normalize = TRUE,
  normalize.method = "median",
  scale = TRUE,
  ...
)
```

## Arguments

| | |
|---|---|
| object | Either a raw read count matrix or a SummarizedExperiment object. |
| ... | Additional parameters to pass to S4 methods. |
| min.section | Integer. Genes expressed in less than min.section sections will be filtered out. |
| normalize | Logical, whether to perform normalization when creating the object. Default is TRUE. |
| normalize.method | Character, must be one of "median", or "cpm". |
| scale | Logical, whether to perform scaling when creating the object. Default is TRUE. |
| matrix.normalized | (Optional) A numeric matrix of normalized read count. |

## Details

This is the generic function to create a SummarizedExperiment object for representing tomo-seq data. Either matrix or SummarizedExperiment object can be used for input.

When using matrix for input, at least one of raw read count matrix and normalized read count matrix (like FPKM and TPM) must be used for input. If normalized matrix is available, input it with argument matrix.normalized. Matrices should have genes as rows and sections as columns. Columns should be sorted according to the order of sections.

When using SummarizedExperiment object for input, it must contain at least one of 'count' assay and 'normalized' assay. Besides, the row data and column data of the input object will be retained in the output object.

By default, all library sizes are normalized to the median library size across sections. Set normalize.method = "cpm" will make library sizes normalized to 1 million counts. Scaling and centering is performed for all genes across sections.

## Value

A SummarizedExperiment object. Raw read count matrix, normalized read count matrix and scaled read count matrix are saved in 'count', 'normalized' and 'scale' assays of the object.

## See Also

- tomoMatrix : creating an object from matrix.
- tomoSummarizedExperiment : creating an object from SummarizedExperiment.

- normalizeTomo : normalization.
- scaleTomo : scaling.
- SummarizedExperiment-class : operations on SummarizedExperiment.

## Examples

```
data(zh.data)
zh <- createTomo(zh.data)

data(zh.data)
se <- SummarizedExperiment::SummarizedExperiment(assays=list(count=zh.data))
zh <- createTomo(se)
```

---

embedPlot *Embedding plot for sections*

---

## Description

Scatter plot for sections with two-dimenstional embeddings in a SummarizedExperiment object. Each point stands for a section.

## Usage

```
embedPlot(object, group = "section", method = "TSNE")
```

## Arguments

| | |
|---|---|
| object | A SummarizedExperiment object. |
| group | Character, a variable in slot meta defining the groups of sections. Sections in the same group have same colors. |
| method | Character, the embeddings for scatter plot. Must be one of "TSNE", "UMAP", or "PCA". |

## Value

A ggplot object.

## Examples

```
data(zh.data)
zh <- createTomo(zh.data)
zh <- runTSNE(zh)
# Plot TSNE embeddings.
embedPlot(zh)

# Plot UMAP embeddings.
zh <- runUMAP(zh)
embedPlot(zh, method="UMAP")
```

```
# Color sections by kmeans cluster labels.
zh <- kmeansClust(zh, 3)
embedPlot(zh, group="kmeans_cluster")
```

---

expHeatmap *Expression heatmap*

---

## Description

Heatmap for gene expression across sections in a SummarizedExperiment object.

## Usage

```
expHeatmap(object, genes, matrix = "scaled", size = 5)
```

## Arguments

| | |
|---|---|
| object | A SummarizedExperiment object. |
| genes | A vector of character, the name of genes to plot heatmap. |
| matrix | Character, must be one of "count", "normalized", or "scaled". |
| size | Character, the size of gene names. Set it to 0 if you do not want to show gene names. |

## Value

A ggplot object.

## Examples

```
data(zh.data)
zh <- createTomo(zh.data)

# Plot some genes.
expHeatmap(zh,
 c("ENSDARG00000002131", "ENSDARG00000003061", "ENSDARG00000076075", "ENSDARG00000076850"))

# Plot peak genes.
peak_genes <- findPeakGene(zh)
expHeatmap(zh, peak_genes$gene)

# Remove gene names if too many genes are in the heatmap.
expHeatmap(zh, peak_genes$gene, size=0)
```

## findPeak *Find peak in a vector*

### Description

Find the position of peak in a vector.

### Usage

```
findPeak(x, threshold = 1, length = 4)
```

### Arguments

| | |
|---|---|
| x | A numeric vector. |
| threshold | Integer, only values bigger than threshold are recognized as part of the peak. |
| length | Integer, minimum length of consecutive values bigger than threshold are recognized as a peak. |

### Value

A numeric vector. The first element is the start index and the second element is the end index of the peak. If multiple peaks exist, only output the start and end index of the one with maximun length. If no peak exist, return c(0, 0).

### Examples

```
# return c(3, 10)
findPeak(c(0:5, 5:0), threshold=1, length=4)

# Most likely return c(0, 0)
findPeak(rnorm(10), threshold=3, length=3)
```

## findPeakGene *Find peak genes*

### Description

Find peak genes (spatially upregulated genes) in a SummarizedExperiment object.

## Usage

```
findPeakGene(
  object,
  threshold = 1,
  length = 4,
  matrix = "scaled",
  nperm = 1e+05,
  method = "BH"
)
```

## Arguments

| | |
|---|---|
| object | A SummarizedExperiment object. |
| threshold | Integer, only scaled read counts bigger than threshold are recognized as part of the peak. |
| length | Integer, scaled read counts bigger than threshold in minimum length of consecutive sections are recognized as a peak. |
| matrix | Character, must be one of "count", "normalized", or "scaled". |
| nperm | Integer, number of random permutations to calculate p values. Set it to 0 if you are not interested in p values. |
| method | Character, the method to adjust p values for multiple comparisons, must be one of "holm", "hochberg", "hommel", "bonferroni", "BH", "BY", "fdr", "none". |

## Details

Peak genes are selected based on scaled read counts. As scaled read counts are Z scores, suggested threshold are $[1, 3]$. Smaller threshold and length makes the function detect more peak genes, and vice versa. P values are calculated by approximate permutation tests. For a given threshold and length, the scaled read counts of each gene is randomly permutated for nperm times. The p value is defined as the ratio of permutations containing peaks. In order to speed up permutation process, genes whose expression exceeds threshold in same number of sections have same p values. To be specific, only one of these genes will be used to calculate a p value by permutation, and other genes are assigned this p value.

## Value

A data.frame with peak genes as rows. It has following columns:

- gene : Character, peak gene names.
- start : Numeric, the start index of peak.
- end : Numeric, the end index of peak.
- center : Numeric, the middle index of peak. If the length of the peak is even, center is defined as the left-middle index.
- p : Numeric, p values.
- p.adj : Numeric, adjusted p values.

## Examples

```
data(zh.data)
zh <- createTomo(zh.data)
peak_genes <- findPeakGene(zh)
head(peak_genes)

# Increase threshold so that less peak genes will be found.
peak_genes <- findPeakGene(zh, threshold=1.5)

# Increase peak length so that less peak genes will be found.
peak_genes <- findPeakGene(zh, length=5)

# Set nperm to 0 so that p values will not be calculated. This will save running time.
peak_genes <- findPeakGene(zh, nperm=0)
```

---

# geneCorHeatmap *Correlation heatmap of genes*

---

## Description

Heatmap of correlation coefficients between any two queried genes in a `SummarizedExperiment` object.

## Usage

```
geneCorHeatmap(
  object,
  gene.df,
  group = "center",
  matrix = "scaled",
  size = 5,
  cor.method = "pearson"
)
```

## Arguments

| | |
|---|---|
| `object` | A `SummarizedExperiment` object. |
| `gene.df` | Data.frame. The first column must be a vector of gene names, and has the name `"gene"`. Additional columns in `gene.df` can be used to set the colors of genes. |
| `group` | Character, a column name in `gene.df` defining the groups of genes. Genes in the same group have same colors on the side bar. |
| `matrix` | Character, must be one of `"count"`, `"normalized"`, or `"scaled"`. |
| `size` | Numeric, the size of gene names. Set it to 0 if you do not want to show gene names. |
| `cor.method` | Character, the method to calculate correlation coefficients. must be one of `"pearson"`, `"kendall"`, or `"spearman"`. |

## Details

This method can create a pure heatmap or a heatmap with side bar. If you prefer a pure heatmap, input a `gene.df` with a single column of gene names. However, you may want to show additional information of genes with a side bar, and the grouping information should be saved as additional column(s) of `gene.df`, and declared as `group`. By default, you can use the output by `findPeakGene` as input `gene.df`. Peak genes will be grouped by their centers on the side bar.

## Value

A `ggplot` object.

## Examples

```
data(zh.data)
zh <- createTomo(zh.data)

# Correlation heatmap for all peak genes.
peak_genes <- findPeakGene(zh)
geneCorHeatmap(zh, peak_genes)

# Use Spearman correlation coefficients.
geneCorHeatmap(zh, peak_genes, cor.method="spearman")

# Group genes by peak start.
geneCorHeatmap(zh, peak_genes, group="start")

# Plot without side bar.
geneCorHeatmap(zh, data.frame(
 gene=c("ENSDARG00000002131", "ENSDARG00000003061", "ENSDARG00000076075", "ENSDARG00000076850")))
```

---

# geneEmbedPlot *Embedding plot for genes*

---

## Description

Scatter plot for genes with two-dimenstional embeddings in a `SummarizedExperiment` object. Each point stands for a gene.

## Usage

```
geneEmbedPlot(object, gene.df, group = "center", method = "TSNE")
```

## Arguments

| | |
|---|---|
| `object` | A `SummarizedExperiment` object. |
| `gene.df` | Data.frame. The first column must be a vector of gene names, and has the name `"gene"`. Additional columns in `gene.df` can be used to set the colors of genes. |

| | |
|---|---|
| group | Character, a column name in gene.df defining the groups of genes. Genes in the same group have same colors. |
| method | Character, the embeddings for scatter plot. Must be one of "TSNE", "UMAP", or "PCA". |

## Value

A ggplot object.

## Examples

```
data(zh.data)
zh <- createTomo(zh.data)
peak_genes <- findPeakGene(zh)
zh <- runTSNE(zh, peak_genes$gene)
# Color genes by peak centers.
geneEmbedPlot(zh, peak_genes)

# Color genes by peak starts.
geneEmbedPlot(zh, peak_genes, group="start")

# Do not color genes.
geneEmbedPlot(zh, peak_genes["gene"])
```

---

# hierarchClust *Hierarchical clustering across sections*

---

## Description

Performs hierarchical clustering across sections in a SummarizedExperiment object.

## Usage

```
hierarchClust(
  object,
  matrix = "normalized",
  measure = "euclidean",
  p = 2,
  agglomeration = "complete"
)
```

## Arguments

| | |
|---|---|
| object | A SummarizedExperiment object. |
| matrix | Character, must be one of "count", "normalized", or "scaled". |
| measure | Character, must be one of "euclidean", "maximum", "manhattan", "canberra", "binary" or "minkowski". |

| | |
|---|---|
| p | Numeric, the power of the Minkowski distance. |
| agglomeration | Character, must be one of "ward.D", "ward.D2", "single", "complete", "average", "mcquitty", "median" or "centroid". |

## Value

A hclust object.

## See Also

dist for measuring distance and hclust for performing hierarchical clustering on a matrix.

## Examples

```
data(zh.data)
zh <- createTomo(zh.data)
hclust_zh <- hierarchClust(zh)
plot(hclust_zh)

# Use other agglomeration method
hclust_zh <- hierarchClust(zh, agglomeration="average")

# (Not recommended) Use scaled read counts to calculate distance
zh <- scaleTomo(zh)
hclust_zh <- hierarchClust(zh, matrix="scaled")
```

---

# kmeansClust *K-Means clustering across sections*

---

## Description

Performs K-Means clustering across sections in a SummarizedExperiment object.

## Usage

```
kmeansClust(object, centers, matrix = "normalized", ...)
```

## Arguments

| | |
|---|---|
| object | A SummarizedExperiment object. |
| centers | Integer, number of clusters, namely *k*. |
| matrix | Character, must be one of "count", "normalized", or "scaled". |
| ... | other parameters passed to kmeans. |

## Value

A SummarizedExperiment object. The obtained cluster labels are saved in slot meta.

## See Also

kmeans for performing K-Means clustering on a matrix.

## Examples

```
data(zh.data)
zh <- createTomo(zh.data)
zh <- kmeansClust(zh, 3)

# Use scaled read counts to calculate distance
zh <- scaleTomo(zh)
zh <- kmeansClust(zh, 3, matrix="scaled")
```

---

# linePlot *Line plot for expression traces*

---

## Description

Plot expression traces for genes across sections in a SummarizedExperiment object.

## Usage

```
linePlot(object, genes, matrix = "normalized", facet = FALSE, span = 0.3)
```

## Arguments

| | |
|---|---|
| object | A SummarizedExperiment object. |
| genes | A character vector of gene names for plotting expression traces. |
| matrix | Character, must be one of "count", "normalized", or "scaled". |
| facet | Logical. Plot the expression trace of each gene in a facet if it is TRUE. |
| span | Numeric, the amount of smoothing for the default loess smoother. Smaller numbers produce wigglier lines, larger numbers produce smoother lines. Set it to 0 for non-smoothing lines. |

## Value

A ggplot object.

## See Also

geom_smooth for plotting smooth lines, facet_wrap for faceting genes.

## Examples

```
data(zh.data)
zh <- createTomo(zh.data)
linePlot(zh,
 c("ENSDARG00000002131", "ENSDARG00000003061", "ENSDARG00000076075", "ENSDARG00000076850"))

# Do not smooth lines.
linePlot(zh,
 c("ENSDARG00000002131", "ENSDARG00000003061", "ENSDARG00000076075", "ENSDARG00000076850"), span=0)

# Plot genes in different facets.
linePlot(zh,
 c("ENSDARG00000002131", "ENSDARG00000003061", "ENSDARG00000076075", "ENSDARG00000076850"),
 facet=TRUE)
```

---

`normalizeTomo` *Normalize data*

---

## Description

Normalize the raw read count in a `SummarizedExperiment` object.

## Usage

```
normalizeTomo(object, method = "median")
```

## Arguments

| | |
|---|---|
| `object` | A `SummarizedExperiment` object. |
| `method` | Character, must be one of `"median"`, or `"cpm"`. |

## Details

This function should be run for `SummarizedExperiment` object created from raw read count matrix. If the `SummarizedExperiment` object already has a normalized count matrix. The function simply return the original object. Library sizes of all sections are normalized to the median library size (method='median') or one million (method='cpm').

## Value

A `SummarizedExperiment` object with normalized read count matrix saved in assay `'normalized'`.

## Examples

```
data(zh.data)
zh <- createTomo(zh.data, normalize=FALSE)
zh <- normalizeTomo(zh)
```

runPCA *Perform PCA*

## Description

Perform PCA on sections or genes in a SummarizedExperiment object for dimensionality reduction.

## Usage

```
runPCA(object, genes = NA, matrix = "auto", scree = FALSE, ...)
```

## Arguments

| | |
|---|---|
| object | A SummarizedExperiment object. |
| genes | NA or a vector of character. Perform PCA on sections if it is NA, or on given genes if it is a vector of gene names. |
| matrix | Character, must be one of "auto", "count", "normalized", or "scaled". If "auto", normalized matrix is used for sections and scaled matrix is used for genes. |
| scree | Logical, plot the scree plot for PCs if it is TRUE. |
| ... | Other parameters passed to prcomp. |

## Value

A SummarizedExperiment object. The PC embeddings are saved in slot meta if PCA is performed on sections, or saved in slot gene_embedding if PCA is performed on genes.

## See Also

prcomp for performing PCA on a matrix.

## Examples

```
data(zh.data)
zh <- createTomo(zh.data)

# Perform PCA on sections.
zh <- runPCA(zh)

# Plot the scree plot.
zh <- runPCA(zh, scree=TRUE)

# Perform PCA on some genes.
zh <- runPCA(zh, genes=rownames(zh)[1:100])
```

runTSNE *Perform TSNE*

## Description

Perform TSNE on sections or genes in a SummarizedExperiment object for dimensionality reduction.

## Usage

```
runTSNE(object, genes = NA, matrix = "auto", perplexity = NA, ...)
```

## Arguments

| | |
|---|---|
| object | A SummarizedExperiment object. |
| genes | NA or a vector of character. Perform TSNE on sections if it is NA, or on given genes if it is a vector of gene names. |
| matrix | Character, must be one of "auto", "count", "normalized", or "scaled". If "auto", normalized matrix is used for sections and scaled matrix is used for genes. |
| perplexity | Numeric, perplexity parameter for Rtsne (default: 0.25 *(number of observations - 1)). |
| ... | Other parameters passed to Rtsne. |

## Value

A SummarizedExperiment object. The TSNE embeddings are saved in slot meta if TSNE is performed on sections, or saved in slot gene_embedding if TSNE is performed on genes.

## See Also

Rtsne for performing TSNE on a matrix.

## Examples

```
data(zh.data)
zh <- createTomo(zh.data)

# Perform TSNE on sections.
zh <- runTSNE(zh)

# Perform TSNE on sections with other perplexity.
zh <- runTSNE(zh, perplexity=10)

# Perform TSNE on some genes.
zh <- runTSNE(zh, genes=rownames(zh)[1:100])
```

runUMAP *Perform UMAP*

## Description

Perform UMAP on sections or genes in a SummarizedExperiment object for dimensionality reduction.

## Usage

```
runUMAP(object, genes = NA, matrix = "auto", ...)
```

## Arguments

| | |
|---|---|
| object | A SummarizedExperiment object. |
| genes | NA or a vector of character. Perform UMAP on sections if it is NA, or on given genes if it is a vector of gene names. |
| matrix | Character, must be one of "auto", "count", "normalized", or "scaled". If "auto", normalized matrix is used for sections and scaled matrix is used for genes. |
| ... | Other parameters passed to umap. |

## Value

A SummarizedExperiment object. The UMAP embeddings are saved in slot meta if UMAP is performed on sections, or saved in slot gene_embedding if UMAP is performed on genes.

## See Also

umap for performing UMAP on a matrix.

## Examples

```
data(zh.data)
zh <- createTomo(zh.data)

# Perform UMAP on sections.
zh <- runUMAP(zh)

# Perform UMAP on some genes.
zh <- runUMAP(zh, genes=rownames(zh)[1:100])
```

## scaleTomo *Scale data*

### Description

Scale the normalized read count in a SummarizedExperiment object.

### Usage

```
scaleTomo(object)
```

### Arguments

| | |
|---|---|
| object | A SummarizedExperiment object. |

### Details

This function should be run for SummarizedExperiment object with normalized read count matrix. The normalized read counts of each gene are subjected to Z score transformation across sections.

### Value

A SummarizedExperiment object with scaled read count matrix saved in assay 'scaled'.

### Examples

```
data(zh.data)
zh <- createTomo(zh.data, scale=FALSE)
zh <- scaleTomo(zh)
```

## tomoMatrix *Create an object from matrix*

### Description

tomoMatrix creates an object from raw read count matrix or normalized read count matrix.

### Usage

```
tomoMatrix(
  matrix.count = NULL,
  matrix.normalized = NULL,
  min.section = 3,
  normalize = TRUE,
  normalize.method = "median",
  scale = TRUE
)
```

### Arguments

| | |
|---|---|
| matrix.count | A numeric matrix or matrix-like data stucture that can be coverted to matrix, with genes with rows, sections as columns and values as raw read counts. Columns should be sorted according to section numbers. |
| matrix.normalized | A numeric matrix or matrix-like data stucture that can be coverted to matrix, with genes as rows, sections as columns and values as normalized read counts. Columns should be sorted according to order of sections. |
| min.section | Integer. Genes expressed in less than min.section sections will be filtered out. |
| normalize | Logical, whether to perform normalization when creating the object. Default is TRUE. |
| normalize.method | Character, must be one of "median", or "cpm". |
| scale | Logical, whether to perform scaling when creating the object. Default is TRUE. |

### Value

A SummarizedExperiment object

### See Also

createTomo for the generic function.

### Examples

```
data(zh.data)
zh <- tomoMatrix(zh.data)
```

---

## tomoSummarizedExperiment

*Create an object from SummarizedExperiment*

---

### Description

tomoSummarizedExperiment creates an object from a SummarizedExperiment object.

### Usage

```
tomoSummarizedExperiment(
  se,
  min.section = 3,
  normalize = TRUE,
  normalize.method = "median",
  scale = TRUE
)
```

### Arguments

| | |
|---|---|
| se | A SummarizedExperiment object, it must contain at least one of 'count' assay and 'normalized' assay. |
| min.section | Integer. Genes expressed in less than min.section sections will be filtered out. |
| normalize | Logical, whether to perform normalization when creating the object. Default is TRUE. |
| normalize.method | Character, must be one of "median", or "cpm". |
| scale | Logical, whether to perform scaling when creating the object. Default is TRUE. |

### Value

A SummarizedExperiment object

### See Also

createTomo for the generic function.

### Examples

```
data(zh.data)
se <- SummarizedExperiment::SummarizedExperiment(assays=list(count=zh.data))
zh <- tomoSummarizedExperiment(se)
```

---

## zh.data *A raw read count matrix of zebrafish injured heart.*

---

### Description

A dataset containing gene expression across 40 sections of **z**ebrafish **h**eart generated with Tomo-seq. The zebrafish heart is 3 days post cryoinjury (3 dpi).

### Usage

```
data(zh.data)
```

### Format

A numeric matrix with 16495 genes as rows and 40 section as columns. Its row names are gene names and column names are section names.

### Source

https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE74652

## References

Wu CC, Kruse F, Vasudevarao MD, et al. Spatially Resolved Genome-wide Transcriptional Profiling Identifies BMP Signaling as Essential Regulator of Zebrafish Cardiomyocyte Regeneration. *Dev Cell*. 2016;36(1):36-49. doi:10.1016/j.devcel.2015.12.010

# Index

∗ **datasets**
zh.data, 20

corHeatmap, 2
createTomo, 3, *19, 20*
createTomo,matrix-method *(*createTomo*)*, 3
createTomo,missing-method *(*createTomo*)*, 3
createTomo,SummarizedExperiment-method *(*createTomo*)*, 3

dist, *12*

embedPlot, 5
expHeatmap, 6

findPeak, 7
findPeakGene, 7

geneCorHeatmap, 9
geneEmbedPlot, 10

hclust, *12*
hierarchClust, 11

kmeans, *13*
kmeansClust, 12

linePlot, 13

normalizeTomo, *5*, 14

prcomp, *15*

Rtsne, *16*
runPCA, 15
runTSNE, 16
runUMAP, 17

scaleTomo, *5*, 18

tomoMatrix, *4*, 18
tomoSummarizedExperiment, *4*, 19

umap, *17*

zh.data, 20